REVIEWS

Sex differences in molecular neuroscience: from fruit flies to humans

Elena Jazin * and Larry Cahill *

Abstract | A plethora of discoveries relating to sex influences on brain function is rapidly moving this field into the spotlight for most areas of neuroscience. The domain of molecular or genetic neuroscience is no exception. The goal of this article is to highlight key developments concerning sex-based dimorphisms in molecular neuroscience, describe control mechanisms regulating these differences, address the implications of these dimorphisms for normal and abnormal brain function and discuss what these advances mean for future work in the field. The overriding conclusion is that, as for neuroscience in general, molecular neuroscience has to take into account potential sex influences that might modify signalling pathways.

Epigenetics

Changes in phenotype caused by mechanisms other than changes in the underlying DNA sequence (hence the name epi — 'in addition to' — genetics).

Sex bias

(Also known as sexual dimorphism.) The systematic difference in form or function between individuals of a different sex in the same species. Body features that are affected by sex bias include colour of skin or coat (fur, feathers, et cetera), size and the presence or absence of body parts or behaviours.

*Department of Genetics and Development, Evolutionary Biology Centre, Uppsala University, SE-75236, Sweden. *Center for the Neurobiology of Learning and Memory, University of California, Irvine, California 92697-3800, USA. Correspondence to E.J. e-mail: <u>elena.jazin@ebc.uu.se</u> doi:10.1038/nrn2754

NATURE REVIEWS | NEUROSCIENCE

Sex influences are being uncovered at an accelerating pace at all levels of neuroscience — including the molecular level, with sex differences in gene expression and epigenetic regulation. Recent progress has been made by mapping differential expression of genes according to sex in several species from worms to man¹-7. These studies show that genes are expressed in a sex-biased manner in many tissues, including the brain. Together with substantial prior evidence of morphological and functional brain dimorphisms, this has raised awareness of the importance of sex in molecular neuroscience⁸. Little is known about the mechanisms that control these sex differences in development and in the adult vertebrate brain.

In principle, all differences are ultimately controlled by the gonadal sex determination systems in each species9. Clearly, among the factors controlled by the sex chromosomes, hormonal influences are highly important, and extensive reviews on gonadal hormones and their actions, in particular in the CNS, are available 10-12. In short, testosterone is the main male hormone. It is secreted by the testes of mammals during late gestational and neonatal periods and causes significant brain sexual dimorphism. It is also produced in small amounts by the ovaries of females. Oestrogens, usually known as female hormones, have vital roles in both sexes. In females they are produced primarily by developing follicles in the ovaries, the corpus luteum and the placenta, circulate in the bloodstream and bind to oestrogen receptors in many target tissues, including the brain. Although it may seem counterintuitive, the male brain is masculinized

by testosterone only after it has been transformed by the enzyme aromatase into oestrogen, which crosses the blood-brain barrier and enters the male brain. Proposed mechanisms of action of sex hormones in the brain include influences on neurogenesis, cell migration, cell differentiation, cell death, axon guidance and synaptogenesis. However, multiple lines of evidence indicate that hormone-independent mechanisms under the control of the sex chromosome complement are also important¹³⁻²⁰ — the extensive effects of hormones and their regulators are insufficient to explain all of the sex differences in the CNS. Exactly how the different sex chromosomes influence a myriad of somatic genes in both a tissue- and a developmental stage-specific manner remains largely unknown. We focus here on a few known sex-biased regulatory mechanisms operating in the brain independently of steroid hormones. The emerging view is that although many sex-biased genes evolved recently, others show conserved patterns of sex-biased activation. This is true not only for genes expressed in the gonads but also for genes with sex bias in the brain^{5,21-32}.

This Review begins by surveying some of the many, often surprising, sex differences in gene expression reported to date, including those involving sex and somatic chromosomes (FIG. 1). We then consider mechanisms that potentially underlie these differences, such as those involving gene splicing and epigenetics, and also discuss the contribution of bioinformatics to illuminating such control mechanisms. Finally, we close with a brief consideration of the vast clinical relevance of this topic.

Gonadal sex determination
The biological mechanism that
induces the development of
the ovaries or testes in an
organism. In many species it
is genetically determined by
the presence of specific
chromosomes called
sex chromosomes

Sex-biased brain gene expression in many species

Early studies based on the analysis of very few gene sets³³ suggested that sex bias in gene expression is large in some somatic tissues but limited in the human brain³⁴. More recent genome-wide expression studies revealed extensive sex dimorphism in gene expression levels in the rodent brain⁴. This preceded gonadal differentiation¹⁵, indicating that it is independent of hormone action. Interestingly, sex-dependent gene expression patterns differ between the gonads and non-reproductive tissues, suggesting that different control mechanisms operate in these tissues. It is now becoming clear that sex bias in gene expression is extensive in the adult brain of all mammalian species⁷ and also occurs in other vertebrates and invertebrates, such as *Drosophila melanogaster*.

Sex differences in human and non-human primates. Influences of aging on gene expression are different in the brains of women and men³⁵. One key question concerns whether these dimorphisms result from environmental differences between the genders — including, for example, food intake, drug use, education differences and other cultural influences that have been shown to affect gene expression differently in both sexes — or whether these gender differences are inherited. A recent evolutionary study showed that a 'gene signature' of sexbiased expression (defined as a pattern of two or more genes altered in a sex-specific way in two or more primates) was conserved during the evolution of primates, including humans5. These results not only show that genes are involved in sex bias in the brain of primates but also suggest that at least some of the mechanisms for the control of sex biases are inherited.

Sex differences in rodents. A general mechanism or regulatory cascade for controlling sex differences in gene expression in the mammalian brain is not known. However, the rapidly increasing numbers of reports of sex differences in gene expression in rodents might lead to the discovery of potential mechanisms. In the past, the extent of sex-biased effects in mammals was obscured by the fact that most investigators examined animals of only one sex (typically males)36-41 or did not specifically mention the sex of the tested animals 42-44. For example, in knockouts of the arginine vasopressin receptor1a, anxiety-like behaviour was initially reported only in male mice45, and only later was it reported that the changes were sex specific, with lack of anxiety-like effects in female knockout mice46. It is noteworthy that there has been a recent increase in knockout mouse studies that have reported results from both sexes in separate analyses. Not uncommonly, key effects prove to be sex specific: a genetic manipulation may have an effect in only one sex or may have the opposite effect in the sexes (TABLE 1). Indeed, such sex differences are the 'elephant in the room' for knockout studies.

Drosophila studies. Recent investigations in D. melanogaster stimulated sex-related research efforts in other organisms. As much as 50% of the D. melanogaster genome exhibits sexually dimorphic expression⁴⁷. A large sex bias remains when the gonads are removed⁴⁸, indicating that this bias is independent of hormonal control. These differences are even more prominent when alternative splicing is taken into account, as up to 22% of the alternatively spliced genes that produce multiple transcripts have sex-specific bias in the proportion of their splice variants⁴⁹. This extreme sex divergence is one of the most important mechanisms for molecular evolution and speciation in flies⁵⁰. As in rodents and humans, genderbiased gene expression in flies is tissue specific⁴⁸. One caveat, however, in evaluating the importance of previous studies in *D. melanogaster* for neuroscience is that all the work was carried out using the complete head, which is mostly composed of large eyes. Future specific analysis of different brain regions will probably clarify this point and might provide exciting insights.

