

# BI 8 LECTURE 18

## OTHER RNAS AS REGULATORS AND RNAS AS ENZYMES

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3 March 2016

Read: Alberts et al.: Chapter 6, pp. 317-324, 346-347, 362-366

# RNA is more than a protein coding molecule

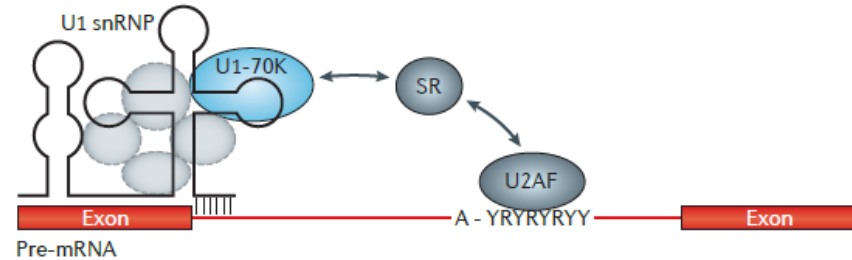
- RNA as regulator of other mRNAs' translation and lifespans (miRNA and artificial versions)
- RNA as a local orientation device to recruit chromatin modifier function to specific sites (supplementing action of transcription factors)
- RNA as an enzyme

# Multiple regulatory modes exist for RNA molecules

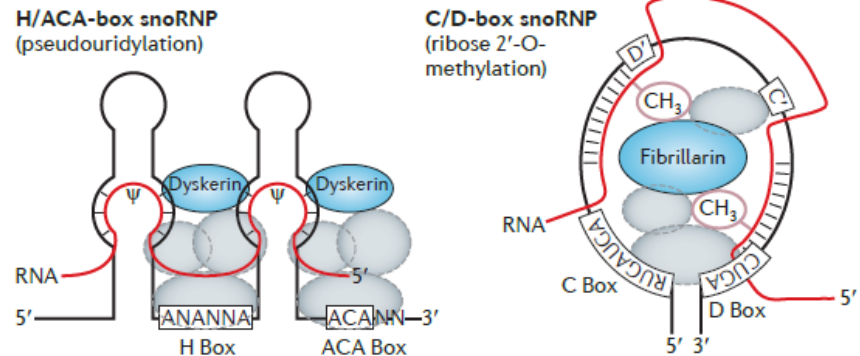
Guidance for protein-  
based enzymes:  
using RNA  
complementarity for  
target specificity

## Box 2 | Examples of ncRNAs with guiding activity for proteins

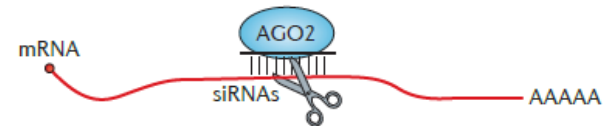
### a Pre-mRNA splicing



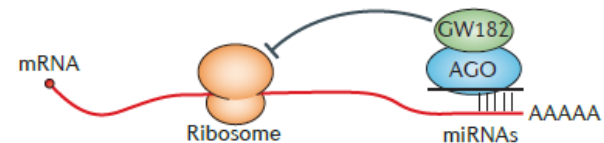
### b Site-specific RNA modification



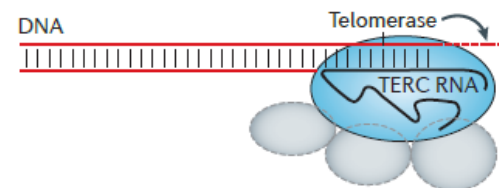
### c RNA interference



### miRNP-mediated silencing

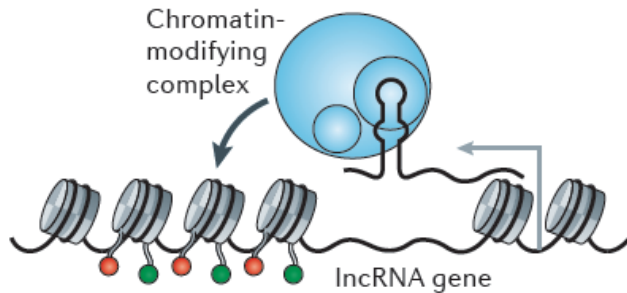


### d Telomere formation



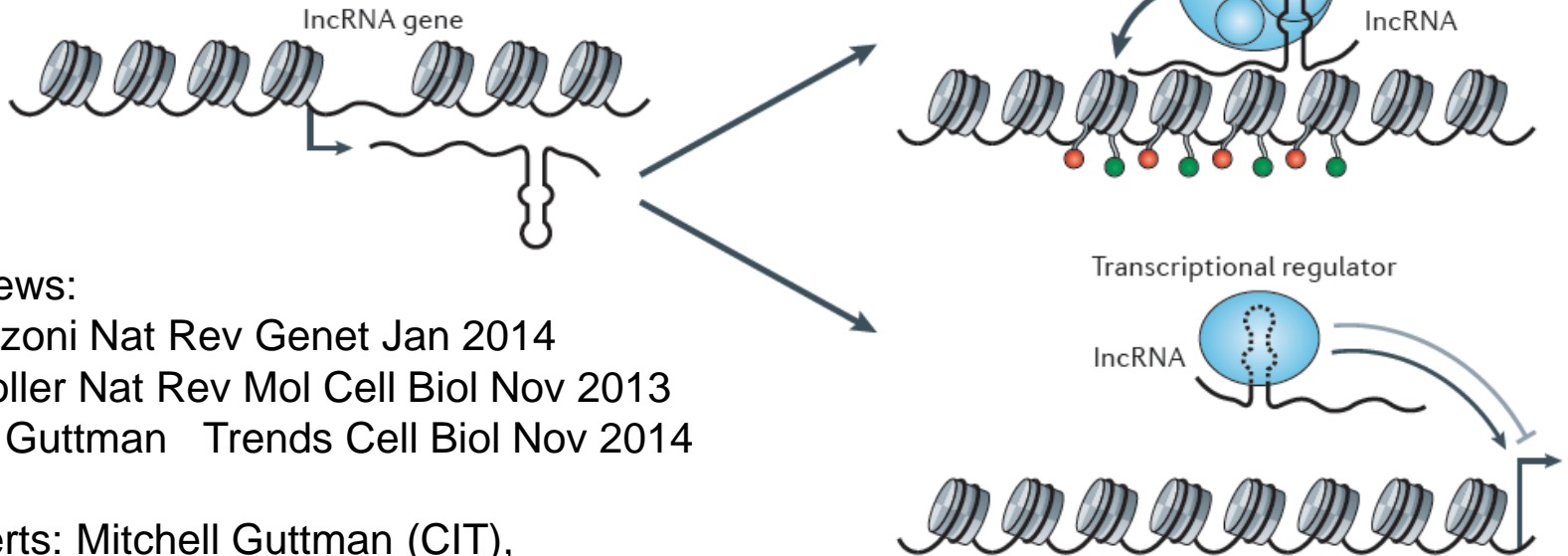
# Long noncoding RNAs: from the dark matter of the genome

## A *Cis*-acting lncRNAs



A variety of roles in guidance of chromatin modifiers, still being discovered, and other roles

## B *Trans*-acting lncRNAs



Some reviews:

Fatica, Bozzoni Nat Rev Genet Jan 2014

Geisler, Coller Nat Rev Mol Cell Biol Nov 2013

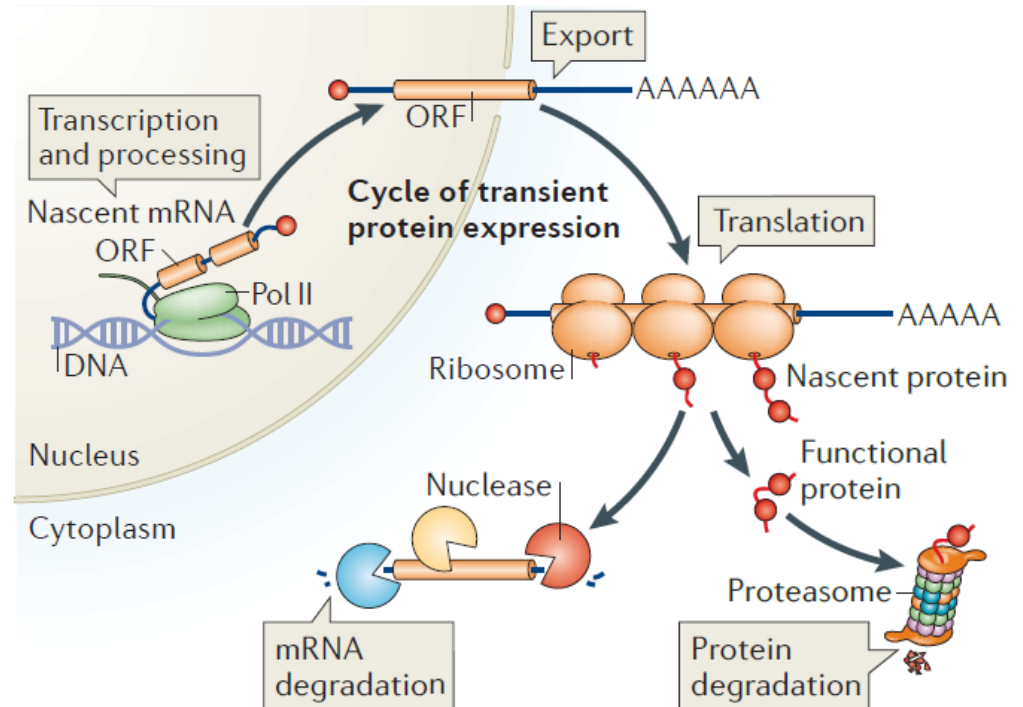
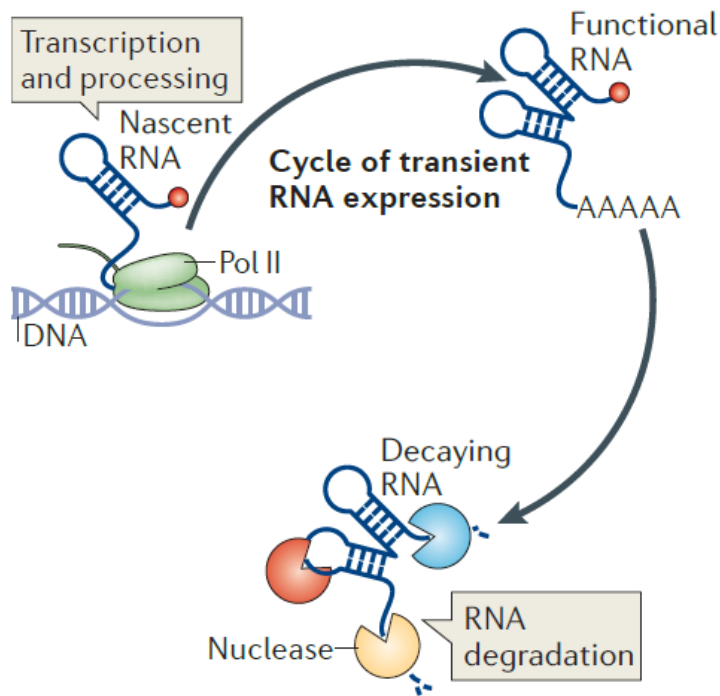
Quinodoz, Guttman Trends Cell Biol Nov 2014

