

# Towards single-molecule nanomechanical mass spectrometry

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**Mass spectrometry provides rapid and quantitative identification of protein species with relatively low sample consumption. The trend towards biological analysis at increasingly smaller scales, ultimately down to the volume of an individual cell, continues, and mass spectrometry with a sensitivity of a few to single molecules will be necessary. Nanoelectromechanical systems provide unparalleled mass sensitivity, which is now sufficient for the detection of individual molecular species in real time. Here, we report the first demonstration of mass spectrometry based on single biological molecule detection with a nanoelectromechanical system. In our nanoelectromechanical-mass spectrometry system, nanoparticles and protein species are introduced by electrospray injection from the fluid phase in ambient conditions into vacuum, and are subsequently delivered to the nanoelectromechanical system detector by hexapole ion optics. Precipitous frequency shifts, proportional to the mass, are recorded in real time as analytes adsorb, one by one, onto a phase-locked, ultrahigh-frequency nanoelectromechanical resonator. These first nanoelectromechanical system-mass spectrometry spectra, obtained with modest mass sensitivity from only several hundred mass adsorption events, presage the future capabilities of this approach. We also outline the substantial improvements that are feasible in the near term, some of which are unique to nanoelectromechanical system based-mass spectrometry.**

Nanoelectromechanical systems (NEMS)<sup>1</sup> are enabling important emerging applications in diverse fields ranging from quantum measurement to biotechnology<sup>2–9</sup>. In general, the smaller a device, the more susceptible are its physical properties to perturbation by external influences. This enhanced sensitivity of NEMS is opening a variety of unprecedented opportunities for applications such as mass spectrometry, which is now widely used for proteomics<sup>10,11</sup>. Furthermore, to reliably detect the expression of low-level signals and to understand the fundamental biological processes, it is important to develop techniques capable of single-cell or single-molecule analyses<sup>12,13</sup>. In this work, the exceptional mass sensitivity of ultrahigh-frequency (UHF) NEMS resonators<sup>14–18</sup>—derived from their minuscule masses, high frequencies, and high resonance quality factors—is used to demonstrate a new paradigm for mass spectrometry. Our approach enables the first real-time detection of individual protein molecules and nanoparticles as they adsorb on a sensitive NEMS detector. We use these to carry out an initial form of mass spectrometry based on discrete adsorption events.