Sex-specific neuronal networks Drosophila melanogaster • Ratio of sex chromosomes to autosomes • Female-only expression of Sxl • Female-only expression of tra • Sex-specific splicing of fru • Sex-specific splicing of dsx Caenorhabditis elegans Hermaphrodite • No central (i.e. gonadal) regulator of sexual dimorphism • tra-1 acts in sexually dimorphic somatic cells • mab-3 involved in sex-specific Sex-specific neuronal networks? Mammals e.g. mice and humans • Genes that escape X inactivation • Y chromosome genes Genes controlling sex hormone production • Epigenetic regulation? • DMRT genes?

Figure 1 | Early genetic control of sex determination in the CNS. The top panel lists the main genetic determinants of somatic sex differences in the brains of Drosophila melanogaster and Caenorhabditis elegans. In both cases, a cascade of expression of regulatory genes results in the formation of male- and female-specific neuronal networks. The regulatory pathway of genes includes a homologous gene named doublesex in D. melanogaster and mab-3 in C. elegans. More specific information about each of the genes, their functions and regulation can be found in the main text. The resulting sex-specific neuronal networks are labelled in green in the D. melanogaster brains. Sex-specific neuronal differences in C. elegans are indicated by red and yellow circles in the hermaphrodites and blue circles in the males (the circles represent neurons contributing to sex-specific networks). The bottom panel lists proposed early determinants of sex differences in the brains of mammals. Among them, the DMRT genes deserve special mention as they are the only genes known to be conserved during evolution from flies to mammals. Some of the members of this family have well-known functions in the sex determination of gonads. Others are exclusively expressed in the brain and may therefore be important for the sex determination of this tissue. Similar to the situation in flies and nematodes, genetic control of sex-specific brain development in mammals may result in the formation of sex-specific neuronal networks — the search for these is an important challenge for the future. dsx, doublesex; fru, fruitless; Sxl, Sex lethal; tra, transformer.

Gonadal hormones

(Also called sex steroids or sex hormones.) Hormones produced in the gonads, including oestrogen and testosterone. These hormones interact with oestrogen or androgen receptors.

Gonad

The organ that makes gametes, the germ cells used for fertilization. The gonads in males are the testes or testicles and the gonads in females are the ovaries.

Hormone dependent or independent?

In most of the examples in TABLE 1 it is not known whether the control of sex differences is independent of sex hormones. Recently, the evaluation of hormone-dependent versus -independent effects in the mouse brain has been greatly facilitated by genetic manipulations of the *SRY* gene that led to the generation of XX males and XY females^{20,51}. *SRY* is the key regulator for the formation of the testes in mammals. When the testes are formed, testosterone is produced, with well-known masculinizing consequences for all somatic tissues, including the brain. In *Sry*-manipulated animals, the sex of the gonads is separated from the chromosomal sex (BOX 1). Recent experiments using *Sry*-manipulated mice

have revealed multiple sex chromosome influences that are independent of hormonal effects, including on protein translation⁵², nociception¹⁹ and apoptosis as well as behaviours such as aggression⁵³ and habit formation¹⁸ (BOX 1). Therefore, future studies should systematically examine and report potential sex biases, including the lack of an effect of a genetic manipulation in, for example, females, as these results are equally important for the overall understanding of gene–function relationships.

X chromosome inactivated genes

There is large diversity in the genetic mechanisms that determine gonadal sex in different species. In many animals, such as flies, birds and mammals, gender is

Table 1	Some sex-s	pecific effects	in knockout mice
---------	------------	-----------------	------------------

Gene symbol	Protein name	Mouse type	Effect	Sex	Refs
<u>Htr1b</u>	Serotonin receptor 1B	KO	Enhanced aggressive behaviour	Male	101
Htr1b	Serotonin receptor 1B	КО	Increased body weight from birth Increased body weight after 8 weeks of age	Male Female	102
Acvr2a	Activin receptor type II	KO	Deficit in reproductive behaviour	Male*	103
Apoe	Apolipoprotein E		Impairments in learning a water maze task and in vertical exploratory behaviour	Female > male	104
<u>App</u>	Amyloid precursor protein	MP	Lower Cu ²⁺ and higher Mn ²⁺ levels	Female	105
Арр	Amyloid precursor protein	MP	Higher levels of AB_{1-40} and higher levels of lipid peroxidation products	Female	106
<u>Ar</u>	Androgen receptor gene	NM	Better perfomance in the water maze	Female	107
<u>Bdnf</u>	Brain-derived neurotrophic factor	CON-forebrain	Hyperactivity Increased depression-like behaviour	Male Female	98
<u>Camkk2</u>	Calcium/calmodulin-dependent protein kinase kinase 2, beta	КО	Long-term memory impairment and regulation of alternative splicing factors	Male	76
<u>Cb1</u> (also known as Cnr1)	Cannabinoid receptor type 1	КО	Increased struggling in the forced-swim test	Males	108
<u>Er</u> (also known as <i>Sfn</i>)	Oestrogen receptor	Luc-ERE	Oestrogen receptor activity in CNS before gonad formation	Female	109
<u>Gabrd</u>	GABA _A receptor, delta subunit	KO	Enhanced acquisition of tone and context fear	Female	110
M1 (also known as Chrm1)	Muscarinic receptor 1	КО	Decreased corticosterone response to muscarinic agonist	Female	111
M2 (also known as Chrm2)	Muscarinic receptor 2	КО	Increased ACTH responses to muscarinic agonist	Male	111
<u>Mc1r</u>	Melanocortin 1 receptor	KO	к-Opioid analgesia	Female	112
Npas2	Neuronal PAS domain protein 2	KO	No rebound in NREM sleep after sleep deprivation	Male	113
<u>Park2</u>	Parkin 2	КО	Increased density of cannabinoid receptors in substantia nigra	Female	114
<u>Sert</u> (also known as Slc6a4)	Serotonin transporter	КО	Increased serotonin synthesis	Female > male	115
<u>Th</u>	Tyrosine hydroxylase	LacZ	Increased TH expression after gonadectomy Decreased TH expression	Males Female	116
<u>Trp2</u>	tRNA proline 2	КО	Unable to recognize the sexual identity of their conspecifics	Males*	117
Not applicable	Not applicable	Ts65Dn	Lower ACTH and higher corticosterone levels after predator exposure	Female	118
<u>V1ar</u> (also known as Avpr1a)	Arg vasopressin receptor 1a	КО	Impaired social recognition and reduced anxiety-like behaviour	Male	46

ACTH, adrenocorticotropic hormone; CON, conditional knockout; ERE, estrogen-responsive element; KO, knockout; LacZ, LacZ-transgenic mice containing the LacZ gene under the control of the promoter of the gene under study; Luc, mice containing a luciferase reporter; MP, transgenic animals with a mutated protein; NM, natural mutant; NREM, non-rapid eye movement; Ts65Dn, mice with trisomy in 136 genes used as a model for Down syndrome. *No females were tested.

determined ultimately by the presence of sex chromosomes in the fertilized egg, whereas other organisms, such as many reptiles, fish and amphibians, display temperature- or behaviour-dependent sex determination⁵⁴. Among the organisms with sex chromosomes, different complements have evolved. For example, whereas it is the male flies, rodents and humans that have two different sex chromosomes (X and Y), it is the female birds (with Z and W) that are hemizygous⁹.