Local experts: Mitchell Guttman (CIT),  
Howard Chang (Stanford)

# Regulatory RNA as a scaffold

- Long non-coding RNAs probably use base pairing to interact with other nascent RNAs
- Long non-coding RNAs use secondary structure to recruit multiple proteins
- Often long non-coding RNAs recruit chromatin modifying enzymes, including histone deacetylases (repression)
- Local binding, local action, maybe helping to organize domains of different gene regulation activities in cell nucleus

# Functional noncoding RNA: synthesis & turnover kinetics make it a quick-change artist

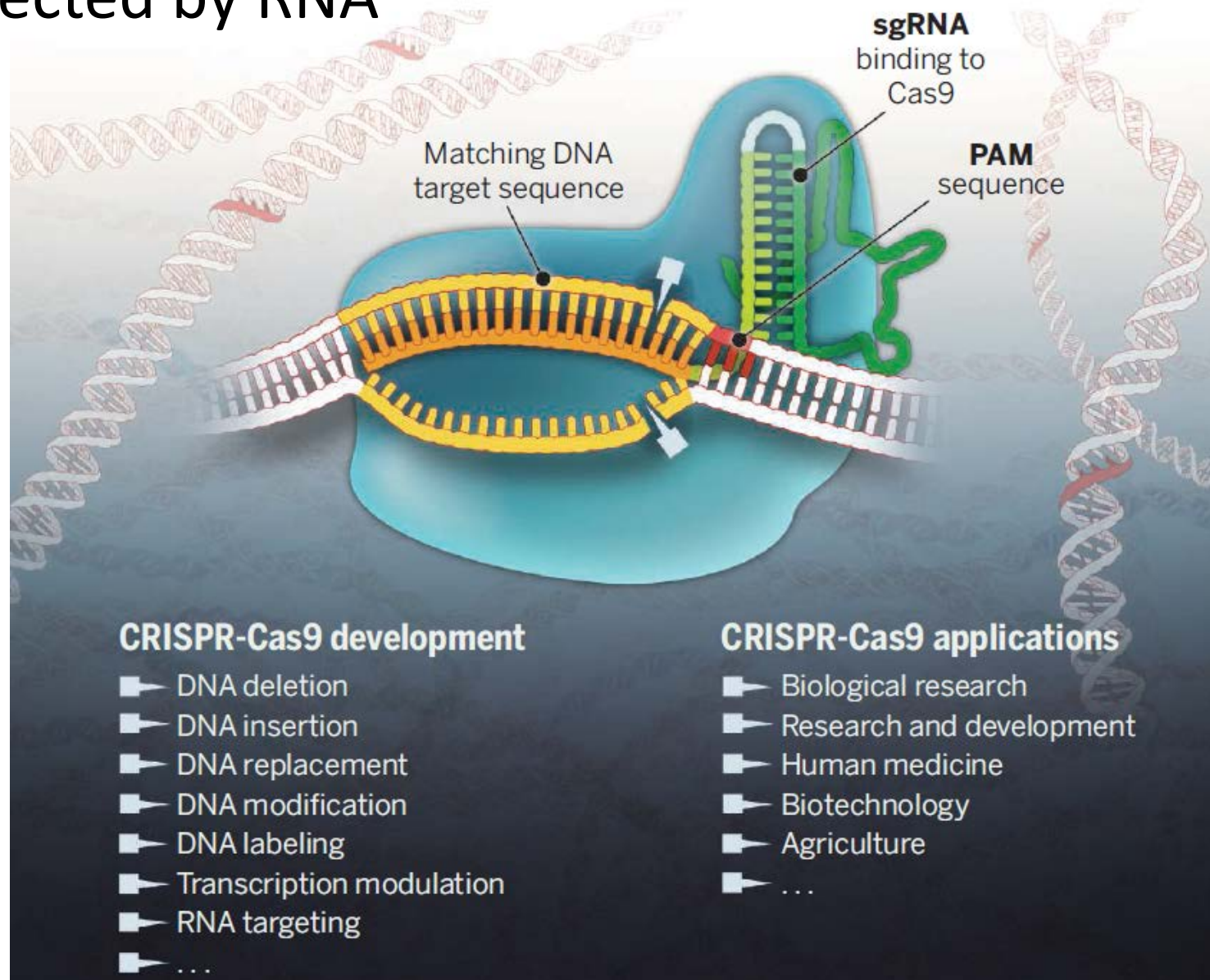


Many extra steps needed to make, destroy protein

# Ways that RNA contributes to enzymatic activity

- RNA as guide, by base pairing with substrate
- RNA as catalyst
  - stabilizing transition state &/or
  - creating it through attack on substrate
- These are primary roles of multiple classes of RNAs

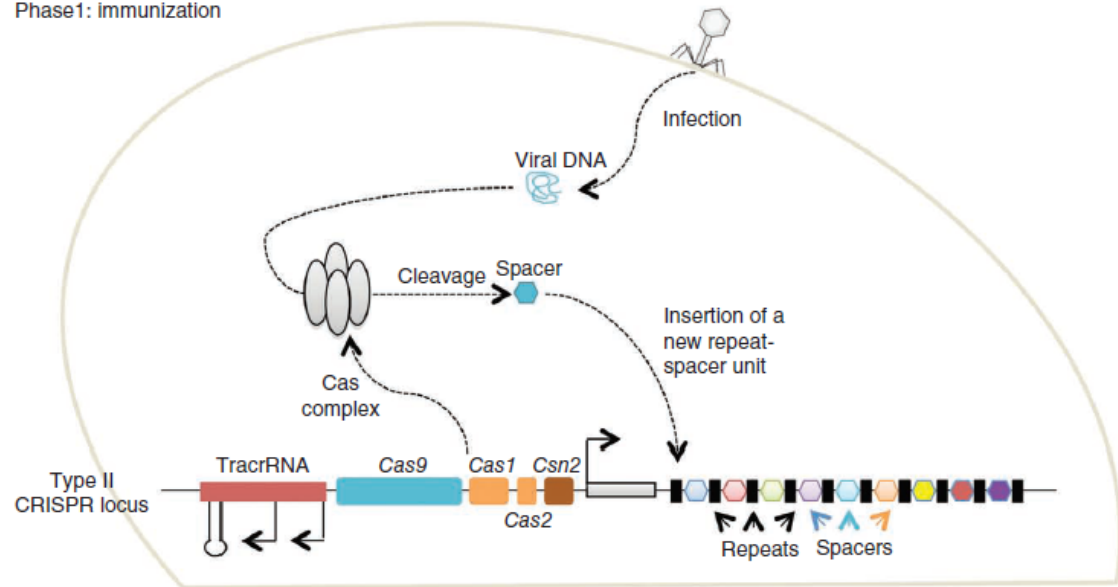
# CRISPr—Cas9: a revolution in genome editing, directed by RNA