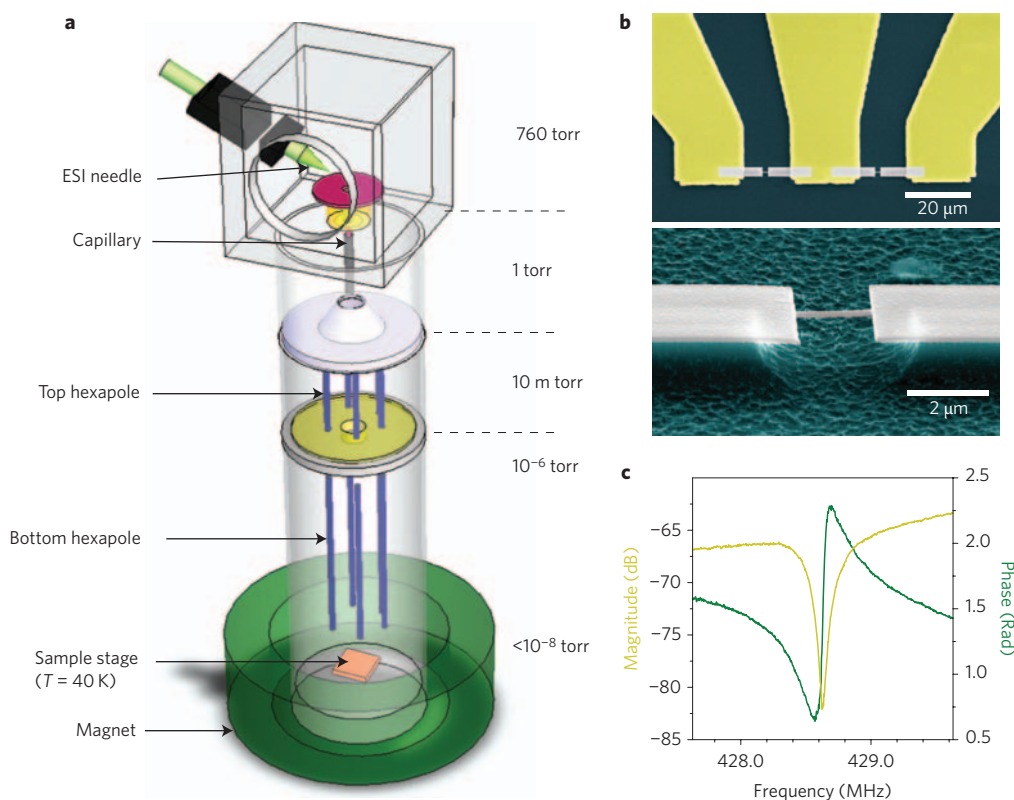
The vibrational frequency of a NEMS resonator is an exquisitely sensitive function of its total mass. Small variations in mass, for example, from adsorbed addenda, can measurably alter its resonant frequency. Theoretical calculations for physically realizable devices indicate that NEMS mass sensitivity below a single dalton (1 Da = 1 AMU) is achievable<sup>19,20</sup>. Experimental measurements of NEMS mass sensing at the ~1,000 Da level<sup>17,21</sup> and, more recently, below the 200 Da level<sup>22,23</sup> have been demonstrated. Our NEMS-MS (mass spectrometry) paradigm is also quite distinct from existing approaches to mass spectrometry in that the inertial mass of each arriving species—atom, molecule or nanoparticle—is ‘weighed’ as the analyte adsorbs upon the detector. Hence, a mass analyser is not needed to pre-separate and aggregate similar species. In fact, it is possible to contemplate circumventing analyte ionization entirely if alternative injection and transport methods for neutral species are used. This may offer significant advantages for whole-protein MS of

high-mass species by circumventing electrostatic fragmentation. It should also dramatically reduce analyte consumption by permitting the mass detector to be positioned in close proximity to the protein source. The singular advantage of NEMS-MS is that each NEMS sensor in the single-molecule limit acts as an individual mass spectrometer. This NEMS-based mass spectrometry system, combined with other micro- and nanoscale technology<sup>24,25</sup>, offers the possibility of compact, massively parallel MS, limited only by the number of NEMS mass sensors incorporated on a chip.

## NEMS-based mass spectrometry

Typically, mass spectrometers comprise three separate components that provide the critical functions of operation: analyte ionization, analyte separation and detection. Analyte species in the fluid phase are first ionized, and bare (unsolvated) ions are produced using electrospray ionization (ESI)<sup>26,27</sup>. Second, ion separation is undertaken in vacuum based on the charge-to-mass ( $m/z$ ) ratio of the analytes. Third, detection of clustered groups of these analytes with similar  $m/z$  values is carried out to determine the presence of a given species. Our new paradigm of NEMS-MS combines the latter two functions into one: the NEMS sensor is used as both mass analyser and mass detector. This NEMS mass analyser/detector, in this first realization described here, is preceded by well-validated mass spectrometry components for analyte injection and delivery. Figure 1 schematically depicts our prototype experimental system, which introduces, transports and measures the masses of analytes. Protein ions or charged nanoparticles are produced and stripped of fluidic solvent in the course of ESI. These bare ions traverse through a three-stage differentially pumped vacuum system and land onto the NEMS mass analyser/detector situated ~2 m away from the ESI source. Two stages of hexapole ion optics<sup>28</sup> driven at radio frequency (an RF-only hexapole) are used to guide the species to the NEMS with minimal  $m/z$  discrimination, as desired (see Supplementary Information). As the individual protein molecules and nanoparticles arrive and accrete onto the NEMS

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**Figure 1 | First-generation NEMS-MS system.** **a**, Simplified schematic of the experimental configuration (not to scale), showing the fluid-phase electrospray ionization (ESI) and injection, the system's three-stage differential pumping and its two-stage ion optics. **b**, Scanning electron micrographs showing one of the doubly clamped beam NEMS devices used in these experiments. It is embedded in a nanofabricated three-terminal UHF bridge circuit. **c**, Magnitude and phase of the UHF NEMS resonator's response displaying a prominent fundamental-mode resonance near 428 MHz.

sensor, its resonant frequency jumps downward abruptly with each individual molecular or nanoparticle adsorption event (Fig. 2). These precipitous events, which are absent during the control runs (see Supplementary information), are the hallmark of NEMS mass sensing with single-molecule (-nanoparticle) sensitivity. They are observed here directly for the first time, and are the centrepiece of this work.