Genes on the sex chromosomes could potentially control brain development and brain function in many ways, should they function in a sexually dimorphic manner during brain development. Well-tuned control systems ensure that many of the X chromosome genes are equally expressed in both sexes despite being present at the ratio 2/1 in females and males. Compensation can be achieved by increasing or decreasing gene expression, and such mechanisms are known to occur in nematodes55,56, D. melanogaster57,58, birds59 and mammals⁶⁰. In mammals, for example, gene expression is tuned down through the inactivation of one of the X chromosomes in females, a process that is initiated by a gene named XIST. The neuroscientific significance of these compensation mechanisms is illustrated by the fact that individuals with greater or fewer copies of the sex chromosomes exhibit multiple neurodevelopmental and behavioural abnormalities61,62. However, it is also now apparent that many genes that are located on the X chromosome are expressed at different levels in females and males⁶³, escaping the regulatory compensation system. This is called 'escape of X inactivation'. For example, six X-linked genes, which are paralogues of Y-encoded genes (*Usp9x*, *Ube1x* (also known as *Uba1*), Smcx (also known as Kdm5c), Eif2s3x, Utx (also known as Kmd6a) and $\underline{Dbx1}$), are expressed in the adult female mouse brain at significantly higher levels than in the brains of males⁶⁴.

In addition, a disproportionately high number of genes on the X chromosome are involved with mental function⁶⁵. Future studies of X chromosome genes that escape inactivation during brain development will be

required for a complete understanding of the genetically based control of sex differences in brain function and cognition.

Regulation of Y and X chromosome paralogues

As mentioned above, some of the genes encoded on the Y chromosome are paralogues for X chromosome genes. It has been suggested that the expression of these genes might compensate for differences in gene dosage due to escape from X inactivation. However, in several cases the two paralogues are regulated in different ways and therefore may have different functions as they may not be expressed in the same cells at the same time.

One example is the protocadherins (PCDHs), members of the cadherin superfamily that are involved in cell-cell interactions during the development of the CNS31. In humans the X-linked (PCDHX) and Y-linked (PCDHY) genes share 98.3% amino acid identity, but the small differences in amino acid sequence result in a longer signal peptide (that may contribute to differences in processing and export) and a shorter cytoplasmic domain (that may result in the activation of different intracellular signalling pathways) in the Y paralogue³¹. Furthermore, PCDHX escapes from X inactivation66 and although both the PCDHX and the PCDHY genes are mainly expressed in the brain, the promoters of the two paralogues are different and there is regional specificity, for example PCDHX is predominantly expressed in the cerebellum

Another example of X and Y chromosome paralogues that potentially operate in a different manner in the brain are the histone demethylases UTX and UTY⁶⁷. In mice UTX escapes X inactivation, and UTX and UTY have different expression patterns in the brain, particularly in the hypothalamus and amygdala. These differences may result in altered demethylation of histone 3 and therefore in differences in the epigenetic regulation of gene expression between the sexes (see the section on epigenetic regulation below).

The Y chromosome encodes SRY, which acts as the key regulator for the formation of the testes in mammals

Genome-wide expression analysis

Examination of RNA expression variation across the human genome, designed to identify associations with observable traits.

Alternative splicing

(Also known as differential splicing.) Variations of the splicing mechanism in which the exons of the primary gene transcript are separated and reconnected so as to produce alternative ribonucleotide arrangements.

SRY

(Sex-determining region Y). A gene encoded on the Y chromosome in many placental mammals. It encodes a transcription factor that initiates the formation of the testicles in males.

X inactivation

The process by which one of the two X chromosomes in female mammals is not expressed. Inactivation occurs at random in each cell, resulting in a mosaic of expression in each XX individual.

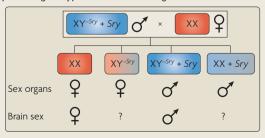
Paralogues

One type of homologous gene. Homologous genes are those that have a common ancestor. Paralogous genes were separated during evolution by a gene duplication event.

Box 1 | Addressing the dependence of sex differences on sex hormones in the brain

An ideal way to evaluate whether a sex bias in the brain is controlled by sex hormones or by the chromosome complement is to dissociate the production of sex hormones from the presence of a Y chromosome. To accomplish this, transgenic mice were first constructed so that the sex determination gene *Sry* was removed from the Y chromosome. The resulting heterochromosomal individuals were called 'XY^{-Sry*} to indicate the absence of *Sry*. In the absence of this gene, no testicles are formed and no testosterone is produced. In a second set of constructs, the *Sry* gene was reinserted into the genome, but this time on one of the autosomes. These 'XY^{-Sry*} the original seven one of these males is crossed with a regular female, the progeny has any of the four possible genotypes outlined in the figure.

A gene with sexual dimorphism in the brain of normal females and males would present a female phenotype in the XY^{-5ry} individuals if the dimorphism were controlled by sex hormones but a male phenotype if it were controlled by the presence of a Y chromosome. Similarly, gonadal control would produce a male phenotype in 'XX + Sry' brains, whereas Y chromosome-mediated control would result in a female phenotype. This 'four core genotype' model thus provides a powerful new way to disentangle hormonal and chromosomal effects on brain development and function.



(discussed above). SRY is also expressed in the human brain^{13,68,69}, but its precise role during brain development remains unknown.

Differential splicing of somatic genes with sex bias

The male-specific expression of sex-biased alternative splice variants in the brain has recently been observed in multiple species. At present, the expression of different splice variants of the somatic gene *fruitless* (*fru*) in *D. melanogaster* neurons is the most elegant example of sex-biased differential gene splicing that is necessary for phenotypic sex differences in the brain⁷⁰. In this case, the expression of FRUM induces the formation of a male-specific neuronal network in the brain that is involved in male mating behaviour.

The cascade responsible for the sex-specific splicing pattern of fru transcripts is well known (FIG. 1). Briefly, the ratio of sex chromosomes to autosomes activates in females the expression of the gene Sex lethal (Sxl). Sxl regulates the splicing of the transformer (tra) gene transcripts, resulting in female-only expression of TRA. TRA, together with TRA2 (expressed in both sexes), controls the splicing of fru transcripts, producing female- and male-specific isoforms of a transcription factor (FRUF and FRUM, respectively). FRUM is specifically expressed in a subset of neurons that promote male-specific behaviour, including courtship⁷¹. The neuronal network that these neurons constitute is stimulated by a male-specific pheromone called *cis*-vaccenyl acetate (cVA) that binds to the odorant receptor OR67D, which is present in one set of FRUM-expressing cells⁷².

This well-established example in flies suggests that differential splicing of somatic genes may also be a key mechanism for the control of sex differences in the brains of other species. For example, gender-biased differential alternative-splicing patterns of the transcriptional cofactor CA150 are observed in the parasitic worm *Schistosoma mansoni*⁷³. This could generate a cascading effect, influencing the transcription and differential splicing of several mRNAs, including those involved in egg laying. Interestingly, the human homologue of the CA150 protein, which is expressed in the brain particularly in striatal neurons, has been described as a transcriptional cofactor that influences the alternative splicing of genes *in vitro*.

In teleost fish, the transcription factor KISSPEPTIN-1 receptor, which has been implicated in the onset of puberty, was recently shown to be spliced in a sex-dependent and developmental-stage-dependent manner in the brain and developmental stage-dependent manner in the brain was different from that observed in the gonads, suggesting that there are different functions and control mechanisms in these two tissues.