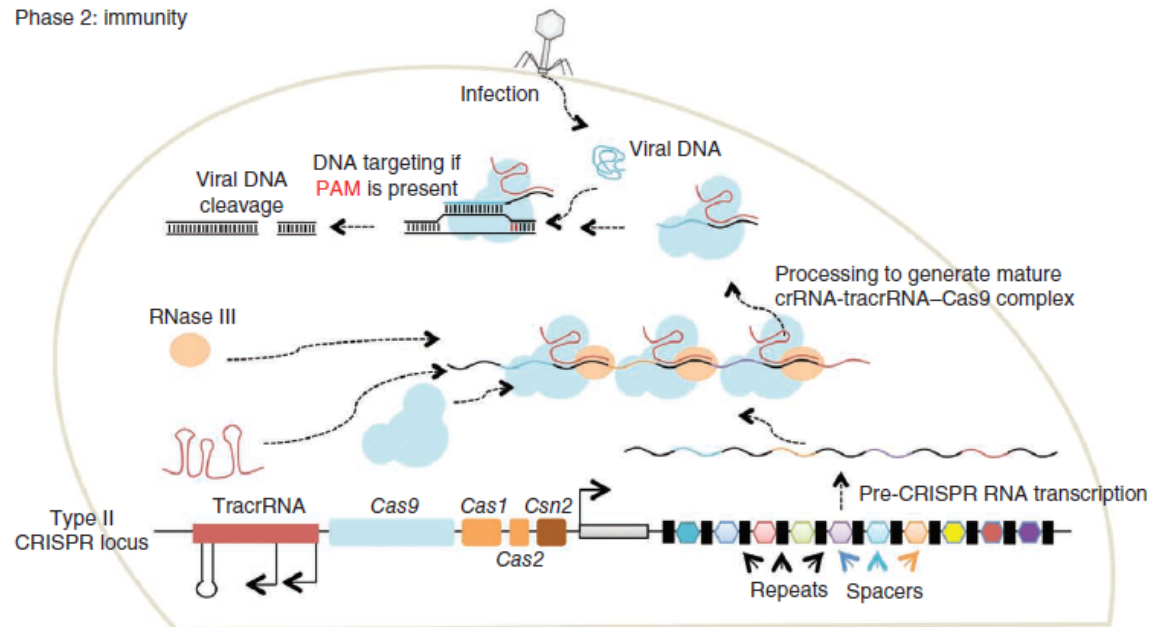


# Evolution of a defense system: know your enemy, really really well

Phase 1: Immunization



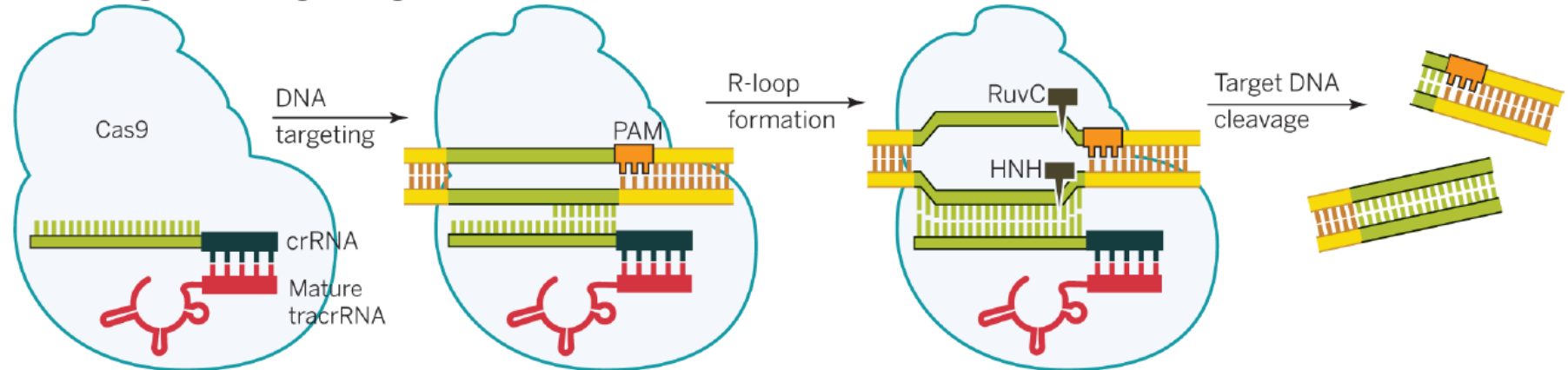
Phase 2: immunity



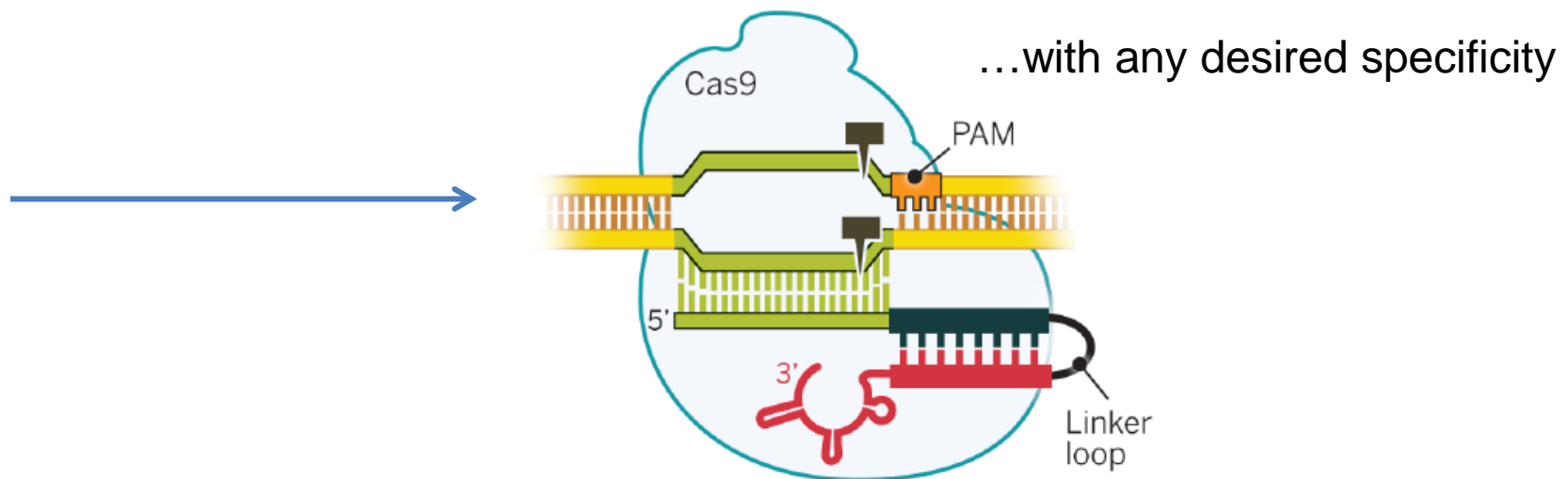
Invading phage DNA is integrated into an array of DNA repeats: a common part of a template for RNA synthesis that will be used to guide Cas9 nuclease vs. future infections!

# Tracer RNA replaced by “Guide RNA”: a bacterial mechanism for “immunity” can be made into a tool

## C RNA-guided cleavage of target DNA

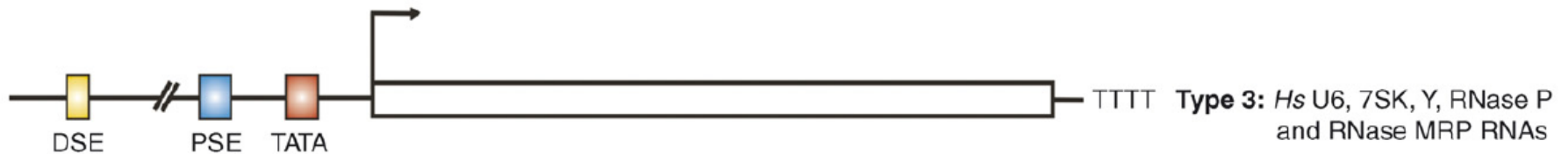
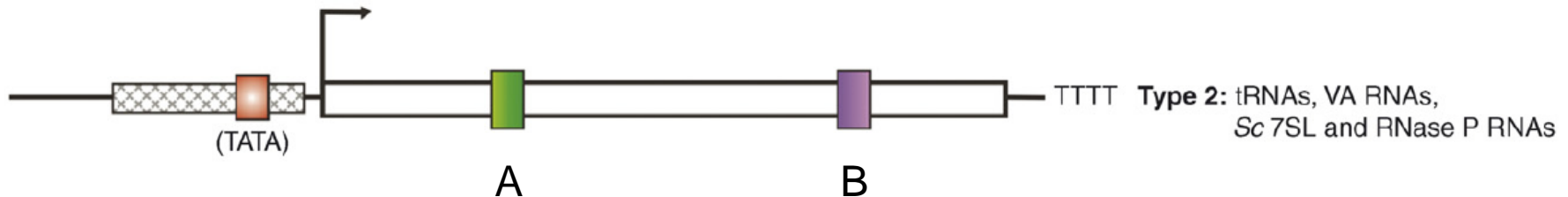
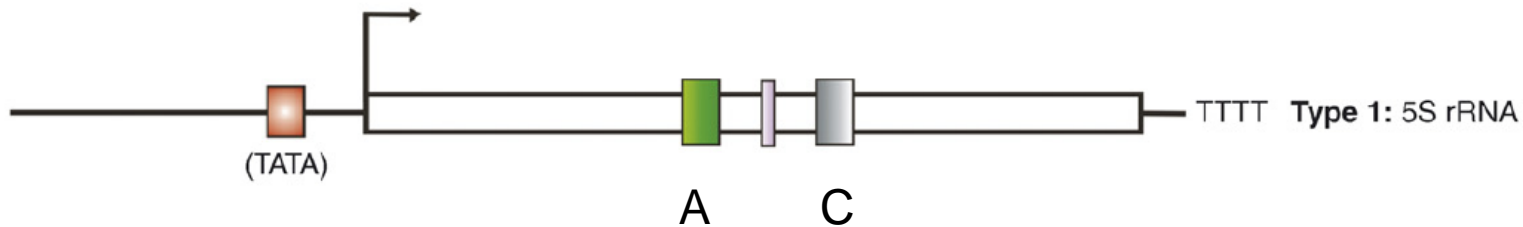


Cas9 programmed by  
single guide RNA



# Many catalytic-like small RNAs are synthesized by RNA polymerase III rather than I or II

(contrast with miRNAs, synthesized by RNA pol II)

