

Digital Images

Bi/BE177: Principles of Modern Microscopy



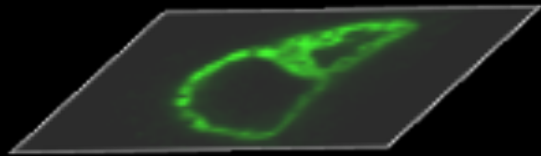
Vikas Trivedi

(viktri@caltech.edu)

California Institute of Technology



Outline



Digital image formation

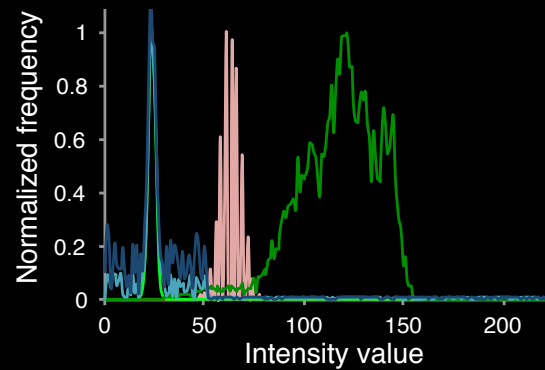


Image properties

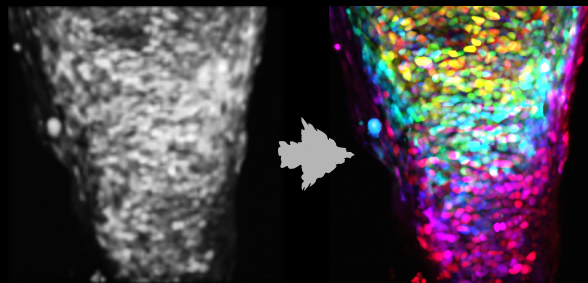


Image processing

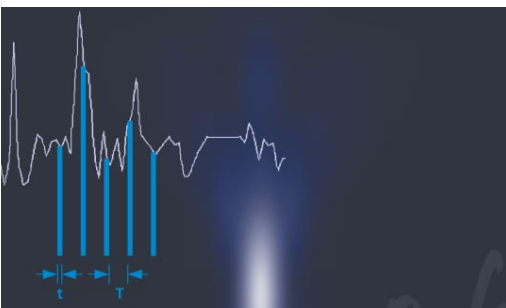


Available softwares

Resources

Principles



Confocal Laser Scanning Microscopy



axial $\Delta z = \frac{\lambda}{2 \text{NA}^2}$

Optical Image Formation

Electronic Signal Processing



Carl Zeiss Microscopy Online Campus

zeiss-campus.magnet.fsu.edu/index.html

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Fluorescent Proteins
Microscope Light Sources
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Microscope Light Sources
Fluorescent Proteins
Live-Cell Imaging
Optical Sectioning
Superresolution

A Whole World of Microscopy Knowledge

Exclusively from Carl Zeiss and MOLECULAR EXPRESSIONS™

Welcome

The Carl Zeiss Microscopy Online Campus website explores the fascinating world of optical microscopy and digital imaging.

Imaging Solutions

Nikon MicroscopyU

www.microscopyu.com

INSTRUMENTS NIKON EU NIKONUSA NIKON.COM

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- Live-Cell Imaging
- Fluorescence Microscopy
- Optical Systems
- Phase Contrast
- DIC Microscopy
- Confocal Microscopy
- Superresolution Microscopy
- Stereomicroscopy
- Polarized Light Microscopy
- Cell Motility Video Gallery
- Swept Field Video Gallery
- FRET Pair Combinations

Introduction to DIC Microscopy



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Small World Competition

- Enter the 2015 Contests
- 2015 Small World
- Competition Rules
- Competition Prizes

Olympus Microscopy Resource Center

www.olympusmicro.com/index.html

Olympus America Research Imaging Software Confocal Clinical FAQ's

MICROSCOPY RESOURCE CENTER

OLYMPUS

SITE NAVIGATION

- Home Page
- Interactive Tutorials
- Microscopy Primer
- Physics of Light & Color
- Microscopy Basic Concepts
- Special Techniques
- Fluorescence Microscopy
- Confocal Microscopy

Olympus BioScapes Contest

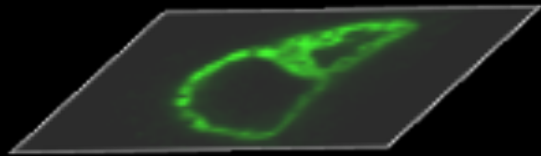
Fluorescent Proteins Intro

Physics of Light and Color
Total Internal Reflection
Fundamentals of FRET
Confocal Microscopy
Microscope Basics



A broad range of fluorescent protein genetic variants have been developed over the past several years that feature fluorescence emission spectral profiles spanning almost the entire visible light spectrum.