Sex bias of differential splicing in the brain has recently been observed in multiple species. Interestingly, the Indian mugger crocodile's sex is determined by the incubation temperature of the developing embryos⁷⁵. In these reptiles, a temperature-sensitive promoter and temperature-sensitive splicing factors may regulate the transcription factor Dmrt1, which is encoded by a somatic gene. This gene's transcripts also undergo

sex-biased differential splicing in mammals, but the control mechanisms as well as the function of each splice variant remain to be elucidated.

In mammals, one example of sex-biased alternative splicing in the brain involves upregulation of the splicing factors PTB-associated splicing factor (<u>PSF</u>; also known as SFPQ) and <u>SRP20</u> (also known as SFRS3)⁷⁶, both of which are expressed at higher levels in the male rodent hippocampus. In humans, Ca²⁺ channels — in particular voltage-gated Ca²⁺ channels, which have key roles in neurotransmitter release — are extensively spliced in an age- and gender-biased manner⁷⁷.

Although genome-wide approaches to search for differential expression in mammals are not yet well developed, recent results indicate widespread sex-biased expression of splice variants in mouse liver⁷⁸; genomewide studies investigating brain tissue are pending. These analyses are likely to reveal the extent and importance of sex-biased differential splicing in the brain.

Lessons from nematodes

Nematodes are a classical model used to study the development and composition of all cells in the CNS. Moreover, the process of sexual differentiation of the CNS has been elucidated in detail only in Caenorhabditis elegans. The hermaphrodites and the males of C. elegans share 294 neurons, but both sexes also have sex-specific neurons — 8 in hermaphrodites and 89 in males — that are essential for sex-specific behaviours. These differences, as well as all somatic differences, are ultimately controlled by the tra-1 gene, a master regulator of C. elegans sexual differentiation (for a more detailed review on sex differences in *C. elegans*, including in the nervous system, see REF. 79). This gene encodes a transcriptional repressor that blocks the expression of male-specific genes in hermaphrodites. The terminal effector of the TRA-1-regulated pathway in CNS development in C. elegans is the transcription factor MAB-3, which belongs to a family of somatic transcription factors called the DM-domain genes. This family is the only known conserved molecular link between sex determination systems in metazoans. In fact, these genes are homologous to doublesex in Drosophila spp. and to the DMRT family in vertebrates^{23,26,80–84}. They encode putative transcription factors and have undergone frequent independent events of gene duplication during the course of evolution, resulting in variable numbers of isoforms between phyla. Interestingly, in vertebrates some members of this gene family have lost their expression in the gonads and are expressed only in the brain⁸⁵, suggesting that they have acquired functions related to CNS development or regulation.

Forming a complete understanding of normal brain development in vertebrates will probably involve elucidating the interplay between factors encoded by the sex chromosomes, such as gonad-determining factors, brain-specific proteins and hormonally controlled mechanisms that collectively control somatic sex determination and sex biases in brain function and behaviour (FIGS 1,2). There are probably other, as yet unknown, sex determination factors active during brain development

Genomic imprinting

Different expression of a gene, depending on the sex of the parent who transmits it. One of the alleles is imprinted or marked to be silenced, for example by methylation.

that will be identified in this relatively simple CNS, and their discovery might initiate parallel studies in more complex animals.

Beyond genetics: inherited epigenetic control

Genomic imprinting affects the development of several mental disorders in a sexually dimorphic manner⁸⁶, and there is accumulating evidence for effects of other inherited epigenetic mechanisms, including DNA methylation, histone modifications, nucleosome repositioning, higher-order chromatin remodelling, mechanisms involving non-coding RNA, and RNA and DNA editing87,88. Recent experiments suggest that inherited epigenetic control should be added to the list of hormone-independent mechanisms that regulate sex differences in the brain. For example, histone 3 modifications are sexually dimorphic in the developing mouse brain independently of testosterone treatment89. Moreover, as indicated above, histone demethylases encoded on the X and Y chromosomes⁶⁷ also contribute to epigenetic mechanisms that regulate sex differences. It is likely that epigenetic mechanisms constitute an important. although largely unexplored, part of genetically based sex influences on the brain.

Control of sex differences in the primate brain

Specific regulators acting early during brain development, before the gonads mature, and resulting in the numerous sex differences found in the adult primate brain have not yet been identified. However, indirect regulators acting after the formation of the gonads, such as oestrogens and androgens, are well-known factors for sex differences in the brain. Oestrogen signalling is primarily conveyed by the oestrogen receptors alpha and beta. These receptors bind to specific DNA sequences — estrogen-responsive elements (EREs) — resulting in the transcriptional activation of genes⁹⁰ (FIG. 2). Similar to oestrogens, androgens exert their function by binding to androgen receptors, which in turn cause the expression of target genes by binding to different sets of androgen-responsive elements (AREs). Recent advances using computational methods make it possible to identify different types of AREs, called classical and specific AREs, in mammals⁹¹.

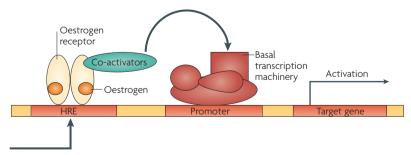


Figure 2 | **Regulation of gene expression by hormone receptors.** Oestrogen molecules form a complex with oestrogen receptors (ERs). This complex can then bind a hormone-responsive element (HRE) (in this case an oestrogen-responsive element (ERE)). The ERE–ER complex interacts through co-activators with the basal transcription machinery to increase the transcription of target genes in a hormone-dependent manner.

More research is required to elucidate the role of these recently identified AREs in the establishment of brainspecific gender differences. The presence of direct regulators acting independently of sex hormones is suggested by systematic bioinformatic searches of sex-biased genes in primate brains, which have demonstrated that many of these genes do not contain any known AREs or EREs5. The absence of hormoneresponsive elements is not by itself proof of the presence of hormone-independent regulators. However, this fact, together with the demonstration that some sex biases in the brain are controlled in many species by sex chromosomes independently of hormones¹³⁻²⁰, strongly suggests the existence of somatic regulators in the brain. These additional regulators should share several characteristics. First, they should be expressed in brain tissues, possibly during development. Second, they should show some form of sexual dimorphism in expression and/or function. Third, their expression in one sex should be finally controlled by the sex chromosome complement. Somatic regulators of sex differences in the brain probably act in a cascade of events similar to those described above for flies and nematodes.

As is the case for rodents, the first candidates in a search for genes that regulate sex determination in somatic tissues in primates are genes encoded on the sex chromosomes. The gene that encodes the transcription factor SRY (discussed above) has been named as potentially the first regulatory gene to have a role during sex-specific brain development in primates, based on its expression in specific cells in the adult CNS^{92,93}. However, although SRY is known to be expressed in specific cells in the adult human brain⁶⁸, its potential brain-specific functions remain to be elucidated. Moreover, in a recent study using brains from human embryos, although onethird of the Y chromosome-encoded genes are highly expressed in the brain before birth, SRY expression could not be detected in any of the regions analysed. This suggests that the main controller of sex bias in the brain is different from the one that controls the formation of the testicles94.