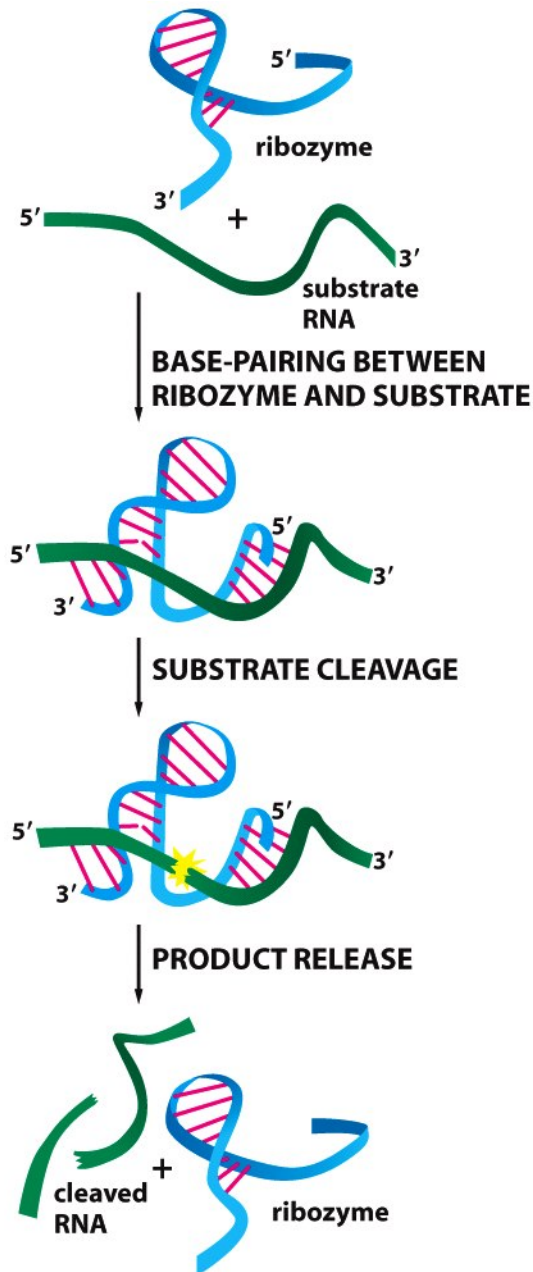


Colored boxes: special cis-regulatory elements for pol III; also oligoT termination signals

(Dieci, ....& Pagano, Trends Genet., 2007)

The benefits of  
2'-OH reactivity:

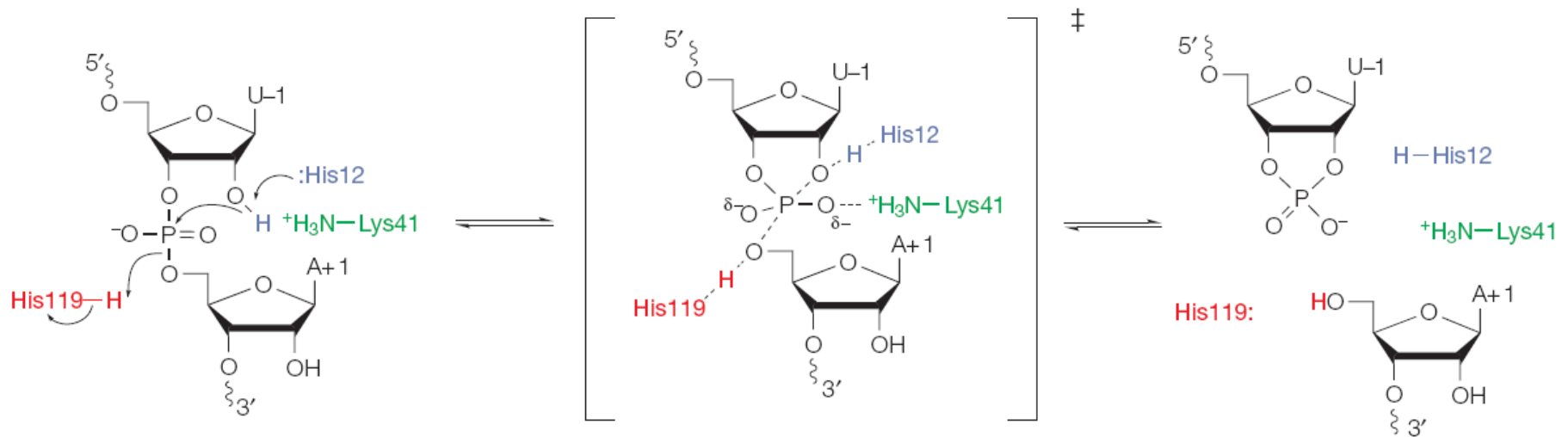
RNA can also  
catalyze  
enzymatic  
reactions, not  
just find  
templates



**Table 6–5 Some Biochemical Reactions That Can Be Catalyzed by Ribozymes**

ACTIVITY	RIBOZYMES
Peptide bond formation in protein synthesis	ribosomal RNA
RNA cleavage, RNA ligation	self-splicing RNAs; RNase P; also <i>in vitro</i> selected RNA
DNA cleavage	self-splicing RNAs
RNA splicing	self-splicing RNAs, perhaps RNAs of the spliceosome
RNA polymerization	<i>in vitro</i> selected RNA
RNA and DNA phosphorylation	<i>in vitro</i> selected RNA
RNA aminoacylation	<i>in vitro</i> selected RNA
RNA alkylation	<i>in vitro</i> selected RNA
Amide bond formation	<i>in vitro</i> selected RNA
Glycosidic bond formation	<i>in vitro</i> selected RNA
Oxidation/reduction reactions	<i>in vitro</i> selected RNA
Carbon–carbon bond formation	<i>in vitro</i> selected RNA
Phosphoamide bond formation	<i>in vitro</i> selected RNA
Disulfide exchange	<i>in vitro</i> selected RNA

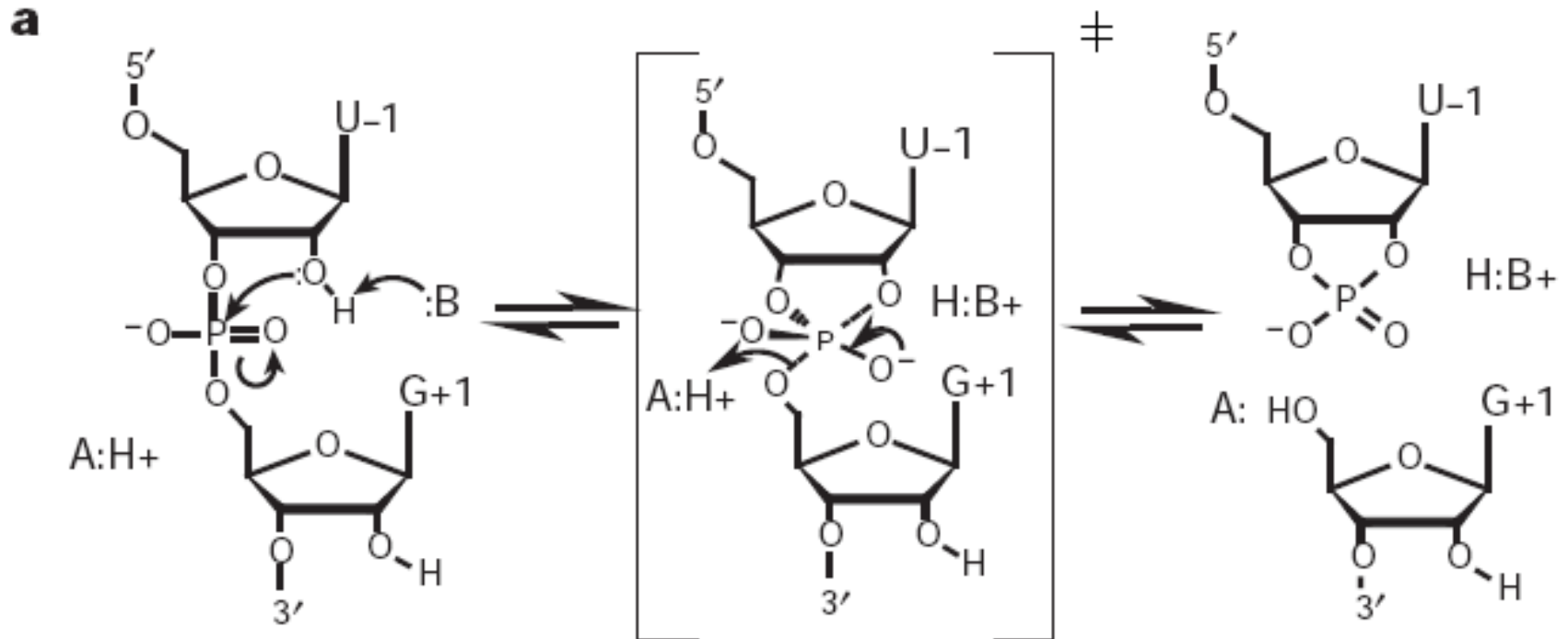
# How a protein catalyzes RNA cleavage (RNase A)



Concerted acid-base catalysis: (1) His12 abstracts proton to create oxyanion for attack; (2) protonated His 12, unprotonated His119 and protonated Lys41 stabilize the transition state; (3) cleaved polynucleotide is released; later (not shown) His12 “gives back”

(Fedor, Williamson Nat Rev Mol Cell Bio 2005)

# RNA self-cleavage: the easiest job for an RNA



Extremely similar to RNase cleavage... especially if acid and base moieties are there for transition state stabilization