Outline



Digital image formation

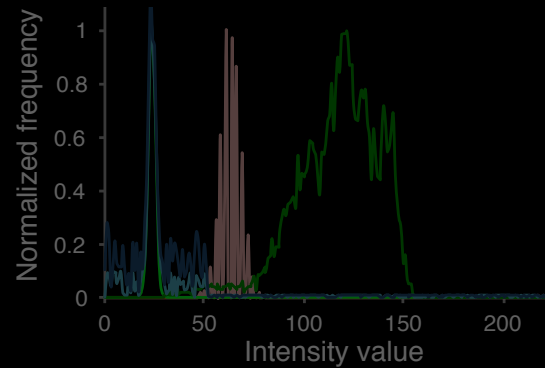


Image properties

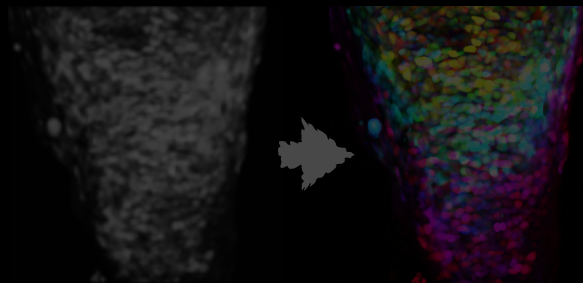


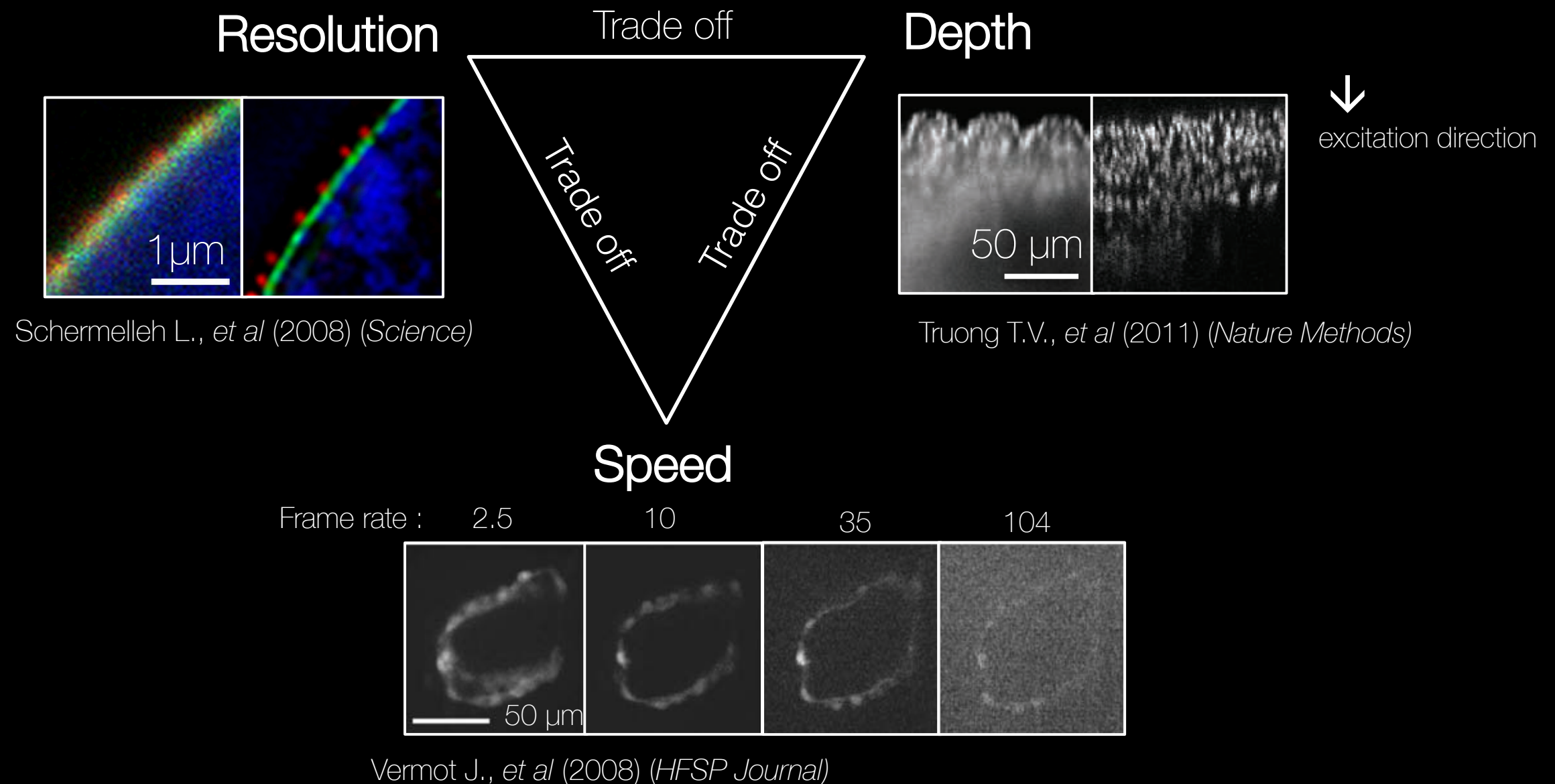
Image processing



Available softwares

Microscopy as a compromise

Competition between different performance parameters



Microscopy in biological context

Competition between different performance parameters

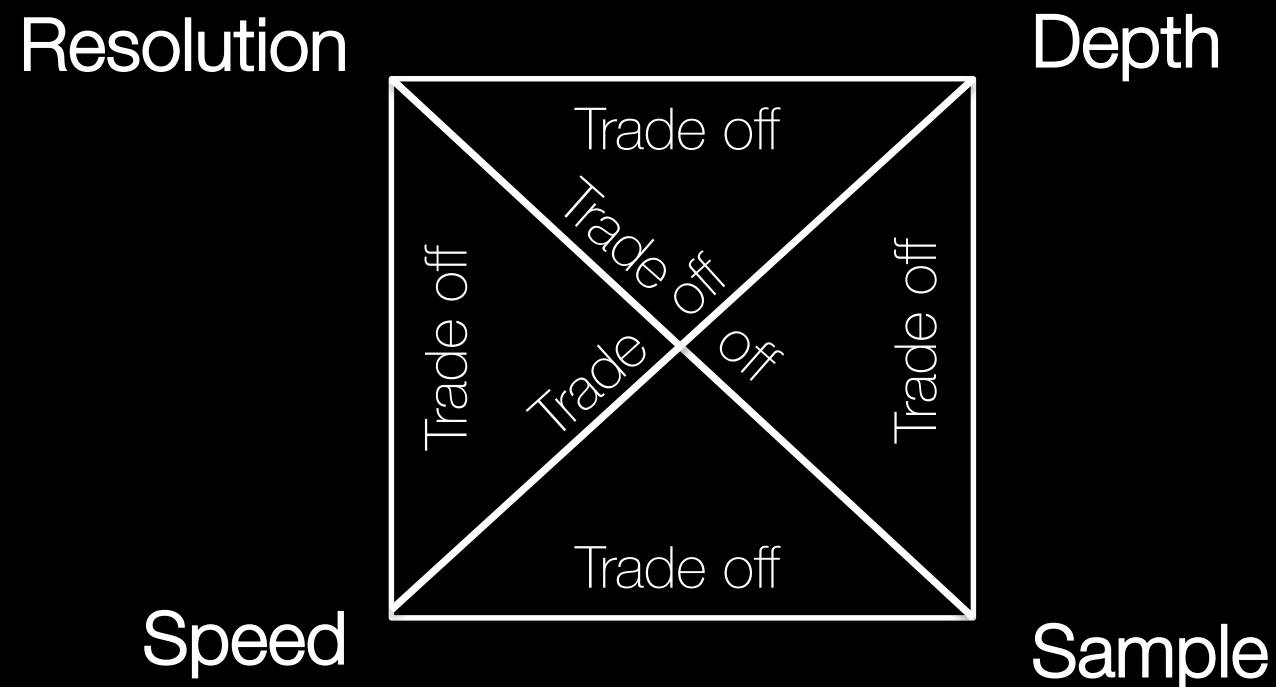
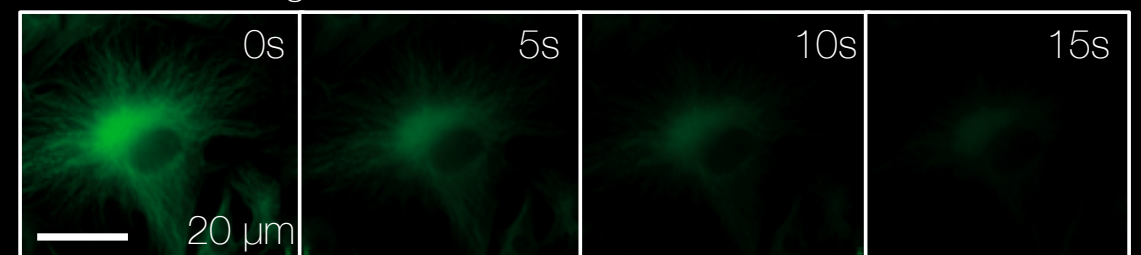