Many other genes encoded in the sex chromosomes are candidates and may act early in regulatory cascades that determine sex differences. The current version of the database Ensembl for Homo sapiens includes 1,155 genes on the X chromosome, of which 39 are transcription factors, and 130 genes on the Y chromosome, of which 5 (including SRY) are transcription factors. Although many genes on the X chromosome are expressed in the developing brain, only a fraction of those escape X inactivation95. In some cases, X inactivation starts at very early stages of embryonic development, well before CNS formation, when the embryos are composed of only a few cells95. Of the genes that escape X inactivation, only those expressed later in development, during the period of brain formation, are potential regulators of sex bias in the brain. A systematic search of sex-biased genes expressed in the primate brain during development will be needed to identify early somatic determinants of dimorphic brain development.

Clinical consequences of sex-biased genes

Attention to sex differences that are caused directly or indirectly by changes in the sex chromosome complement has many implications beyond the understanding of the basic physiology of the brain. In fact, many of the genes associated with animal models of several neurological diseases, such as Huntington's disease (cerebral ischaemia⁹⁷, depression-related phenotypes⁹⁸ and Alzheimer's disease, as well as with cognitive ability models, such as mice lacking functional APOE⁹⁹, have sex biases in disease development, pathological processes and recovery mechanisms. However, precisely which control mechanisms for sex-biased gene expression are affected in these disorders is not known. Understanding the mechanisms that underlie the diseases will be essential for the development of new gender-specific therapies¹⁰⁰.

However, caution should be exercised when translating research from mice to humans, particularly with regard to hormone-dependent effects. Alterations in sex hormone levels can result directly from genetic modification (for example of hormone receptors or their regulators) or from environmental influences, such as the use of contraceptives, diet, hormone therapy and other drugs that modify steroid levels. Both types of influence will have important (although different) implications for understanding and treating clinical disorders. Clearly, more attention needs to be paid to the potential clinical implications of genetically based sex differences, whether direct or indirect.

Conclusions

It is now clear that genetically controlled sex differences are pronounced in the brains of many species, with consequences for normal and abnormal brain physiology. These sex differences result in differences in neuronal activation and network activity and may ultimately influence behavioural traits.

Contrary to well-known pathways for sex determination in the gonads, the main regulators of somatic sex differentiation in most species are not known. Among the mechanisms that control sex-biased gene expression are hormone, sex chromosome, alternative splicing and epigenetic control mechanisms. The lessons learned from *D. melanogaster* and nematodes, together with current research focusing on sex differentiation of the mammalian brain, will help to identify a cascade of somatic regulators involved in the sex differentiation of the brain during development.

Although the identity of the main controllers of brain sex differences is not known in mammals, the sex chromosomes are good candidates for being the first step of the controlling molecular pathways. In this respect, genes on the X chromosome that escape dosage-related compensation of expression levels, as well as genes on the Y chromosome that are expressed during early brain development, are good candidates for mediating the early phase of establishing sex differences. Sex chromosome genes may in turn control regulatory cascades relating to the expression of other genes encoded in the autosomes. At least some of the sex-biased controlling genes, such as SRY, are conserved during evolution, and therefore evolutionary studies might provide a new means of searching for the mechanisms that control sex differences in the brain.

Regardless of the current lack of answers to many important mechanistic questions, most notably those concerning sex-biased expression of somatic genes, there is overwhelming evidence for genetically based sex influences on brain function, clearly indicating that increased awareness of, and systematic investigation into, molecular-level sex influences will yield tremendous dividends for both basic and clinically applied brain science.

- Fitzpatrick, J. M. et al. An oligonucleotide microarray for transcriptome analysis of Schistosoma mansoni and its application/use to investigate genderassociated gene expression. Mol. Biochem. Parasitol. 141, 1–13 (2005).
- 2. Jin, W. et al. The contributions of sex, genotype and age to transcriptional variance in Drosophila melanogaster. Nature Genet. 29, 389–395 (2001). A genome-wide analysis of sex bias in gene expression in flies, demonstrating that around one-half of the transcriptome differs significantly between the two sexes and that adult age has remarkably little effect on transcriptional variance.
- Santos, E. M. et al. Molecular basis of sex and reproductive status in breeding zebrafish. Physiol. Genomics 30, 111–122 (2007).
- Yang, X. et al. Tissue-specific expression and regulation of sexually dimorphic genes in mice. Genome Res. 16, 995–1004 (2006).
- Reinius, B. et al. An evolutionarily conserved sexual signature in the primate brain. PLoS Genet. 4, e1000100 (2008).
 - This article demonstrates that some human sex-specific gene expression patterns are conserved in the brains of other primates. Conservation across species indicates that these patterns may underlie some genetic differences between the sexes.
- Zhang, W., Bleibel, W. K., Roe, C. A., Cox, N. J. & Eileen Dolan, M. Gender-specific differences in expression in human lymphoblastoid cell lines. *Pharmacogenet. Genomics* 17, 447–450 (2007).

- Isensee, J. & Ruiz Noppinger, P. Sexually dimorphic gene expression in mammalian somatic tissue. Gend. Med. 4 (Suppl. B), S75–S95 (2007).
- Cahill, L. Why sex matters for neuroscience. Nature Rev. Neurosci. 7, 477–484 (2006).
 A comprehensive review of multiple sex influences
 - on the anatomy, chemistry and function of the brain, including a discussion of sex effects on brain disorders.
- Arnold, A. P. et al. Minireview: sex chromosomes and brain sexual differentiation. Endocrinology 145, 1057–1062 (2004).
- Flerko, B. Steroid hormones and the differentiation of the central nervous system. *Curr. Top. Exp. Endocrinol.* 1, 41–80 (1971).
- McEwen, B. S. Steroid hormones and the chemistry of behavior. Adv. Behav. Biol. 4, 41–59 (1972).
- Pfaff, D. W. Steroid sex hormones in the rat brain: specificity of uptake and physiological effects. UCLA Forum Med. Sci. 15, 103–112 (1972).
- Mayer, A., Mosler, G., Just, W., Pilgrim, C. & Reisert, I. Developmental profile of Sry transcripts in mouse brain. *Neurogenetics* 3, 25–30 (2000).
- Agate, R. J. et al. Neural, not gonadal, origin of brain sex differences in a gynandromorphic finch. Proc. Natl Acad. Sci. USA 100, 4873–4878 (2003).
- Dewing, P., Shi, T., Horvath, S. & Vilain, É. Sexually dimorphic gene expression in mouse brain precedes gonadal differentiation. *Brain Res. Mol. Brain Res.* 118, 82–90 (2003).
- Glickman, S. E., Short, R. V. & Renfree, M. B. Sexual differentiation in three unconventional mammals:

- spotted hyenas, elephants and tammar wallabies. Horm. Behav. 48, 403–417 (2005).
- Scholz, B. et al. Sex-dependent gene expression in early brain development of chicken embryos. BMC Neurosci. 7, 12 (2006)
- Quinn, J. J., Hitchcott, P. K., Umeda, E. A., Arnold, A. P. & Taylor, J. R. Sex chromosome complement regulates habit formation. *Nature Neurosci.* 10, 1398–1400 (2007).
- Gioiosa, L. et al. Sex chromosome complement affects nociception in tests of acute and chronic exposure to morphine in mice. Horm. Behav. 53, 124–130 (2008).
- Budefeld, T., Grgurevic, N., Tobet, S. A. & Majdic, G. Sex differences in brain developing in the presence or absence of gonads. *Dev. Neurobiol.* 68, 981–995 (2008).
- Sanders, L. E. & Arbeitman, M. N. Doublesex establishes sexual dimorphism in the *Drosophila* central nervous system in an isoform-dependent manner by directing cell number. *Dev. Biol.* 320, 378–390 (2008).
- Suseendranathan, K. et al. Expression pattern of Drosophila translin and behavioral analyses of the mutant. Eur. J. Cell Biol. 86, 173–186 (2007).
- Cao, J., Cao, Z. & Wu, T. Generation of antibodies against DMRT1 and DMRT4 of Oreochromis aurea and analysis of their expression profile in Oreochromis aurea tissues. J. Genet. Genomics 34, 497–509 (2007).
- Holmes, M. M., Goldman, B. D. & Forger, N. G. Social status and sex independently influence androgen receptor expression in the eusocial naked molerat brain. Horm. Behav. 54, 278–285 (2008).