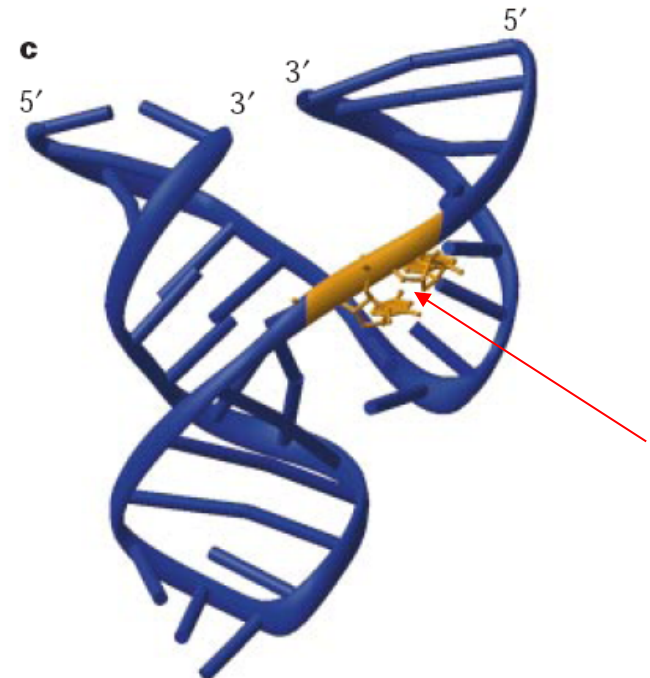
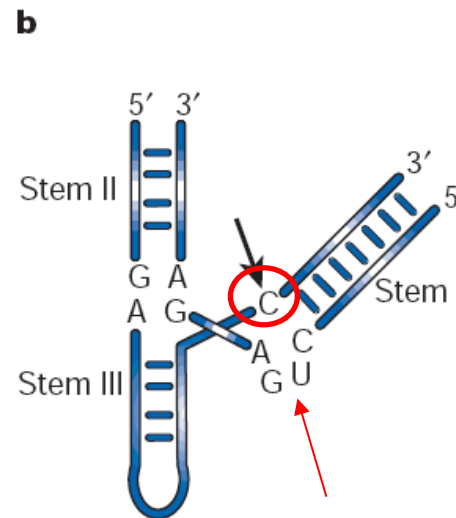
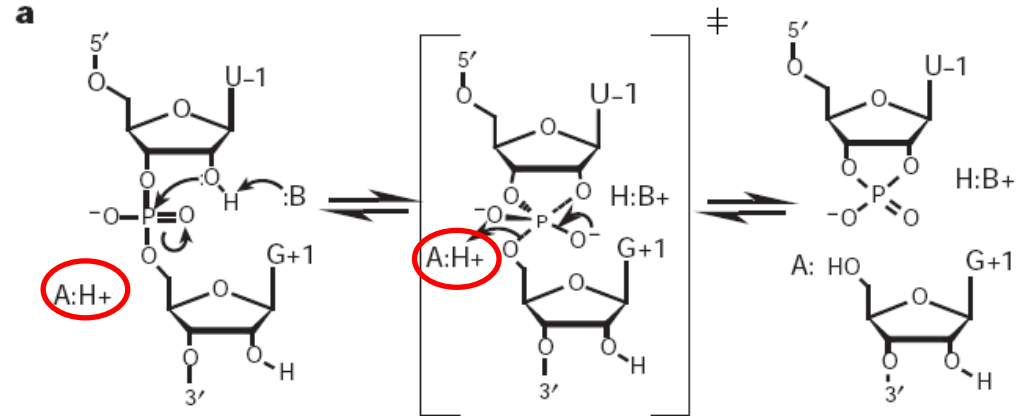
(Doudna, Cech *Nature* 2002)

∴ STRUCTURE CAN HELP!



# “Hammerhead” ribozyme: a naturally occurring RNA enzyme that *catalyzes* its own cleavage

Highly structured “hammerhead” motif allows an RNA plant virus to clip out genome-length RNA pieces from a long concatenated replication product



(Doudna, Cech *Nature* 2002)



RNA-dependent catalysis can also stabilize transition states by *complementary base pairing*

This plays a huge role in splicing: every eukaryote's need for precise RNA cleavage

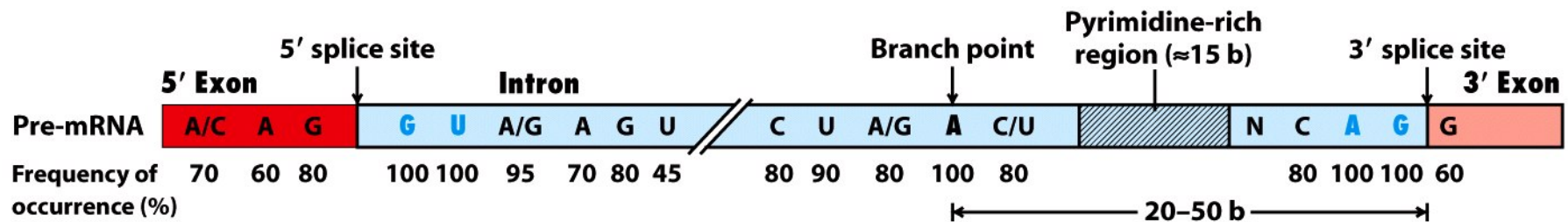


Figure 8-7  
*Molecular Cell Biology, Sixth Edition*  
© 2008 W. H. Freeman and Company

not completely deterministic, but guided by key sequences in substrate RNA... These are targets for base pairing

# Reminder of what happens during splicing

Chemistry is permissive  
but a big problem to find  
the right phosphodiester  
bonds to be attacked

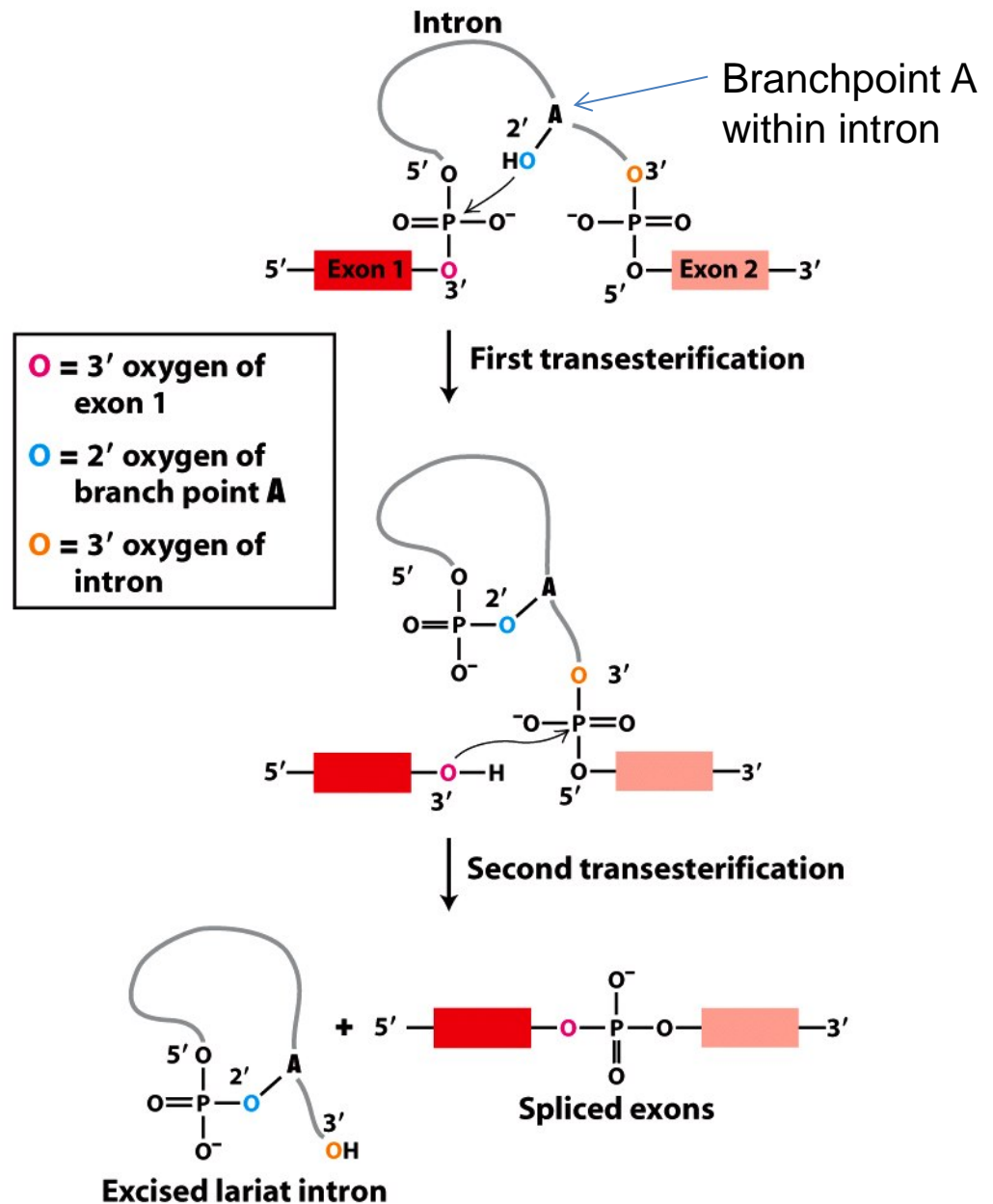
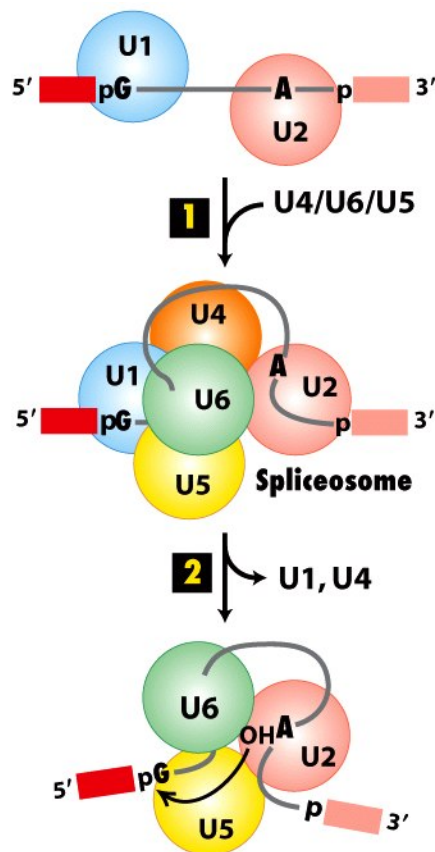
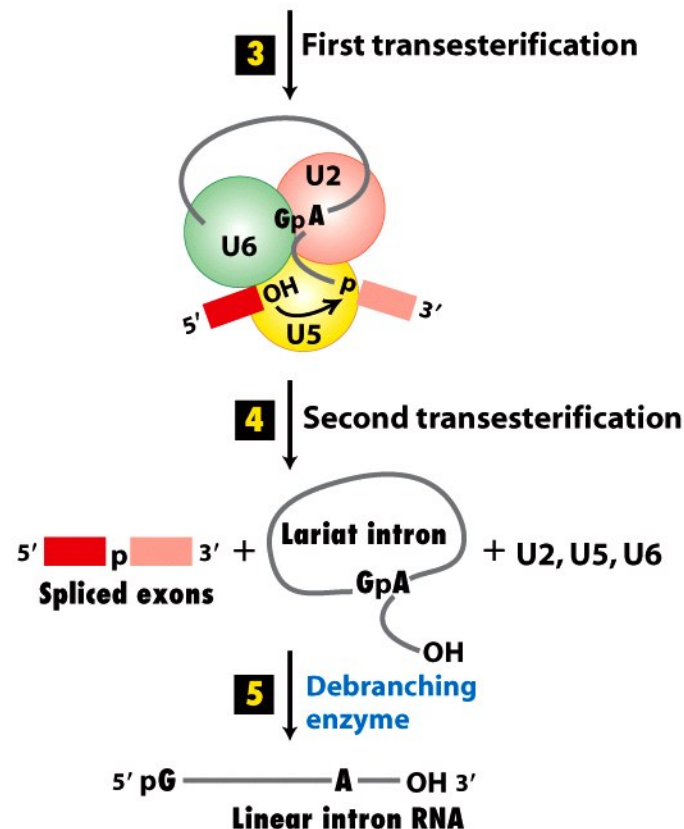