Photo-bleaching



Life technologies, Invitrogen

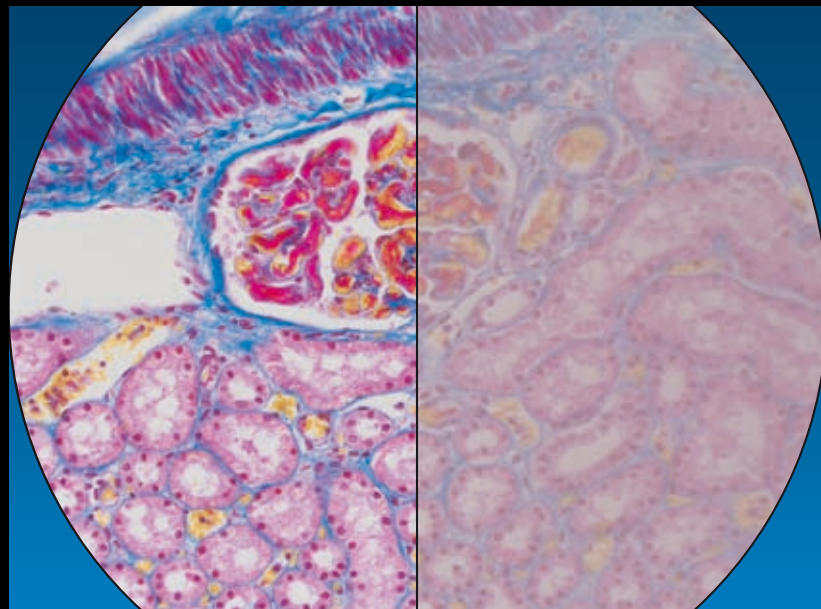
Photo-toxicity



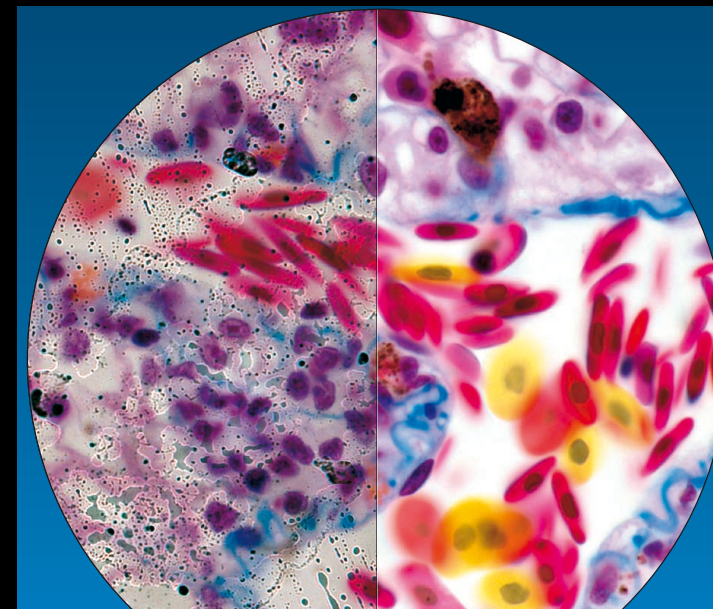
Always best to have the best image settings

-better to optimize acquisition than rely on post-acquisition processing

Something really simple :-)

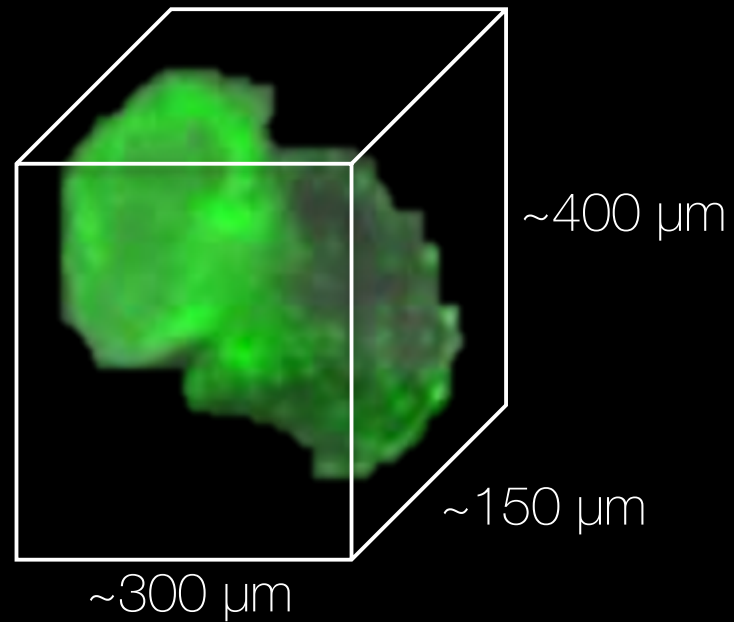
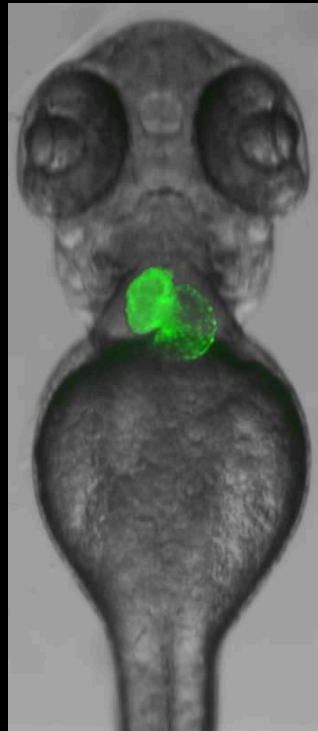