REVIEWS

- Du, Q. Y., Wang, F. Y., Hua, H. Y. & Chang, Z. J. Cloning and study of adult-tissue-specific expression of Sox9 in Cyprinus carpio. J. Genet. 86, 85–91 (2007).
- Guo, Y. et al. Molecular cloning, characterization, and expression in brain and gonad of *Dmrt5* of zebrafish. *Biochem. Biophys. Res. Commun.* 324, 569–575 (2004).
- Blazquez, M. & Piferrer, F. Sea bass (*Dicentrarchus labrax*) androgen receptor: cDNA cloning, tissue-specific expression, and mRNA levels during early development and sex differentiation. *Mol. Cell. Endocrinol.* 237, 37–48 (2005).
- De Vries, G. J. & Panzica, G. C. Sexual differentiation of central vasopressin and vasotocin systems in vertebrates: different mechanisms, similar endpoints. *Neuroscience* 138, 947–955 (2006).
- Alfonso, J. et al. Regulation of hippocampal gene expression is conserved in two species subjected to different stressors and antidepressant treatments. Biol. Psychiatry 59, 244–251 (2006).
- Pask, A. J. et al. SOX9 has both conserved and novel roles in marsupial sexual differentiation. Genesis 33, 131–139 (2002).
- Blanco, P., Sargent, C. A., Boucher, C. A., Mitchell, M. & Affara, N. A. Conservation of PCDHX in mammals; expression of human X/Y genes predominantly in brain. Mamm. Genome 11, 906–914 (2000).
- Delbridge, M. L., McMillan, D. A., Doherty, R. J., Deakin, J. E. & Graves, J. A. Origin and evolution of candidate mental retardation genes on the human X chromosome (MRX). BMC Genomics 9, 65 (2008).
- Galfalvy, H. C. et al. Sex genes for genomic analysis in human brain: internal controls for comparison of probe level data extraction. BMC Bioinformatics 4, 37 (2003).
- Rinn, J. L. & Snyder, M. Sexual dimorphism in mammalian gene expression. *Trends Genet.* 21, 298–305 (2005).
- Berchtold, N. C. et al. Gene expression changes in the course of normal brain aging are sexually dimorphic. Proc. Natl Acad. Sci. USA 105, 15605–15610 (2008)
- Hesen, W. et al. Hippocampal cell responses in mice with a targeted glucocorticoid receptor gene disruption. J. Neurosci. 16, 6766–6774 (1996).
- D'Hooge, R. et al. Mildly impaired water maze performance in male Fmr1 knockout mice. Neuroscience 76, 367–376 (1997).
- Bao, S., Chen, L., Qiao, X., Knusel, B. & Thompson, R. F. Impaired eye-blink conditioning in waggler, a mutant mouse with cerebellar BDNF deficiency. *Learn. Mem.* 5, 355–364 (1998).
- Xue, L. et al. Carbon monoxide and nitric oxide as coneurotransmitters in the enteric nervous system: evidence from genomic deletion of biosynthetic enzymes. Proc. Natl Acad. Sci. USA 97, 1851–1855 (2000).
- Jung, M. Y., Hof, P. R. & Schmauss, C. Targeted disruption of the dopamine D₂ and D₃ receptor genes leads to different alterations in the expression of striatal calbindin-D_{28k}. Neuroscience 97, 495–504 (2000).
- Sora, İ. et al. Molecular mechanisms of cocaine reward: combined dopamine and serotonin transporter knockouts eliminate cocaine place preference. Proc. Natl Acad. Sci. USA 98, 5300–5305 (2001).
- Takahashi, N. et al. VMAT2 knockout mice: heterozygotes display reduced amphetamineconditioned reward, enhanced amphetamine locomotion, and enhanced MPTP toxicity. Proc. Natl Acad. Sci. USA 94, 9938–9943 (1997).
- Iadecola, C., Zhang, F., Casey, R., Nagayama, M. & Ross, M. E. Delayed reduction of ischemic brain injury and neurological deficits in mice lacking the inducible nitric oxide synthase gene. *J. Neurosci.* 17, 9157–9164 (1997).
- Deans, M. R., Gibsón, J. R., Sellitto, C., Connors, B. W. & Paul, D. L. Synchronous activity of inhibitory networks in neocortex requires electrical synapses containing connexin36. *Neuron* 31, 477–485 (2001).
 Bielsky, I. F., Hu, S. B., Szegda, K. L., Westphal, H. &
- Bielsky, I. F., Hu, S. B., Szegda, K. L., Westphal, H. & Young, L. J. Profound impairment in social recognition and reduction in anxiety-like behavior in vasopressin V1a receptor knockout mice.
- Neuropsychopharmacology 29, 483–493 (2004).
 Bielsky, I. F., Hu, S. B. & Young, L. J. Sexual dimorphism in the vasopressin system: lack of an altered behavioral phenotype in female V1a receptor knockout mice. Behav. Brain Res. 164, 132–136 (2005).