Each quasi-instantaneous frequency jump provides information about the specific atom, molecule or nanoparticle that has just adsorbed onto the sensor. The jump height for each event—that is, the resonator's adsorbate-induced frequency deviation—depends upon both the mass of the arriving analyte and its position of adsorption on the NEMS resonator. This response is characterized by a position-dependent mass responsivity for the NEMS resonator (see Supplementary information).

### Single-molecule event analysis

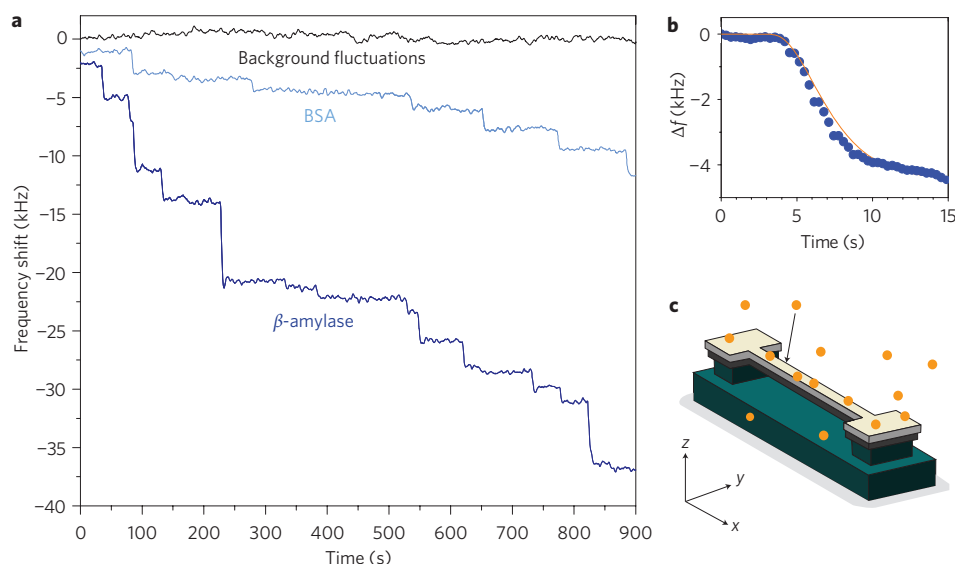
There are two ways through which we can extract the adsorbate mass from the convolved mass- and position-dependence of these adsorption-induced jumps. The more difficult to achieve, but conceptually most transparent, is to orchestrate simultaneous measurement of both jump height and landing position for each species as it arrives, in real time. We shall return to discuss this option below. For our first proof-of-principle demonstration reported here, we take a second, simpler approach—that of building histograms of event probability versus frequency-shift amplitude for small ensembles of sequential single-molecule or single-nanoparticle adsorption events. Figure 3a shows event probabilities calculated for sequential adsorption events originating from a monodisperse source of nanoparticles and their subsequent measurement by a NEMS detector. We assume the resonant mass detector to be a doubly clamped beam operating in fundamental mode, as in our experiments, and

we scale the results to correspond both to our experimentally established NEMS mass responsivity and the masses of the gold nanoparticles we have used, which have a nominal  $\sim 2.5$  nm radius. Figure 3a shows both the ideal case (zero nanoparticle size dispersion and perfect mass sensitivity) as well as more realistic experimental situations that include both the effects of finite size dispersion and the detector's frequency-fluctuation noise. Together, these latter effects reduce the resolution available in a practical system. It is readily apparent from Fig. 3 that the 'canonical' event-probability response is bicuspid, with cusps near zero and at a specific, maximum frequency shift that is associated with adsorption events at the central, most sensitive region of the beam vibrating in fundamental mode. Below, we shall use this characteristic fundamental-mode event-probability shape function to achieve mass analysis in these first NEMS-MS efforts.