Soiled Lens

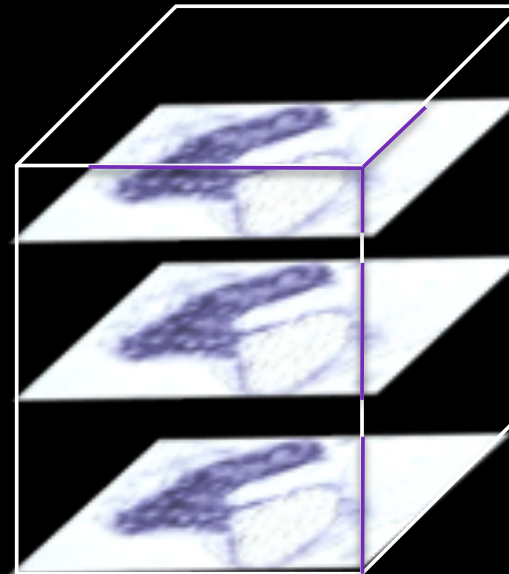


Soiled Camera and dirt

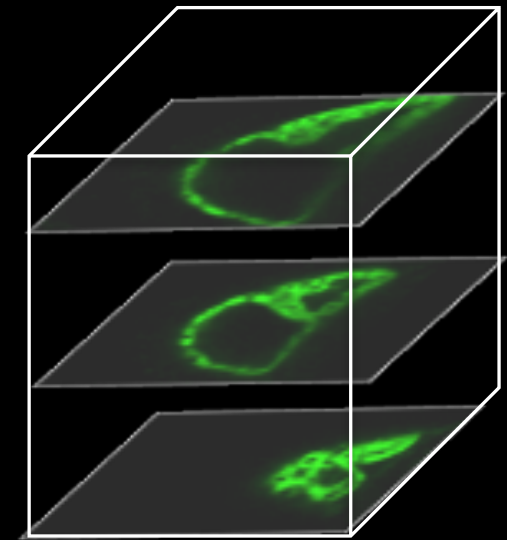
Building up a 3D image



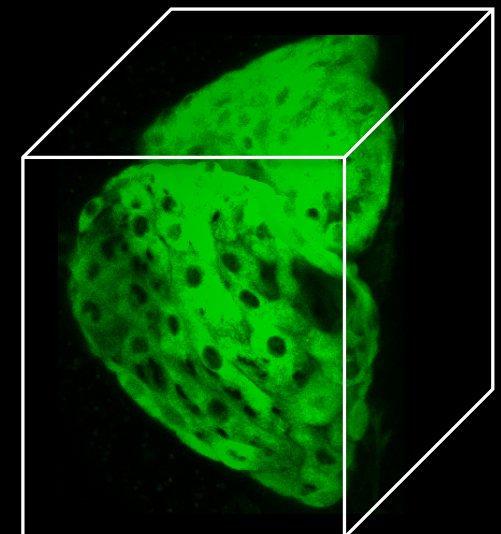
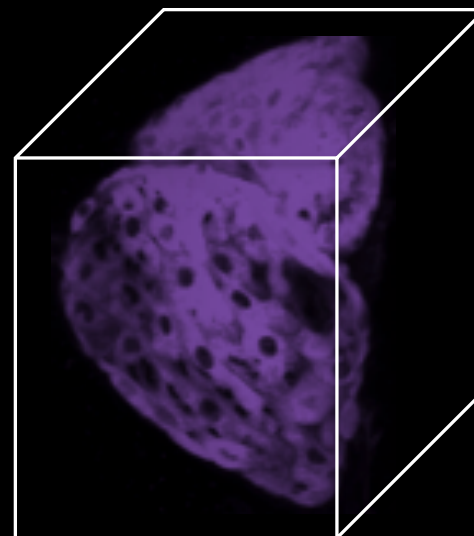
Physical sectioning
(invasive)



Optical sectioning
(non-invasive)

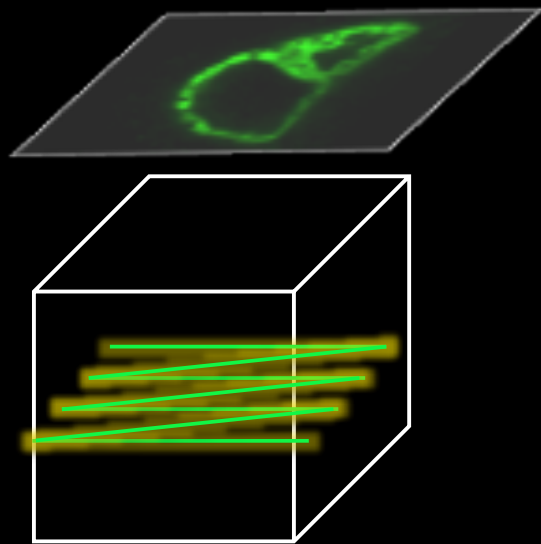


Poss et al, (2002) (*Science*)



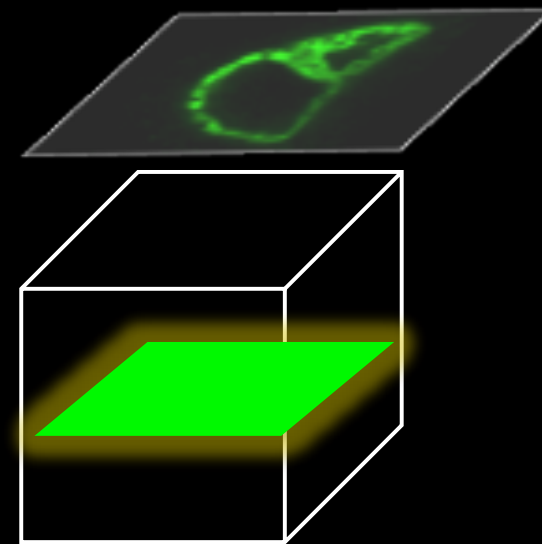
Building up a 2D image

1D detectors (PMT, GaAsP, APD)



Raster scan to
form 2D images

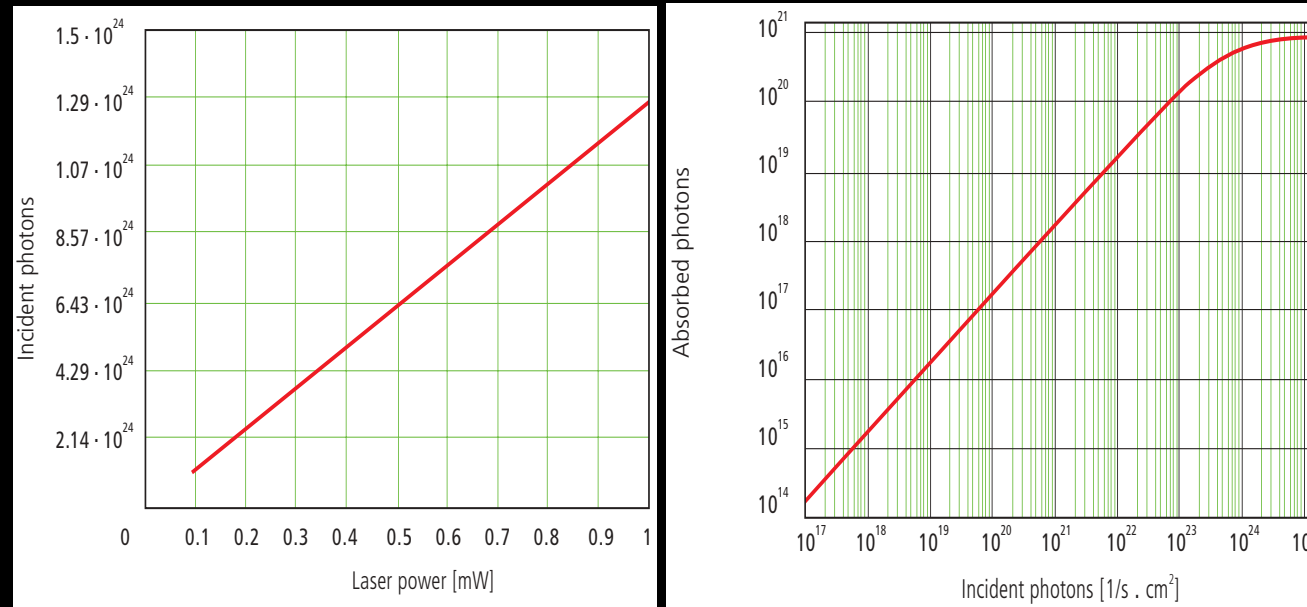
2D detectors (CCD, CMOS, sCMOS)