Figure 8-8  
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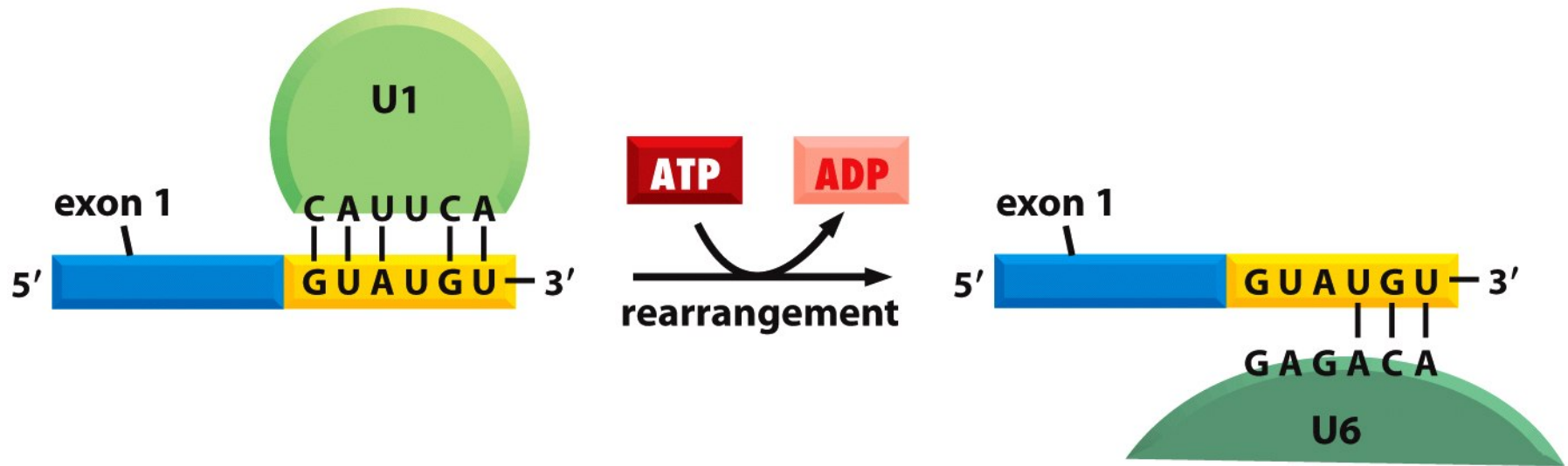
Splicing normally gets a lot of help  
from ribonucleoprotein  
“spliceosomes”:  
key players, snRNA-Protein complexes



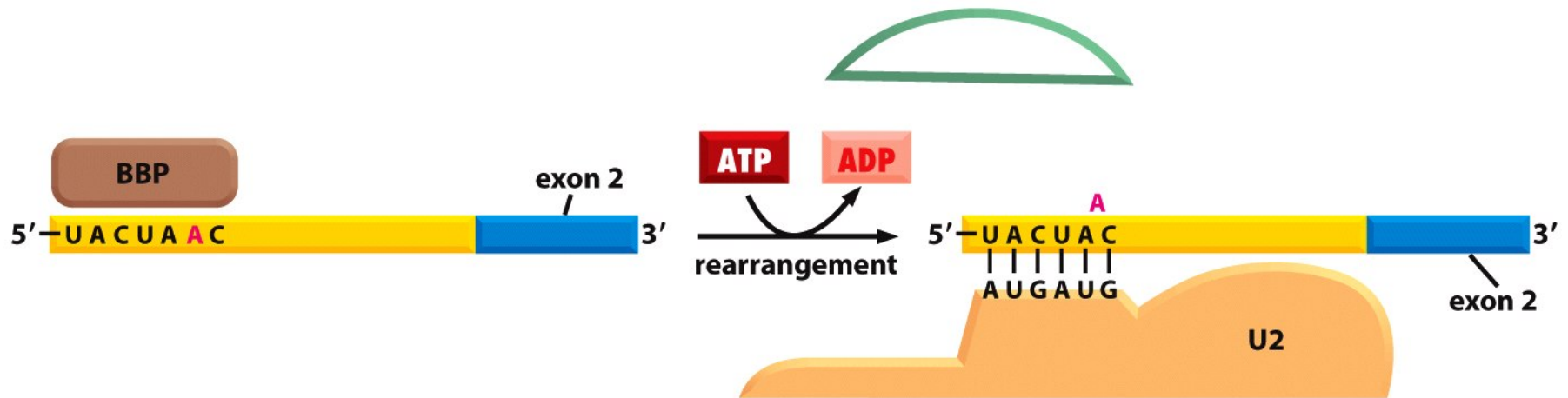
snRNP: small nuclear  
ribonucleoproteins,  
complexes centered  
on snRNAs

Main snRNPs:  
U1: 5' for assembly  
U2: at 3' branch site  
U5: for second attack  
U6: 5' for all catalytic  
events

mRNA intron is base paired with U1, U6 snRNAs:  
handoff requires ATP hydrolysis

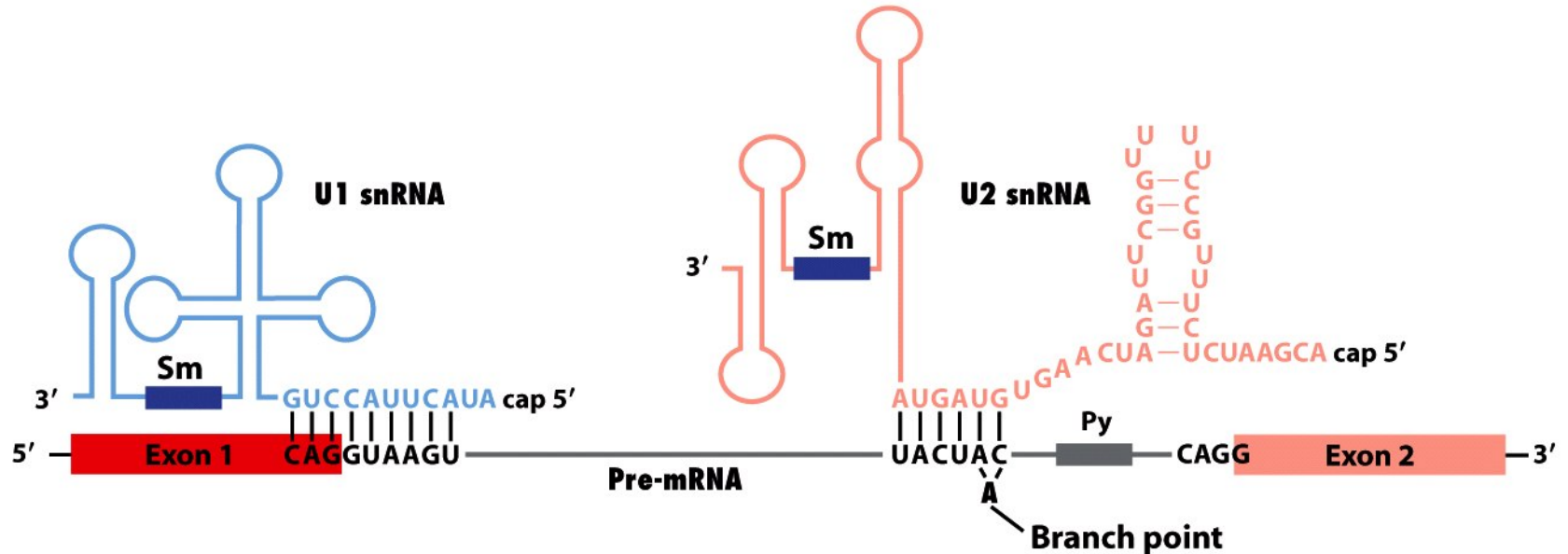


Displacement of original binding protein for 3' end of intron by U2... also requires ATP hydrolysis



# Base pairing is important for splicing efficiency

(a)