### NEMS-MS spectra

We report here the first NEMS-MS spectra for proteins and nanoparticles. Our analyses are carried out by observing, for each species introduced by electrospray, several hundred discrete and abrupt frequency shifts in real time—each associated with an individual protein or nanoparticle adsorption event. Each ESI run is followed by a control run of equal duration, during which the ESI solution pump is shut off to establish the frequency-fluctuation background (see Supplementary information). Given the  $\sim 250$  Hz resolution in these measurements, we construct event probability histograms with 250 Hz bins and reject false positives arising from frequency-fluctuation noise by discarding jumps smaller than  $2\sigma$  ( $= 500$  Hz).

Figure 3b shows an experimental histogram constructed from data obtained by electrospraying a colloidal solution of gold nanoparticles (see Supplementary information). The gold nanoparticles



**Figure 2 | Real-time records of single-molecule adsorption events on a NEMS mass sensor.** **a**, These raw experimental data show the distinctly different, precipitous resonance frequency shifts of the NEMS during ESI-induced adsorption of BSA (66 kDa) and  $\beta$ -amylase (200 kDa). Each frequency jump downward is due to an individual protein adsorption event on the NEMS mass sensor. The height of each frequency jump is a convolved function of the mass of the protein that has adsorbed, and its position of adsorption upon the NEMS. **b**, Raw data from a typical discrete event (blue dots), and a nonlinear least-squares fit to the system's response (orange line), based on the temporal response function of the control loop. **c**, Schematic illustrating single-molecule adsorption events on a NEMS resonator, and the coordinate system used to define its position-dependent mass responsivity. The device itself comprises silicon carbide (dark grey) with a metallic layer (light grey) on top. The silicon substrate (green) beneath the silicon carbide is etched to release (suspend) the doubly clamped beam. The molecules are shown in orange.

used are characterized by the vendor as having an average radius of 2.5 nm and standard deviation of 'less than' 0.375 nm (Sigma-Aldrich). What is ostensibly a small standard deviation in radius actually corresponds to a rather large spread in nanoparticle mass; the advertised specifications translate to an average mass of  $\sim 780$  kDa and  $1\sigma$  mass range of 480–1,190 kDa. This substantial dispersion and our finite frequency-fluctuation noise together result in significant smoothing of the canonical bicuspid spectrum expected for the ideal case (Fig. 3a).

The inset to Fig. 3b shows the residues for a two-parameter least-squares fit of a theoretical event-probability curve to the experimental histogram data. The two fitting parameters are average nanoparticle radius and its dispersion; the theoretical curve incorporates the experimentally measured frequency shift resolution of  $\delta f \approx 250$  Hz. With this approach, by recording just 544 individual nanoparticle adsorption events, we resolve an average nanoparticle radius of 2.15 nm, corresponding to an average mass of 490 kDa, with a standard deviation of nanoparticle radius of 0.5 nm.

Figure 4 shows NEMS-MS spectra obtained for a 'nominally pure' solution of the protein bovine serum albumin (BSA, 66 kDa) (see Supplementary information). From the standpoint of MS, pure solutions of protein are the exception rather than the rule<sup>29,30</sup>, and the NEMS-MS spectra of Fig. 4 bear out this truism. Protein molecules often aggregate in solution to form oligomers (see Supplementary information), and each distinct molecular assemblage present in the sample will produce its own characteristic bicuspid NEMS-MS histogram. A multicomponent solution of such oligomers will thus superpose to produce a complex spectrum.