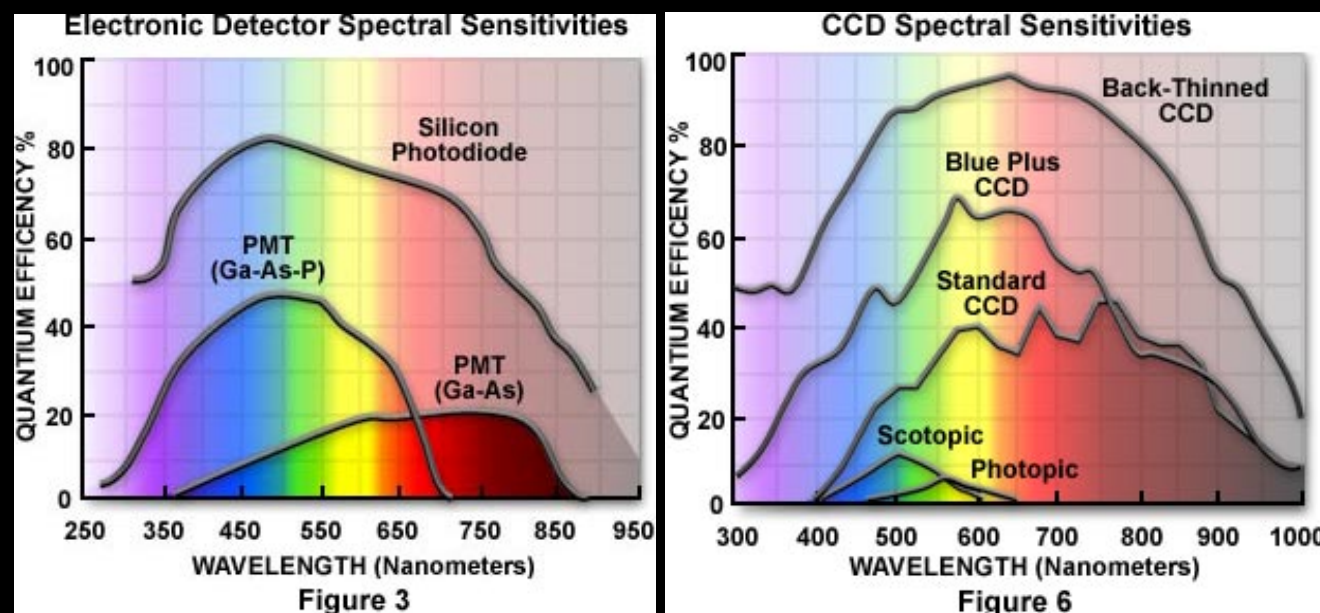
2D illumination

Imaging at the right microscope setting is crucial

Suitable laser power

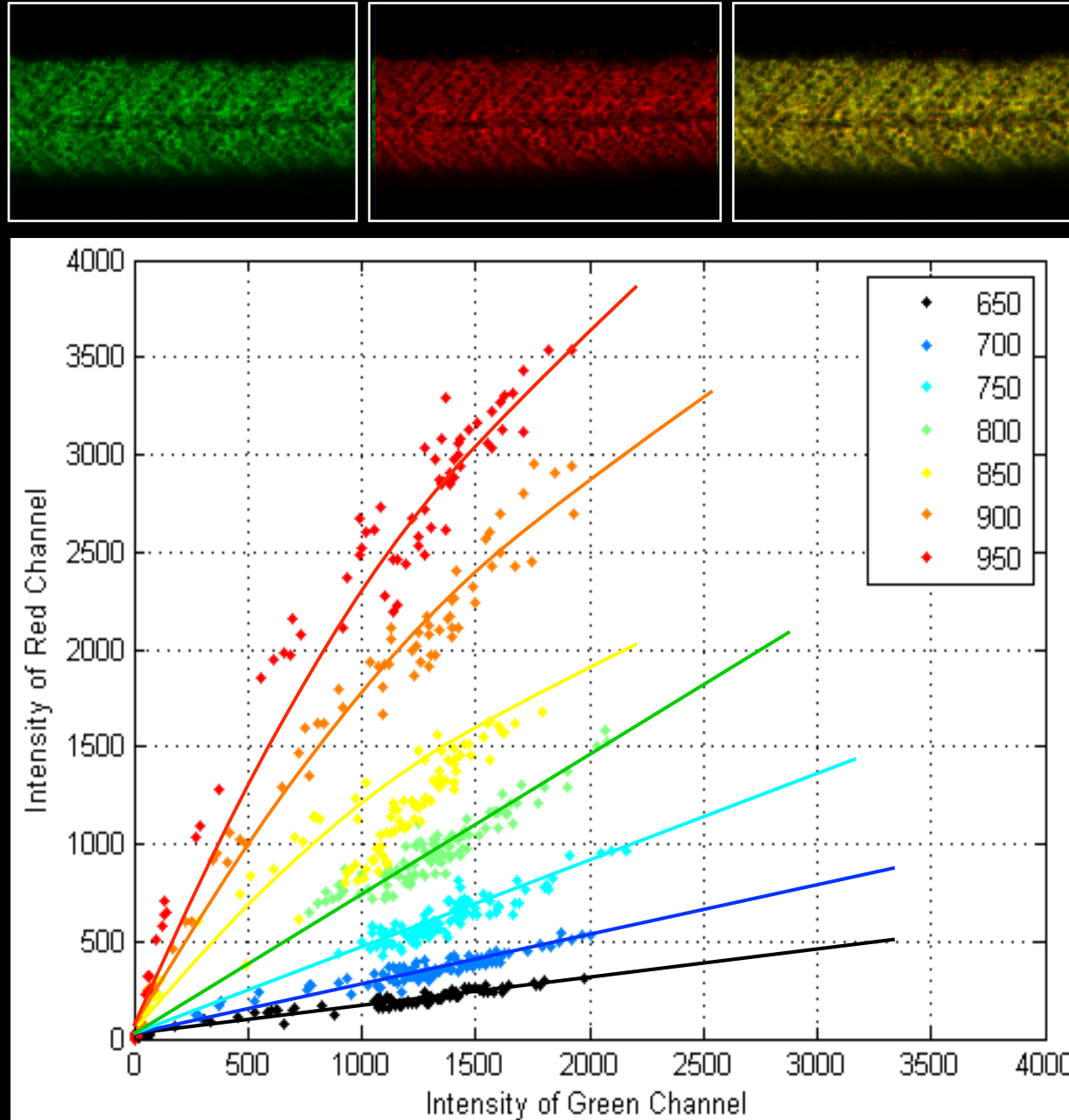


Suitable detector with a good quantum efficiency

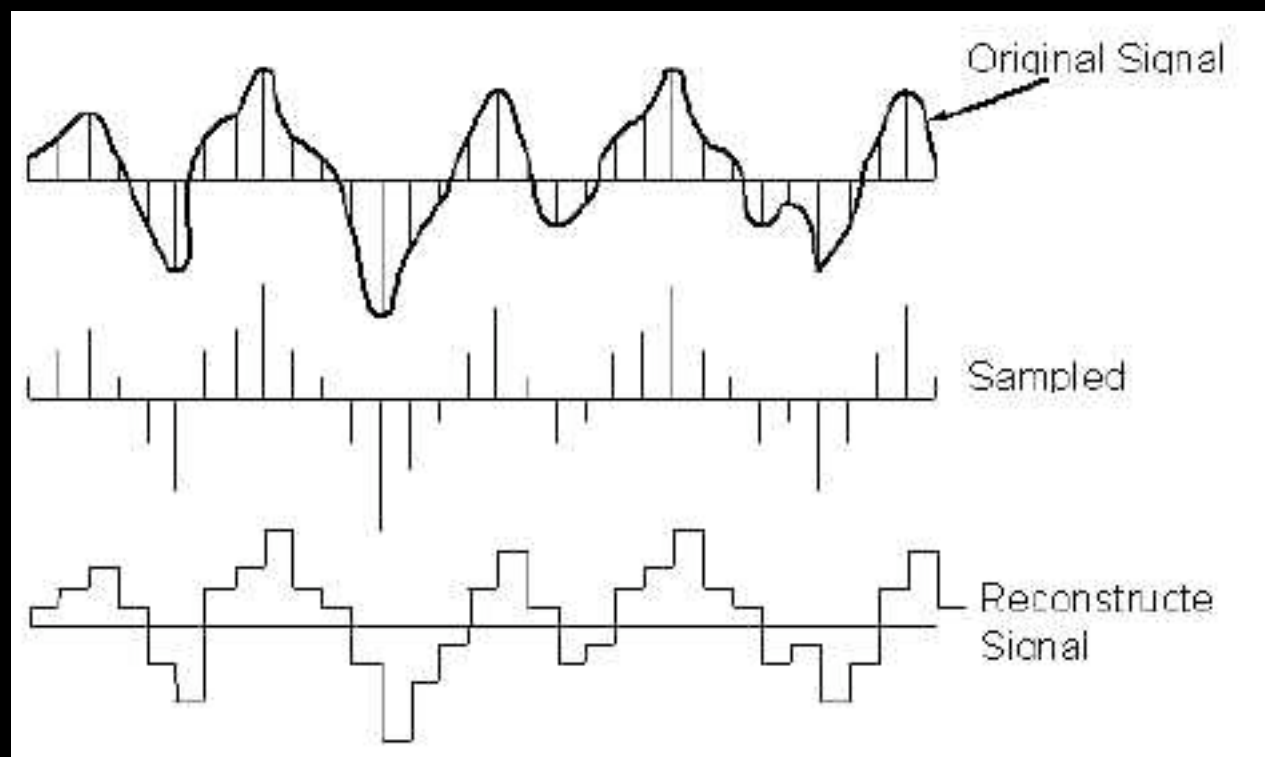