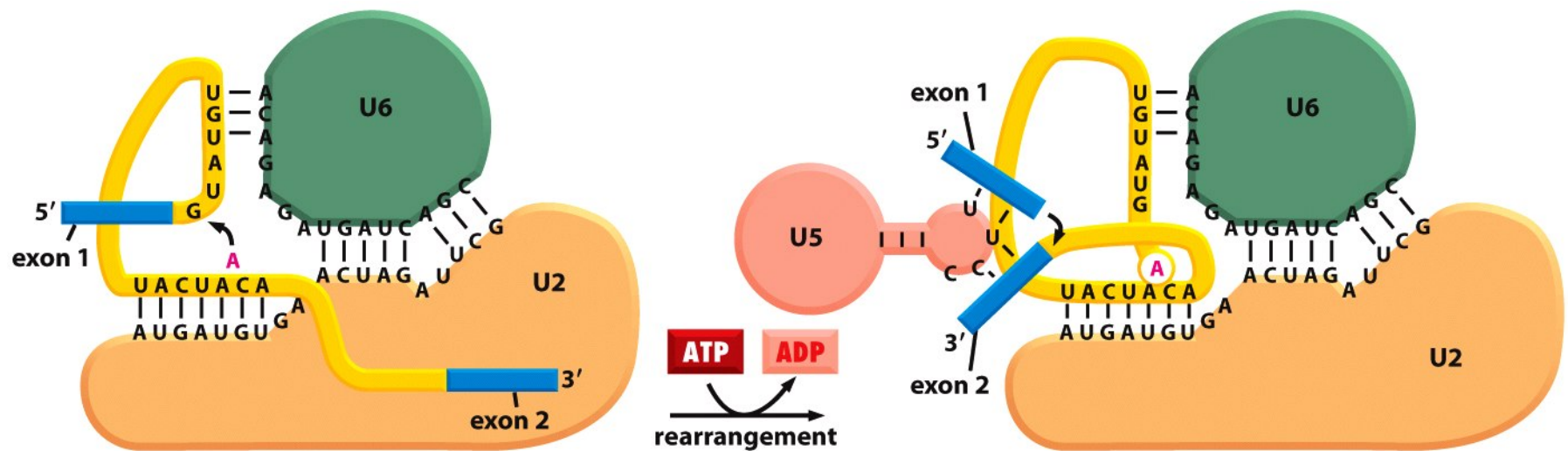


(b)



Figure 8-9  
Molecular Cell Biology, Sixth Edition  
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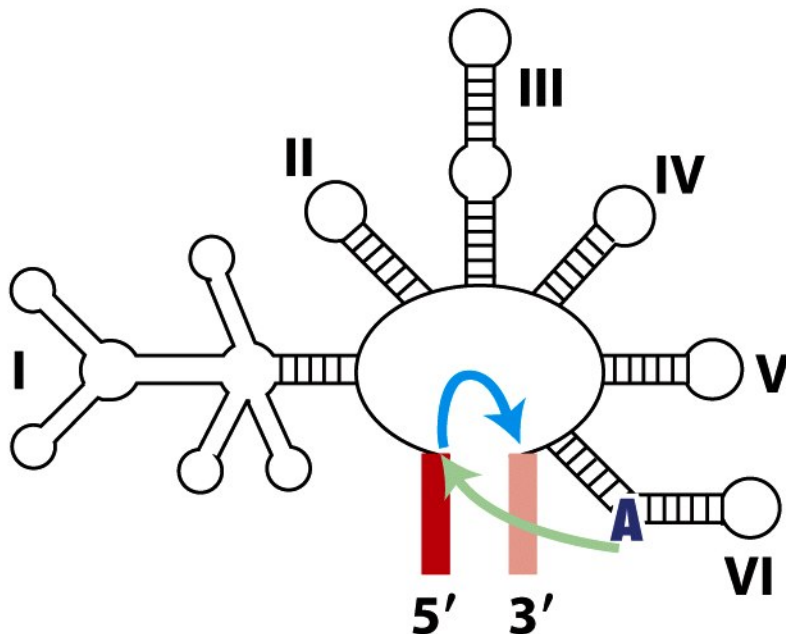
U6/U2 machine carries out the splicing with help from U5 base pairing and more ATP hydrolysis



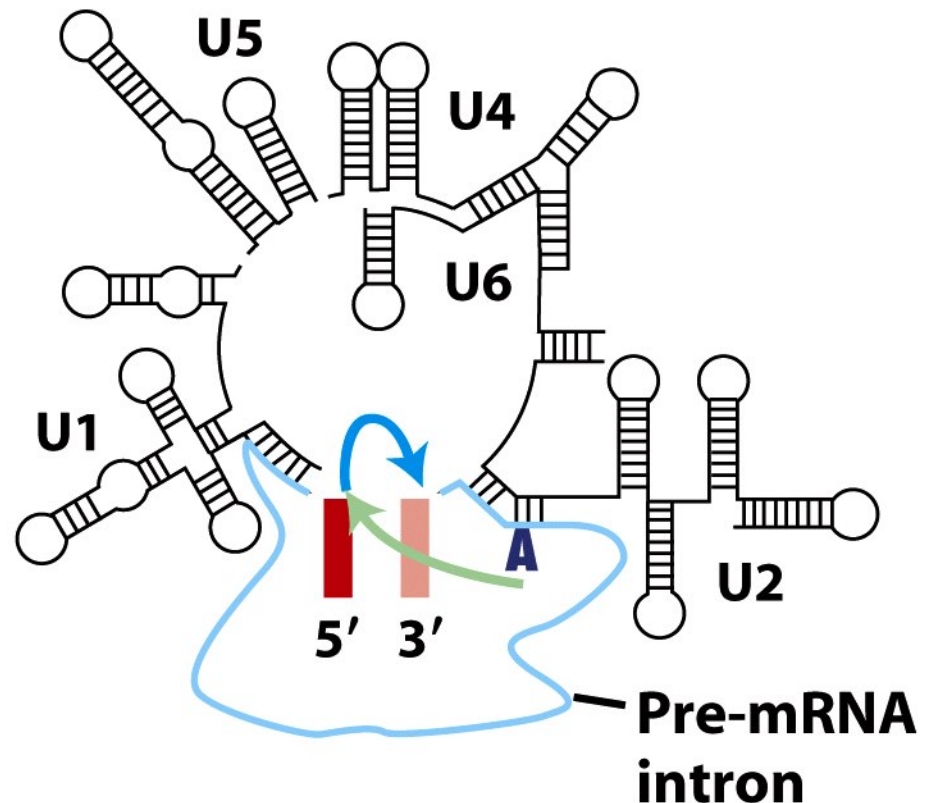


There are small introns in some microorganisms with such a highly specialized structure that they can splice out themselves....Possibly analogous folding to “spliceosomes”