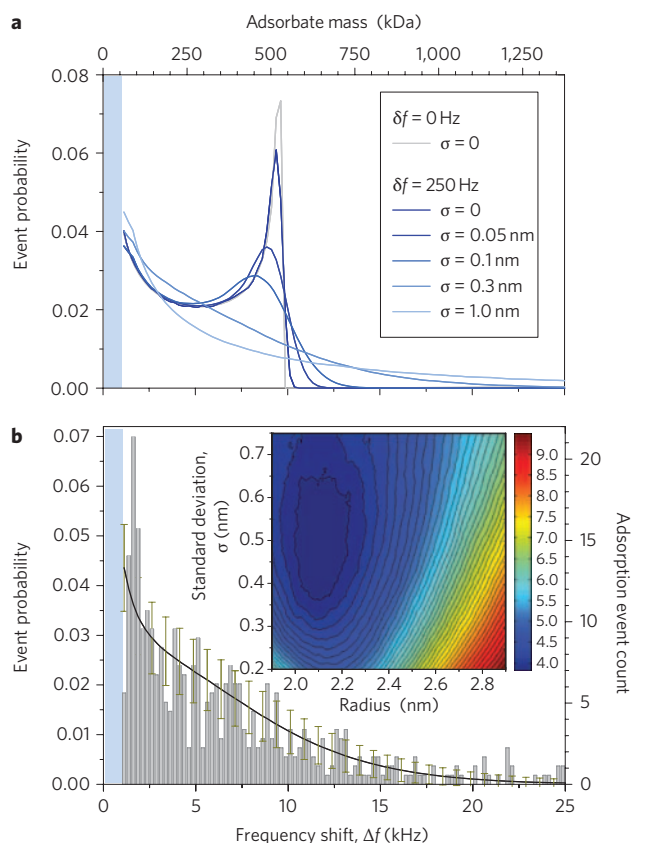
The presence of a family of oligomers has two significant effects on the shape of NEMS-MS histograms. First, the low-frequency shift cusps for each of the oligomers (occurring at the same, zero, frequency shift) superpose to produce a single, prominent peak. Second, the high-frequency shift cusps of the oligomers, which occur at different frequency shifts corresponding to each specific component's mass, become engulfed in the tails from other components. This tends to suppress their overall individual prominence.

Figure 4 illustrates event probabilities as a function of frequency shift for electrosprayed BSA ions that are transported to the NEMS sensor with a hexapole drive frequency of 1.1 MHz. For comparison, also shown are theoretically expected event probabilities for BSA oligomers, generated using the experimentally measured NEMS sensor's mass responsivity of  $\sim 12$  Hz  $zg^{-1}$  and using a least-squares fit to the data—similar to that used for the gold nanoparticle dispersion, but here implemented to extract the spectral weights for the first five oligomers (see Supplementary information). The spectrum in Fig. 4 shows clear peaks at 3,375 and 5,875 Hz, assigned to BSA trimers and pentamers, respectively. The broadly distributed spectral weight below 2 kHz arises from monomers and dimers. Note that this full spectrum has been obtained by recording the individual adsorption of only 578 BSA molecules.

### Prospects for NEMS-MS

We believe these initial results demonstrate the potential of NEMS for MS and provide an unequivocal proof-of-principle for real-time detection of individual proteins and nanoparticles. The full capabilities and sensitivity of the new NEMS-MS paradigm will unfold in a second-generation realization providing both mass- and position-sensing in real time, for each analyte molecule as it arrives. This approach will completely obviate the need for the histogram-based analyses used in the first demonstrations reported here.

The procedure for simultaneous mass and position sensing exists<sup>31,32</sup>, and has been experimentally proven at the microscale<sup>31</sup>. In automated, real-time form, it involves the simultaneous excitation, frequency-locking and tracking of multiple vibrational modes of the resonant NEMS mass sensor. As each analyte adsorbs onto the sensor, it induces a distinct frequency shift for each of the modes monitored. The combined information from the time-correlated shifts from just two modes provides sufficient information to deconvolve the adsorbate's mass and position for each event as it occurs. Tracking additional modes over-determines the solution, providing reduced variance in the deduced values of