Imaging at the right microscope setting is crucial

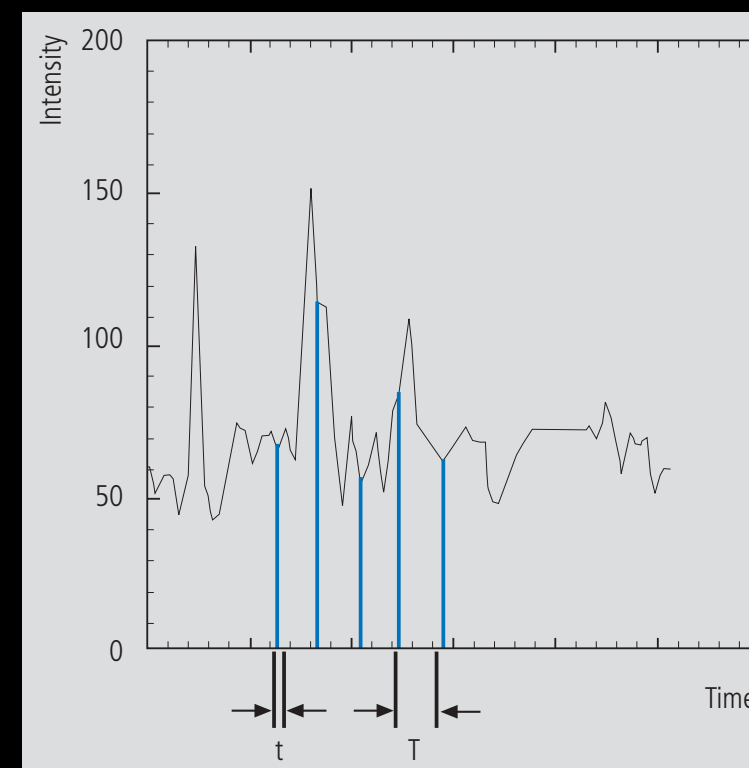
Suitable detector gain



Digitization

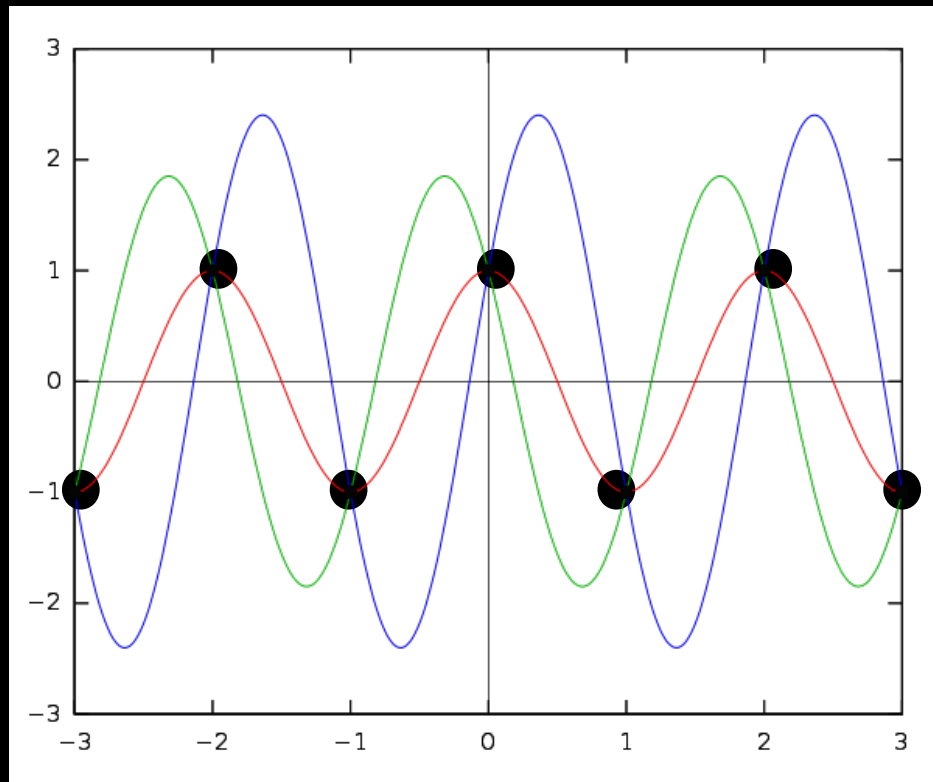


<https://andiemer.files.wordpress.com/2009/11/14-picture-1.png>

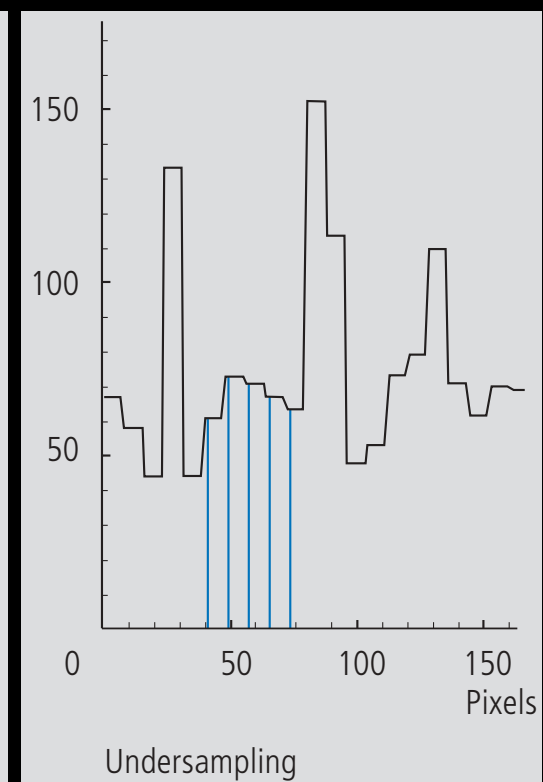