**(a) Group II intron**

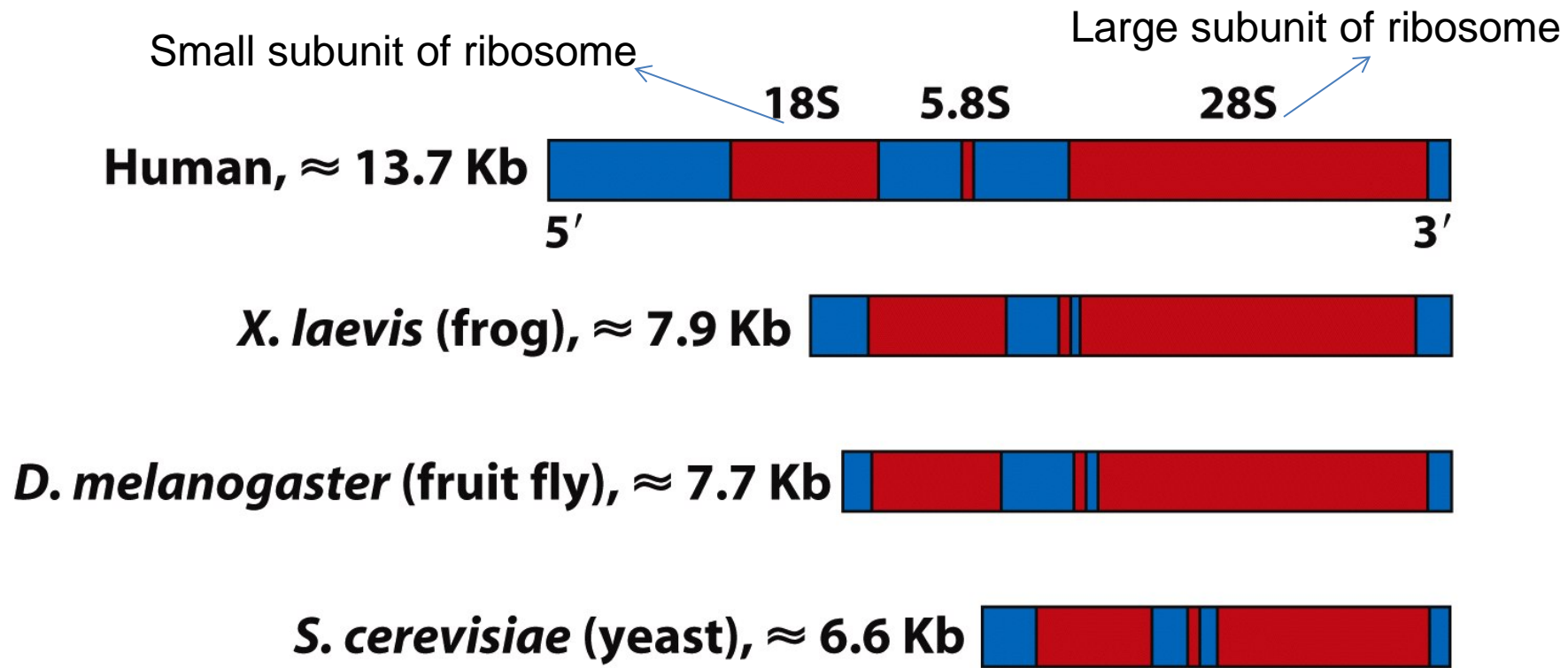


**(b) U snRNAs in spliceosome**





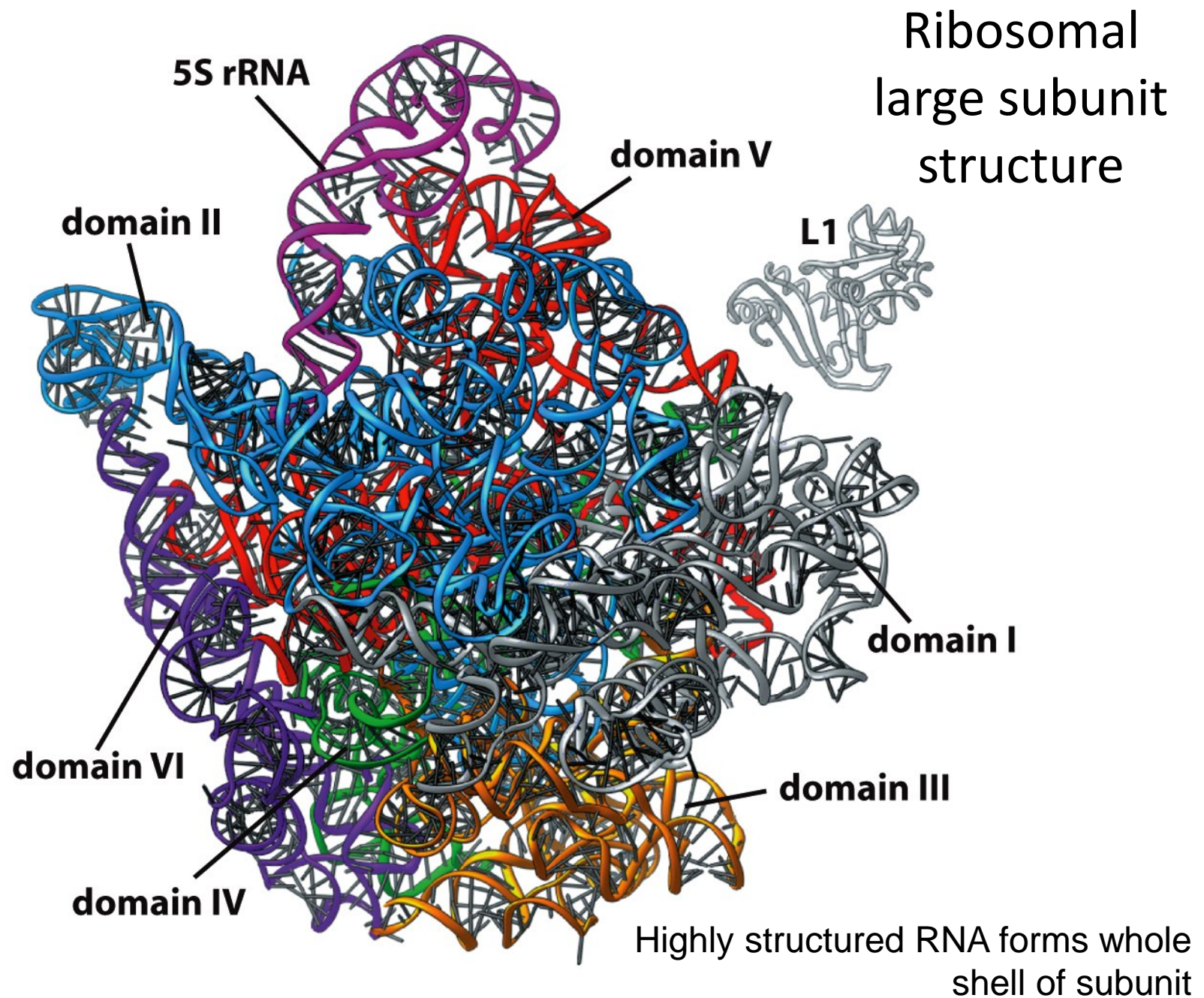
Translation in the ribosome itself may also depend on ribozyme-like activity of rRNAs



Ribosomal RNA  
primary transcripts are  
processed by cleavage  
into the separate  
rRNAs

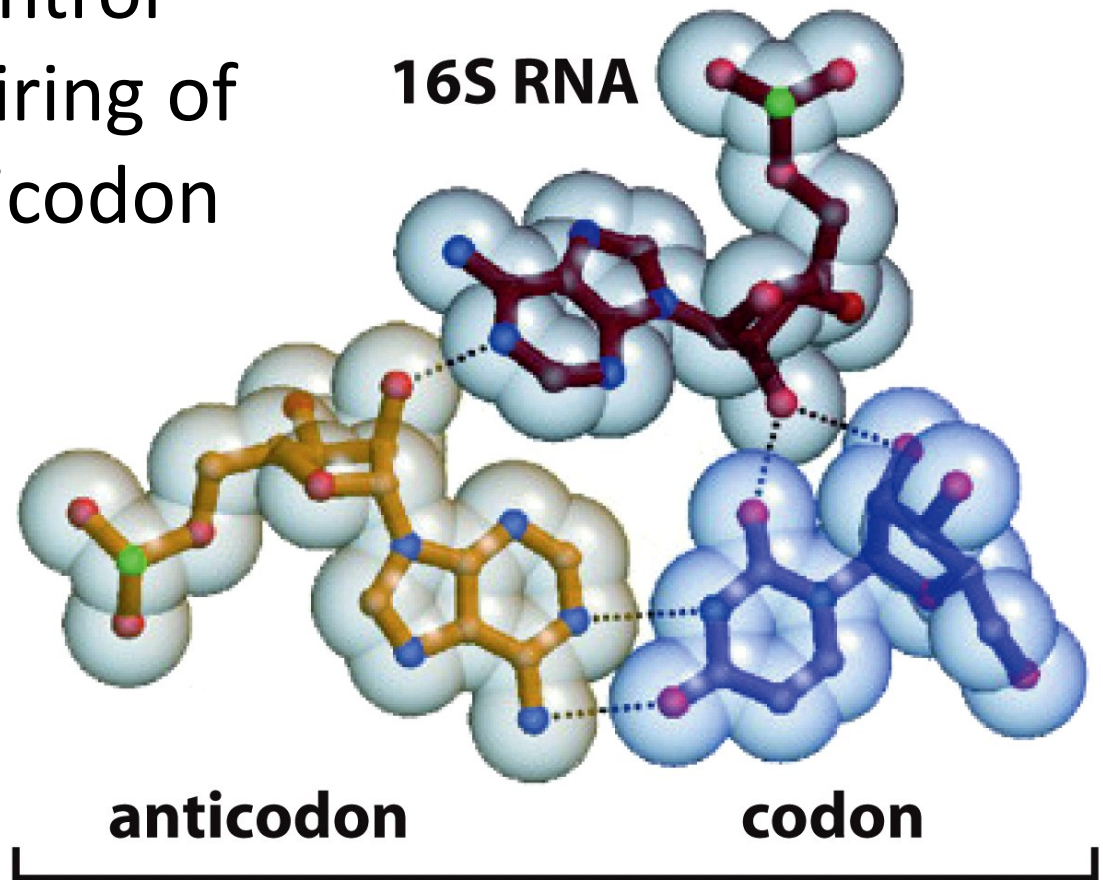
Figure 8-34  
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18S:  $\sim 1.9$  kb      28S:  $\sim 4.7$  kb



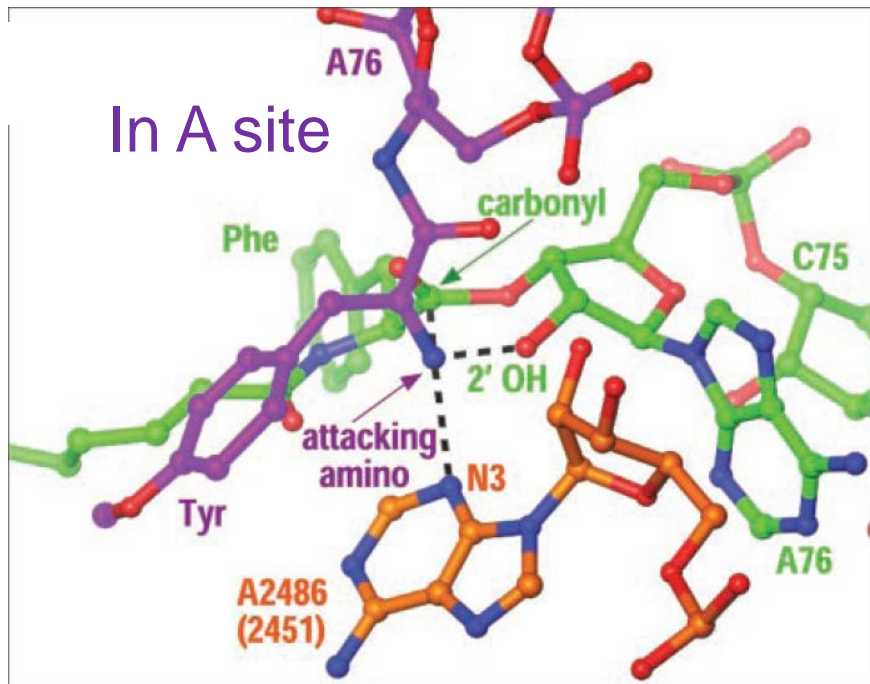
Within ribosome, complementarity to 16S (small rRNA) can define translational start site (Shine Dalgarno box in bacteria), but also...

acts as quality control  
for alignment/pairing of  
codon/ tRNA anticodon  
at each step of  
translation





And a ribozyme-like catalyst lies at the heart of the large ribosomal subunit... crucial for peptidyl transferase reaction



In P site

Large rRNA

(Hansen, ... Steitz  
PNAS 2002;  
Fedor, Williamson  
Nat Rev Mol Cell  
Bio 2005)

# RNA as the jack of all trades of molecular biology

- Base pairing with other molecules for recognition and complex assembly
- Secondary structure for unique features
- Biochemical reactivity for enzyme activity
- Templating possible for replication

# RNA world first?

- Versatility: genetic code and active agent
- Ease of synthesis and turnover, for flexibility
- Inheritance at heart of some of most crucial functions
- Ability of isolated RNAs to do many jobs, including self replication