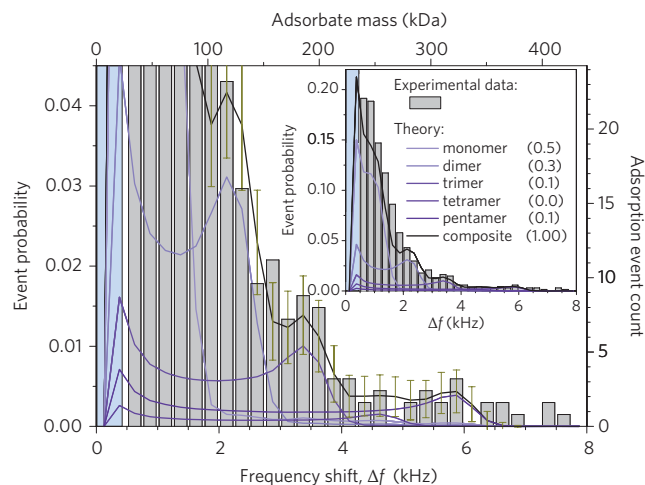


**Figure 3 | NEMS mass spectrometry of a gold nanoparticle dispersion. a,** Theoretically expected event probabilities versus frequency-jump amplitudes are shown for ‘nominal’ 2.5-nm-radius gold nanoparticles (modelled assuming a 2.15 nm mean radius), delivered with an average flux that is uniformly distributed over a doubly clamped beam having peak mass responsivity of  $\sim 12$  Hz  $\text{zg}^{-1}$ . The traces show the expected results for a monodisperse ensemble of particles for the cases of perfect (0 Hz) and experimentally relevant (250 Hz) frequency resolutions, and also for several dispersions (characterized by their radius standard deviations) at 250 Hz. **b,** Experimentally obtained histogram of adsorption event probabilities versus frequency-jump amplitude for electrospayed gold nanoparticles, and the expected curve for an average radius of 2.15 nm and a radial dispersion of 0.375 nm (black trace). The error bars display the theoretically expected deviations corresponding to 544 adsorption events, as registered in this experiment. Inset: contour plot showing the residues for least-squares fits to the experimental data using radius and radial dispersion as the fitting parameter. These data establish the average radius and size dispersion for the 544 nanoparticles measured.

particle mass and position of adsorption. This added technological component will permit the mass of each arriving molecule to be quantitatively measured in real time as it adsorbs upon the NEMS, down to the sensitivity limit imposed by the mass noise floor.

We outline below the anticipated capabilities of NEMS-MS vis-à-vis other current techniques.

**Mass resolution.** Mass resolution in current implementations of MS is typically defined as the ratio of the mass/charge ( $m/z$ ) value and the width of the mass peaks at half maxima. The best mass resolution is obtained with Fourier transform mass spectrometry (FTMS) systems, which attain mass resolutions of the order of 2,000,000, with typical measurement times of about a second. The resolution in FTMS is high at low  $m/z$ , but deteriorates with higher  $m/z$ . Unlike the current MS systems, which measure the



**Figure 4 | NEMS-MS of proteins.** NEMS-MS of BSA enabled by adsorption-event probability analysis. Experimentally obtained frequency-jump data are binned into 250 Hz histograms commensurate with the experimental mass sensitivity. Applying a  $2\sigma$  detection criterion, we reject data below 500 Hz (blue-shaded regions; see text). The main plot shows an expanded view of the low-event-probability region, displaying a clearly detailed decomposition of the simultaneous contributions from oligomers. The theoretical composite curve (grey) is a weighted superposition of adsorption probabilities of the intact monomer and a family of its oligomers deterministically calculated by a least-squares process similar to that of Fig. 3 (see also Supplementary information). The inset shows the entire dataset for the 578 BSA molecular adsorption events recorded in this experiment. The numerically determined best-fit weighting coefficients for the composite curve are displayed.

mass-to-charge ratio, NEMS devices measure the mass of the molecule directly. Achieving a mass sensitivity of 1 Da with a NEMS device<sup>20</sup> will provide the ability to distinguish two species differing by a single Da. For a 1 kDa molecule, this would be equivalent to a mass resolution of 1,000; however, NEMS-MS is capable of measuring molecules greatly exceeding 1 MDa, which means state-of-the-art mass resolution is attainable. With longer measurement times the mass sensitivity can be further enhanced. The upper limit of the mass sensitivity is set by noise sources such as  $1/f$  fluctuations and long-term drifts, which always become predominant at long measurement times. The state-of-the-art of mass sensing with NEMS devices has been improving roughly by about an order of magnitude per year for the last several years; the current record is approaching 100 Da (refs 14, 15, 17, 22, 23). The ultimate limits imposed by thermodynamically driven fluctuation processes have also been theoretically established to be comfortably below 1 Da (refs 19 and 20) for measurement times in the ten millisecond range.