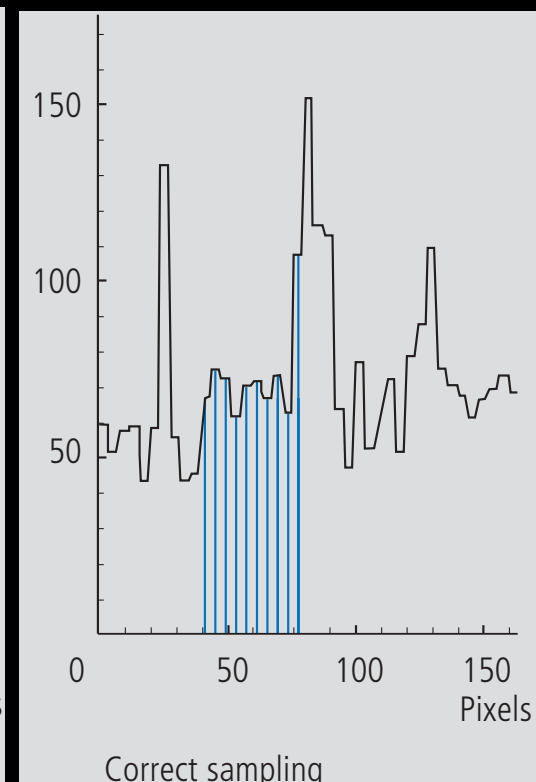
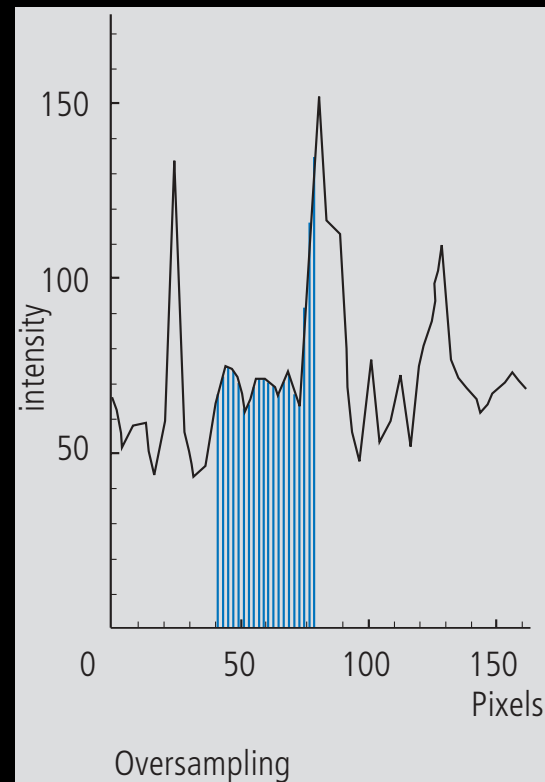


Carl Zeiss

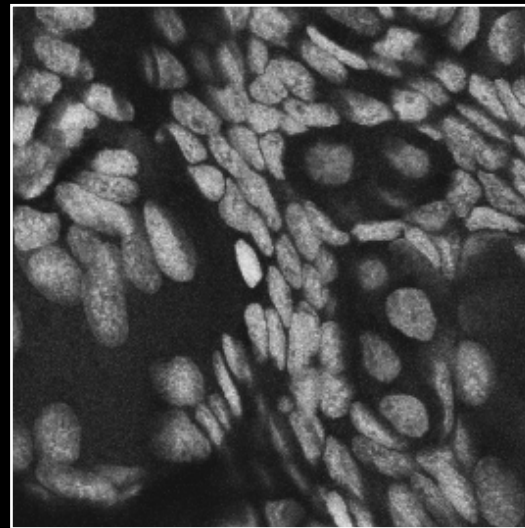
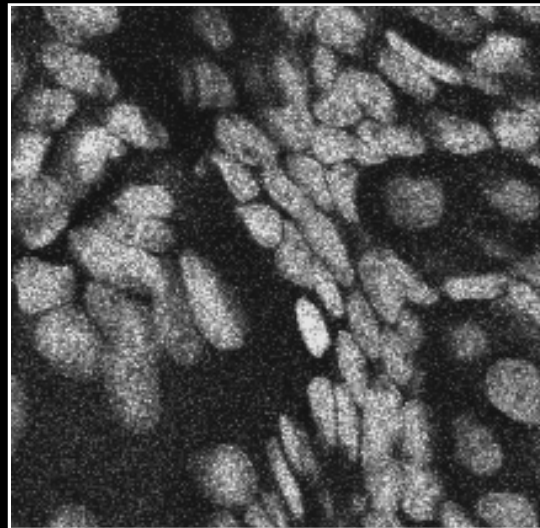
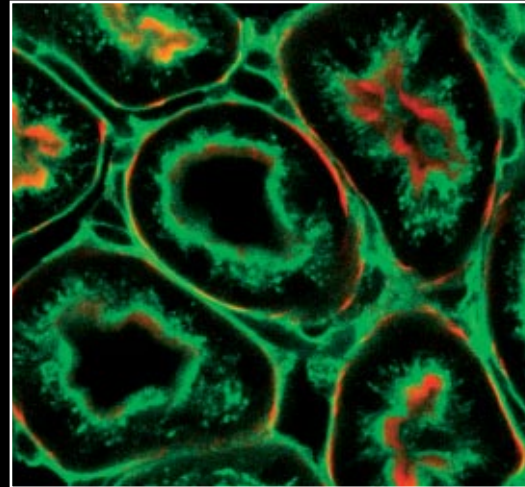
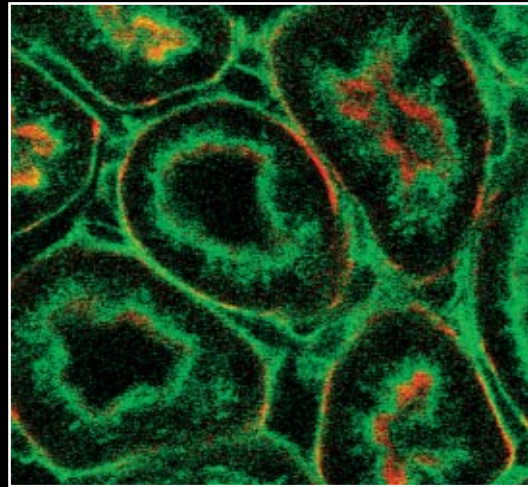
Correct Sampling