- Ranz, J. M., Castillo-Davis, C. I., Meiklejohn, C. D. & Hartl, D. L. Sex-dependent gene expression and evolution of the *Drosophila* transcriptome. *Science* 300, 1742–1745 (2003).
- Parisi, M. et al. A survey of ovary-, testis-, and somabiased gene expression in *Drosophila melanogaster* adults. Genome Biol. 5, R40 (2004).
- McIntyre, L. M. et al. Sex-specific expression of alternative transcripts in *Drosophila*. Genome Biol. 7, R79 (2006).
- Zhang, Y., Sturgill, D., Parisi, M., Kumar, S. & Oliver, B. Constraint and turnover in sex-biased gene expression in the genus *Drosophila*. *Nature* 450, 233–237 (2007).
- De Vries, G. J. et al. A model system for study of sex chromosome effects on sexually dimorphic neural and behavioral traits. J. Neurosci. 22, 9005–9014 (2002)
- Xu, J., Watkins, R. & Arnold, A. P. Sexually dimorphic expression of the X-linked gene *Eli2s3x* mRNA but not protein in mouse brain. *Gene Expr. Patterns* 6, 146–155 (2006).
- 53. Gatewood, J. D. et al. Sex chromosome complement and gonadal sex influence aggressive and parental behaviors in mice. J. Neurosci. 26, 2335–2342 (2006). These results imply that genes on the sex chromosomes affect sex differences in brain and behaviour.
- Crews, D., Coomber, P., Baldwin, R., Azad, N. & Gonzalez-Lima, F. Brain organization in a reptile lacking sex chromosomes: effects of gonadectomy and exogenous testosterone. *Horm. Behav.* 30, 474–486 (1996).
- Meyer, B. J. & Casson, L. P. Caenorhabditis elegans compensates for the difference in X chromosome dosage between the sexes by regulating transcript levels. Cell 47, 871–881 (1986).
- Luz, J. G. et al. XOL-1, primary determinant of sexual fate in C. elegans, is a GHMP kinase family member and a structural prototype for a class of developmental regulators. Genes Dev. 17, 977–990 (2003).
- Amrein, H. & Axel, R. Genes expressed in neurons of adult male *Drosophila*. Cell 88, 459–469 (1997).
- 58. Ruiz, M. F., Esteban, M. R., Donoro, C., Goday, C. & Sanchez, L. Evolution of dosage compensation in Diptera: the gene maleless implements dosage compensation in Drosophila (Brachycera suborder) but its homolog in Sciara (Nematocera suborder) appears to play no role in dosage compensation. Genetics 156, 1853–1865 (2000).
- 59. Arnold, A. P., Itoh, Y. & Melamed, E. A birds-eye view of sex chromosome dosage compensation. Annu. Rev. Genomics Hum. Genet. 9, 109–127 (2008). This excellent review not only describes mechanisms of sex chromosome dosage compensation in birds but also describes and compares these mechanisms with those present in other species, including mammals.
- Nguyen, D. K. & Disteche, C. M. Dosage compensation of the active X chromosome in mammals. *Nature Genet.* 38, 47–53 (2006).
- Tartaglia, N. et al. A new look at XXYY syndrome: medical and psychological features. Am. J. Med. Genet. A 146, 1509–1522 (2008).
- Cutter, W. J. et al. Influence of X chromosome and hormones on human brain development: a magnetic resonance imaging and proton magnetic resonance spectroscopy study of Turner syndrome. Biol. Psychiatry 59, 273–283 (2006).
- Nguyen, D. K. & Disteche, C. M. High expression of the mammalian X chromosome in brain. *Brain Res.* 1126, 46–49 (2006).
- 64. Xu, J., Burgoyne, P. S. & Arnold, A. P. Sex differences in sex chromosome gene expression in mouse brain. *Hum. Mol. Genet.* 11, 1409–1419 (2002).
- Skuse, D. H. X-linked genes and mental functioning. Hum. Mol. Genet. 14, R27–R32 (2005).
- Lopes, A. M. et al. Inactivation status of PCDH11X: sexual dimorphisms in gene expression levels in brain. Hum. Genet. 119, 267–275 (2006).
- 67. Xu, J., Deng, X., Watkins, R. & Disteche, C. M. Sex-specific differences in expression of histone demethylases Utx and Uty in mouse brain and neurons. J. Neurosci. 28, 4521–4527 (2008). A very elegant example of the use of Sry-modified mice to study sex bias effects that are independent from the action of sex hormones.
- 68. Mayer, A., Lahr, G., Swaab, D. F., Pilgrim, C. & Reisert, I. The Y-chromosomal genes SRY and ZFY are transcribed in adult human brain. *Neurogenetics* 1, 281–288 (1998). First description of the expression of SRY, a testis-determining factor, in adult human brain.

- Dewing, P. et al. Direct regulation of adult brain function by the male-specific factor SRY. Curr. Biol. 16, 415–420 (2006).
- Manoli, D. S. et al. Male-specific fruitless specifies the neural substrates of *Drosophila* courtship behaviour. Nature 436, 395–400 (2005).
- Kimura, K., Ote, M., Tazawa, T. & Yamamoto, D. Fruitless specifies sexually dimorphic neural circuitry in the *Drosophila* brain. *Nature* 438, 229–233 (2005).
- Datta, S. R. et al. The Drosophila pheromone cVA activates a sexually dimorphic neural circuit. Nature 452, 473–477 (2008).
- DeMarco, R., Oliveira, K. C., Venancio, T. M. & Verjovski-Almeida, S. Gender biased differential alternative splicing patterns of the transcriptional cofactor CA150 gene in Schistosoma mansoni. Mol. Biochem. Parasitol. 150, 123–131 (2006).
- Mechaly, A. S., Vinas, J. & Piferrer, F. Identification of two isoforms of the kisspeptin-1 receptor (kiss1r) generated by alternative splicing in a modern teleost, the Senegalese sole (Solea senegalensis). Biol. Reprod. 80, 60–69 (2008).
- Anand, A. et al. Multiple alternative splicing of Dmrt1 during gonadogenesis in Indian mugger, a species exhibiting temperature-dependent sex determination. Gene 425, 56–63 (2008).
- Antunes-Martins, A., Mizúno, K., Irvine, E. E., Lepicard, E. M. & Giese, K. P. Sex-dependent up-regulation of two splicing factors, Psf and Srp20, during hippocampal memory formation. *Learn. Mem.* 14, 693–702 (2007).
 Chang, S. Y. *et al.* Age and gender-dependent
- Chang, S. Y. et al. Age and gender-dependent alternative splicing of P/Q-type calcium channel EF-hand. Neuroscience 145, 1026–1036 (2007).
- Su, W. L. et al. Exon and junction microarrays detect widespread mouse strain- and sex-bias expression differences. BMC Genomics 9, 273 (2008).
- Wolff, J. R. & Zarkower, D. Somatic sexual differentiation in *Caenorhabditis elegans*. *Curr. Top. Dev. Biol.* 83, 1–39 (2008).
- Brunner, B. et al. Genomic organization and expression of the doublesex-related gene cluster in vertebrates and detection of putative regulatory regions for DMRT1. Genomics 77, 8–17 (2001).
- Smith, C. A. & Sinclair, A. H. Sex determination in the chicken embryo. *J. Exp. Zool.* 290, 691–699 (2001).
 Volff, J. N., Zarkower, D., Bardwell, V. J. & Schartl, M.
- Volff, J. N., Zarkower, D., Bardwell, V. J. & Schartl, M. Evolutionary dynamics of the DM domain gene family in metazoans. J. Mol. Evol. 57 (Suppl. 1), S241–S249 (2003).
- Huang, X., Hong, C. S., O'Donnell, M. & Saint-Jeannet, J. P. The doublesex-related gene, XDmrt4, is required for neurogenesis in the olfactory system. Proc. Natl Acad. Sci. USA 102, 11349–11354 (2005).
- El-Mogharbel, N. et al. DMRT gene cluster analysis in the platypus: new insights into genomic organization and regulatory regions. Genomics 89, 10–21 (2007).
- 85. Hong, C. S., Park, B. Y. & Saint-Jeannet, J. P. The function of *Dmrt* genes in vertebrate development: it is not just about sex. *Dev. Biol.* **310**, 1–9 (2007).
- Davies, W., Isles, A. R. & Wilkinson, L. S. Imprinted genes and mental dysfunction. *Ann. Med.* 33, 428–436 (2001).
- Vige, A., Gallou-Kabani, C. & Junien, C. Sexual dimorphism in non-Mendelian inheritance. *Pediatr. Res.* 63, 340–347 (2008).
- Breedlove, S. M., Cooke, B. M. & Jordan, C. L. The orthodox view of brain sexual differentiation. *Brain Behav. Evol.* 54, 8–14 (1999).
- 89. Tsai, H. W., Grant, P. A. & Rissman, E. F. Sex differences in histone modifications in the neonatal mouse brain. *Epigenetics* 4, 47–53 (2009). This is the first demonstration that histone modification is associated with neural sexual differentiation.
- Bourdeau, V. et al. Genome-wide identification of highaffinity estrogen response elements in human and mouse. Mol. Endocrinol. 18, 1411–1427 (2004).
- Moehren, U., Denayer, S., Podvinec, M., Verrijdt, G. & Claessens, F. Identification of androgen-selective androgen-response elements in the human aquaporin-5 and Rad9 genes. *Biochem. J.* 411, 679–686 (2008).
- Maatouk, D. M. & Capel, B. Sexual development of the soma in the mouse. *Curr. Top. Dev. Biol.* 83, 151–183 (2008).
- Sekido, R. & Lovell-Badge, R. Sex determination and SRY: down to a wink and a nudge? *Trends Genet.* 25, 19–29 (2009).