**Mass accuracy.** Mass accuracy is the ability of the instrument to accurately establish the absolute  $m/z$  (mass in NEMS-MS) of the species and is usually expressed in parts per million. A NEMS device with single dalton mass sensitivity, calibrated with an appropriate mass standard, will have a mass accuracy of 100 ppm for a 10 kDa molecule. In NEMS-MS the accuracy increases in direct proportion to the mass of the analyte molecule.

**Mass or  $m/z$  range.** NEMS devices have an exceptionally large mass dynamic range. These devices can easily measure biomolecules with masses of tens of MDa and still be sensitive enough to detect mass changes of a single dalton. The upper limit on the mass is set by the mass of the NEMS device itself. Depending upon the details of adsorption, the properties of the NEMS become affected only when the mass accreted becomes comparable to that of the device itself.

This translates into an upper mass limit of hundreds of MDa for typical devices<sup>15,20</sup>. The lower mass limit in the case of detection of large biomolecules may ultimately become limited by the spatial extent of the molecule compared to that of the NEMS sensor.

**Scan speed.** NEMS devices, in principle, are capable of mass sensitivities of a single dalton for measurement (integration) times in the tens of milliseconds range. Additionally, in future NEMS-MS systems, species of all  $m/z$  or masses will be measured simultaneously using arrays of NEMS devices.

**Efficiency of protein transport from the source to the NEMS detector.** As mentioned, the NEMS-MS system combines the role of analyser and detector into a single unit. This enables a significant reduction in the distance between the protein source and the detector, and thus a corresponding improvement in the efficiency of transport and capture. In the so-called nanoESI-MS systems, for instance, efficiencies as high as 10% have been observed<sup>33</sup>. Loss of analyte molecules arises from a combination of factors such as incomplete desolvation, transmission losses through the ion optics and detection inefficiency. We anticipate that future NEMS-MS systems will be based upon arrays of NEMS devices to provide maximal capture efficiency.

**Parallel processing of the mass information.** In NEMS-based MS systems each NEMS device acts as an individual mass sensor. Here, the capture cross-sectional area has dimensions of approximately  $100 \text{ nm} \times 1,000 \text{ nm}$ . This small cross-section has implications for the parallel-processing abilities of such a system. In 100 s, a modest NEMS-MS system consisting of 1,000 devices could quantitatively process  $1,000 \times 100 \text{ molecules s}^{-1} \times 100 \text{ s} = 10$  million molecules. Integration densities that greatly exceed this have already been realized<sup>34</sup>. This throughput should be sufficient for intensive analyses, for example, on individual mammalian cells.

### Enhancing NEMS-MS efficiency

Transitioning previous demonstrations of NEMS mass sensing, carried out under controlled laboratory conditions, into practical and useful realizations of NEMS-based biological and chemical mass spectrometry—with the potential to process thousands of proteins in tens of milliseconds using only picolitres of sample—will require surmounting challenges in nanoscale systems integration. In future implementations, analytes must be delivered from aqueous phase, stripped of solvent, then delivered to and captured with high efficiency by the NEMS sensor. More challenging will be attaining high capture efficiency; this necessitates a mass detector with large capture cross-section. Given the size of nanoscale detectors, this will be possible only through the use of large sensor arrays positioned in relatively close proximity to the analyte injection point. This will require practical routes to very large-scale integration of NEMS. It is also clearly essential to increase sample throughput in future-generation NEMS-MS systems. The NEMS-MS paradigm is exceptionally well suited to these ends; highly multiplexed configurations involving, potentially, thousands of injectors and detectors can each be co-integrated with microfluidic pre-analysis and delivery components to create a system capable of efficiently analysing extremely small total volumes down to that of an individual cell. For the foreseeable future, we believe that the recent advances in top-down wafer-scale nanofabrication processes provide the only viable avenue to the requisite level of systems complexity. Accordingly, our current work towards the advancement of NEMS-MS is focused upon NEMS arrays fabricated in this manner<sup>34</sup>.

### Methods

**Protein/nanoparticle detection and frequency jump extraction.** To accommodate the stochastic sequential arrival of individual protein molecules or nanoparticles we automate our real-time analysis process. This involves two separate procedures. First,

we continuously track resonant frequency in real time, using a low-noise UHF phase-locked loop (PLL) while protein ions or charged nanoparticles are injected by electrospray and delivered to the NEMS sensor by the ion optics. Figure 2 shows a typical experimental time record of the changes in resonant frequency of a phase-locked NEMS mass sensor under such conditions. Note, that each abrupt frequency jump downwards is the result of a single protein molecule or nanoparticle landing on the NEMS mass sensor. Second, we automate the numerical extraction of the jump heights (frequency shifts) for each individual adsorption event observed in these time records. Our procedure is to reject jumps smaller than twice the frequency resolution of the phase-locked NEMS sensor, because any such smaller events will be increasingly biased by false counts associated with the 'noise floor' set by the frequency instability of the phase-locked NEMS resonator. In our current measurements this instability is characterized by a typical Allan deviation of  $\sigma_A(\tau) \approx 2 \times 10^{-7}$ , for measurement integration times  $\tau$  of the order of several seconds. This corresponds to a one-standard-deviation frequency resolution of  $\delta f \approx 250 \text{ Hz}$  and mass noise floor of  $\sim 10 \text{ kDa}$  in the experiments (see Supplementary information). For the remaining events, identified by our automated and unbiased numerical procedure as experimentally significant ( $\Delta f \geq 2 \times \delta f$ ), we extract their corresponding jump heights (frequency shifts). This involves fitting the time record of each jump to the known temporal step-response function for our phase-locked NEMS system using a nonlinear least-square fit. The response function is separately calculated by PLL circuit theory, and has been experimentally verified, separately, for each NEMS mass sensor used in this work.

**Physisorption.** To ensure stable adsorption and immobilization of individual proteins on the NEMS detector, the detector stage is maintained at a temperature of  $\sim 40 \text{ K in vacuo}$ . At reduced temperature, physisorption due to van der Waals forces ensures the proteins or nanoparticles adsorb and become immobilized upon the detector's surface. Note that detector cooling is required primarily for stable protein adsorption, not for enhanced noise performance. Physisorption is also an ideal method for analyte immobilization in that it is non-specific and it enables detector 'recycling' by periodically warming the NEMS sensor to desorb accreted species.

**Measurement electronics.** The detection circuitry uses a bridge circuit to null the UHF background near the NEMS resonance<sup>4,16–18</sup> and a frequency-modulated PLL (FM-PLL) to track the NEMS resonant frequency in real time<sup>15</sup>. For the very low particle flux used in this initial work, a PLL time constant of several seconds ensures each abrupt frequency jump event is recorded with many data points (Fig. 2). We determine the temporal stability of the PLL system and characterize the frequency-fluctuation noise background by operating in phase-lock over extended intervals ( $\gg 1,000 \text{ s}$ ), both with and without the activation of ESI.

We also monitor the ion current reaching the detector stage with a Faraday cup placed in close proximity to the NEMS detector, which is connected to a high-resolution electrometer. In this first-generation prototype, we do not attempt to demonstrate the potential of high-throughput NEMS-MS; the present system configuration provides a conveniently infrequent analyte arrival rate at the NEMS sensor. A typical current of  $\sim 1 \text{ pA}$  observed at the Faraday detector yields about two adsorption events per minute.

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### Author contributions

A.K.N. and M.S.H. fabricated devices, performed experiments, analysed results and carried out some simulations. W.K.H. designed and assembled the system and performed the initial experiments. X.L.F. made the devices and did the initial phase-locked loop measurements. M.L.R. conceived the project and provided overall guidance throughout. All authors discussed the results and were involved in the analyses and manuscript preparation.

### Additional information

Supplementary information accompanies this paper at [www.nature.com/naturenanotechnology](http://www.nature.com/naturenanotechnology). Reprints and permission information is available online at <http://npg.nature.com/reprintsandpermissions/>. Correspondence and requests for materials should be addressed to M.L.R.