- 94. Reinius, B. & Jazin, E. Prenatal sex differences in the human brain. *Mol. Psychiatry* **14**, 987, 988–989 (2009)
- Johnston, C. M. et al. Large-scale population study of human cell lines indicates that dosage compensation is virtually complete. PLoS Genet. 4, e9 (2008).
- Bode, F. J. et al. Sex differences in a transgenic rat model of Huntington's disease: decreased 17ss-estradiol levels correlate with reduced numbers of DARPP32 + neurons in males. Hum. Mol. Genet. 17, 2595–2609 (2008).
- Kitano, H. et al. Gender-specific response to isoflurane preconditioning in focal cerebral ischemia. J. Cereb. Blood Flow Metab. 27, 1377–1386 (2007).
 Monteggia, L. M. et al. Brain-derived neurotrophic
- Monteggia, L. M. et al. Brain-derived neurotrophic factor conditional knockouts show gender differences in depression-related behaviors. Biol. Psychiatry 61, 187–197 (2007).
- Villasana, L., Acevedo, S., Poage, C. & Raber, J. Sexand APOE isoform-dependent effects of radiation on cognitive function. *Radiat. Res.* 166, 883–891 (2006).
- Ober, C., Loisel, D. A. & Gilad, Y. Sex-specific genetic architecture of human disease. *Nature Rev. Genet.* 9, 911–922 (2008).
- Saudou, F. et al. Enhanced aggressive behavior in mice lacking 5-HT1B receptor. Science 265, 1875–1878 (1994).
- Bouwknecht, J. A. et al. Male and female 5-HT_{1B} receptor knockout mice have higher body weights than wildtypes. *Physiol. Behav.* 74, 507–516 (2001).
- 103. Ma, X., Reyna, A., Mani, S. K., Matzuk, M. M. & Kumar, T. R. Impaired male sexual behavior in activin receptor type II knockout mice. *Biol. Reprod.* 73, 1182–1190 (2005).
- 104. Raber, J. et al. Isoform-specific effects of human apolipoprotein E on brain function revealed in ApoE knockout mice: increased susceptibility of females. Proc. Natl Acad. Sci. USA 95, 10914–10919 (1998)
- 105. Maynard, C. J. et al. Gender and genetic background effects on brain metal levels in APP transgenic and normal mice: implications for Alzheimer beta-amyloid pathology. J. Inorg. Biochem. 100, 952–962 (2006).

- 106. Schuessel, K. et al. Impaired Cu/Zn-SOD activity contributes to increased oxidative damage in APP transgenic mice. Neurobiol. Dis. 18, 89–99 (2005).
- 107. Rizk, A., Robertson, J. & Raber, J. Behavioral performance of tfm mice supports the beneficial role of androgen receptors in spatial learning and memory. *Brain Res.* 1034, 132–138 (2005).
- 108. Steiner, M. A. et al. Impaired cannabinoid receptor type 1 signaling interferes with stress-coping behavior in mice. Pharmacogenomics J. 8, 196–208 (2008).
- 109. Ciana, P. et al. In vivo imaging of transcriptionally active estrogen receptors. Nature Med. 9, 82–86 (2003).
- 110. Wiltgen, B. J., Sanders, M. J., Ferguson, C., Homanics, G. E. & Fanselow, M. S. Trace fear conditioning is enhanced in mice lacking the delta subunit of the CABAA receptor. *Learn. Mem.* 12, 327–333 (2005).
- 111. Rhodes, M. E., Billings, T. E., Czambel, R. K. & Rubin, R. T. Pituitary-adrenal responses to cholinergic stimulation and acute mild stress are differentially elevated in male and female M₂ muscarinic receptor knockout mice. J. Neuroendocrinol. 17, 817–826 (2005).
- Mogil, J. S. et al. The melanocortin-1 receptor gene mediates female-specific mechanisms of analgesia in mice and humans. Proc. Natl Acad. Sci. USA 100, 4867–4872 (2003).
- 113. Franken, P. et al. NPAS2 as a transcriptional regulator of non-rapid eye movement sleep: genotype and sex interactions. Proc. Natl Acad. Sci. USA 103, 7118–7123 (2006).
- 114. Gonzalez, S. et al. Cannabinoid CB₁ receptors in the basal ganglia and motor response to activation or blockade of these receptors in parkin-null mice. Brain Res. 1046, 195–206 (2005).
- 115. Kim, D. K. et al. Altered serotonin synthesis, turnover and dynamic regulation in multiple brain regions of mice lacking the serotonin transporter. Neuropharmacology 49, 798–810 (2005).
- 116. Thanky, N. R., Son, J. H. & Herbison, A. E. Sex differences in the regulation of tyrosine hydroxylase gene transcription by estrogen in the locus coeruleus

- of TH9-LacZ transgenic mice. *Brain Res. Mol. Brain Res.* **104**, 220–226 (2002).
- 117. Stowers, L., Holy, T. E., Meister, M., Dulac, C. & Koentges, G. Loss of sex discrimination and male-male aggression in mice deficient for TRP2. *Science* 295, 1493–1500 (2002).
- 118. Martinez-Cue, C., Rueda, N., Garcia, E. & Florez, J. Anxiety and panic responses to a predator in male and female Ts65Dn mice, a model for Down syndrome. *Genes Brain Behav.* 5, 413–422 (2006).

Acknowledgements

The authors would like to thank the Swedish Börgströms Foundation (E.J.) and the National Institute of Mental Health (R01 to L.C.) for their support.

Competing interests statement

The authors declare no competing financial interests.

DATABASES

Entrez Gene: http://www.ncbi.nlm.nih.gov/gene
Acvr2a | Appe | App | Ar | Bdnf | Camkk2 | Cb1 | Dbx1 |
doublesex | Eifzs3x | Er | fru | Gabra | Htr1b | M1 | M2 | Mc1r |
Sert | Smcx | SRP20 | SRY | Sxl | Th | tra | Trp2 | Ube1x | Usp9x | Utx
| V1ar | XIST

OMIM: http://www.ncbi.nlm.nih.gov/omim Alzheimer's disease | Huntington's disease UniProtKB: http://www.uniprot.org OR67D | PSE | TRA2

FURTHER INFORMATION

Elena Jazin's homepage: http://www.fu.uu.se/devbiol/lazin/jazin_index.html

Larry Cahill's homepage: http://cahill.bio.uci.edu/ Ensemble for Homo sapiens: http://www.ensembl.org/ Homo_sapiens/Info/Index

EREs in human and mouse: http://www.mapageweb. umontreal.ca/maders/eredatabase/index.html Nurebase: http://www.ens-lyon.fr/LBMC/laudet/nurebase/ nurebase html

Sebida: http://www.sebida.de

ALL LINKS ARE ACTIVE IN THE ONLINE PDF