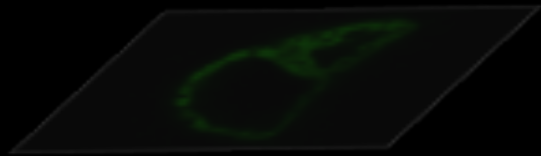
Wikipedia



Pixel dwell-time



Outline



Digital image formation

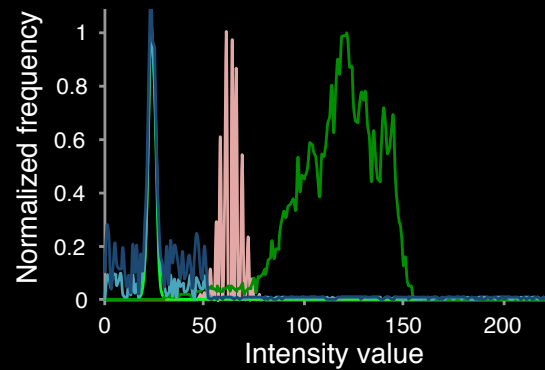


Image properties

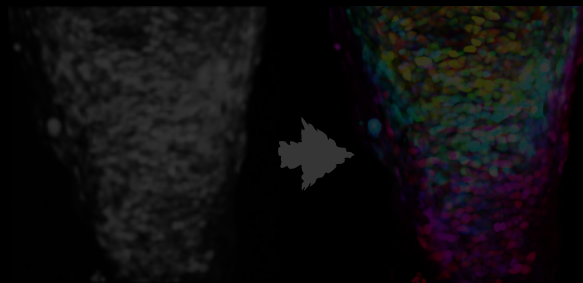
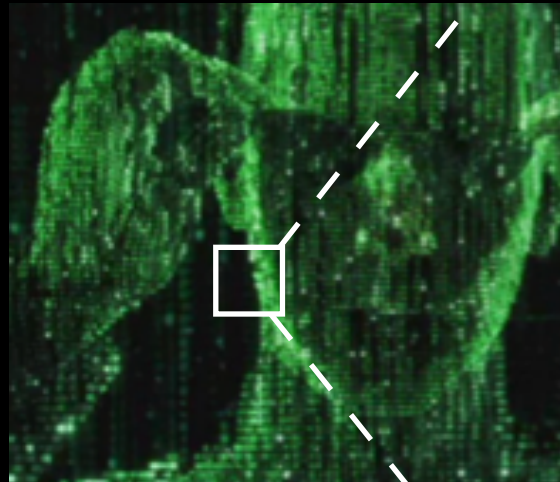


Image processing



Available softwares

Image is a matrix of numbers



			230	221	
			250	203	
			240	202	
				220	200
					230

Gray values

How high can the numbers be ?

			230	221	
			250	203	
			240	202	
				220	200
					230

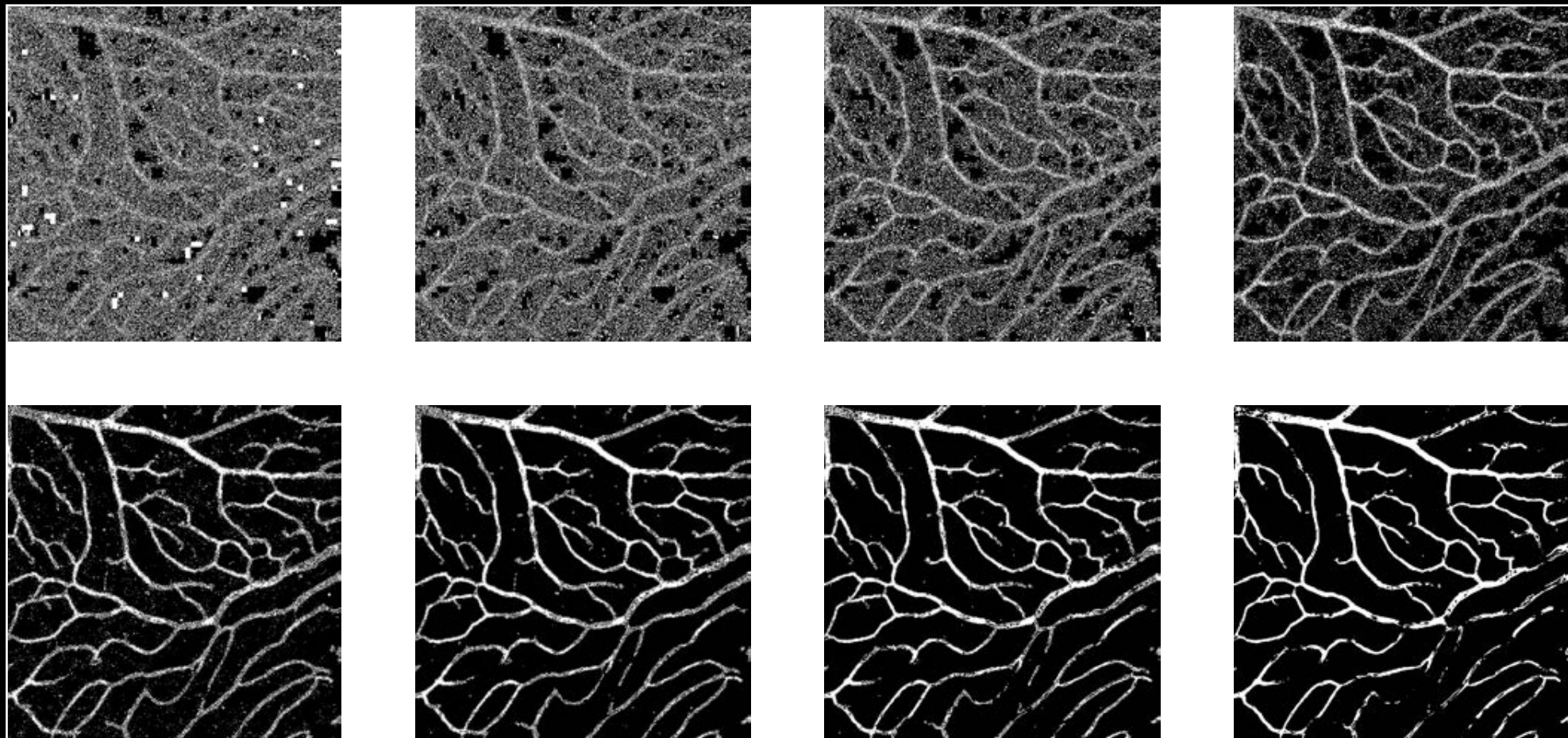
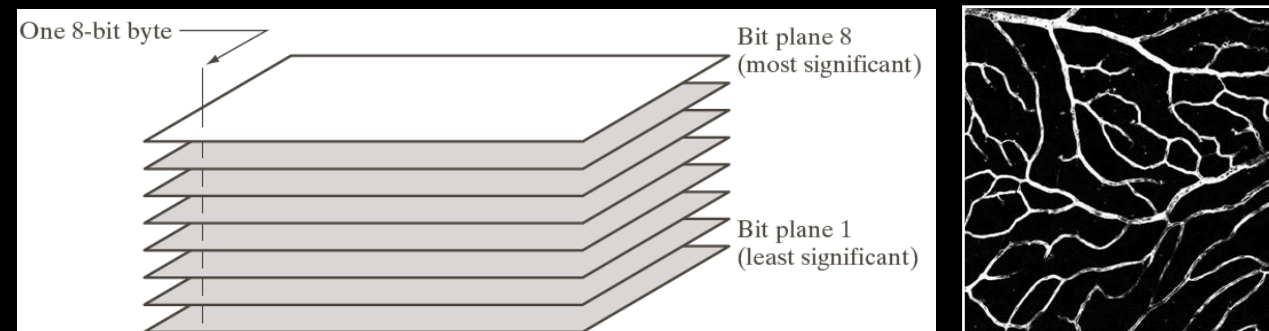
$$(0 - 11)_{\text{binary}} = (0 - 1 \times 2^1 + 1 \times 2^0)_{\text{decimal}}$$

$$= (0 - 3)_{\text{decimal}}$$

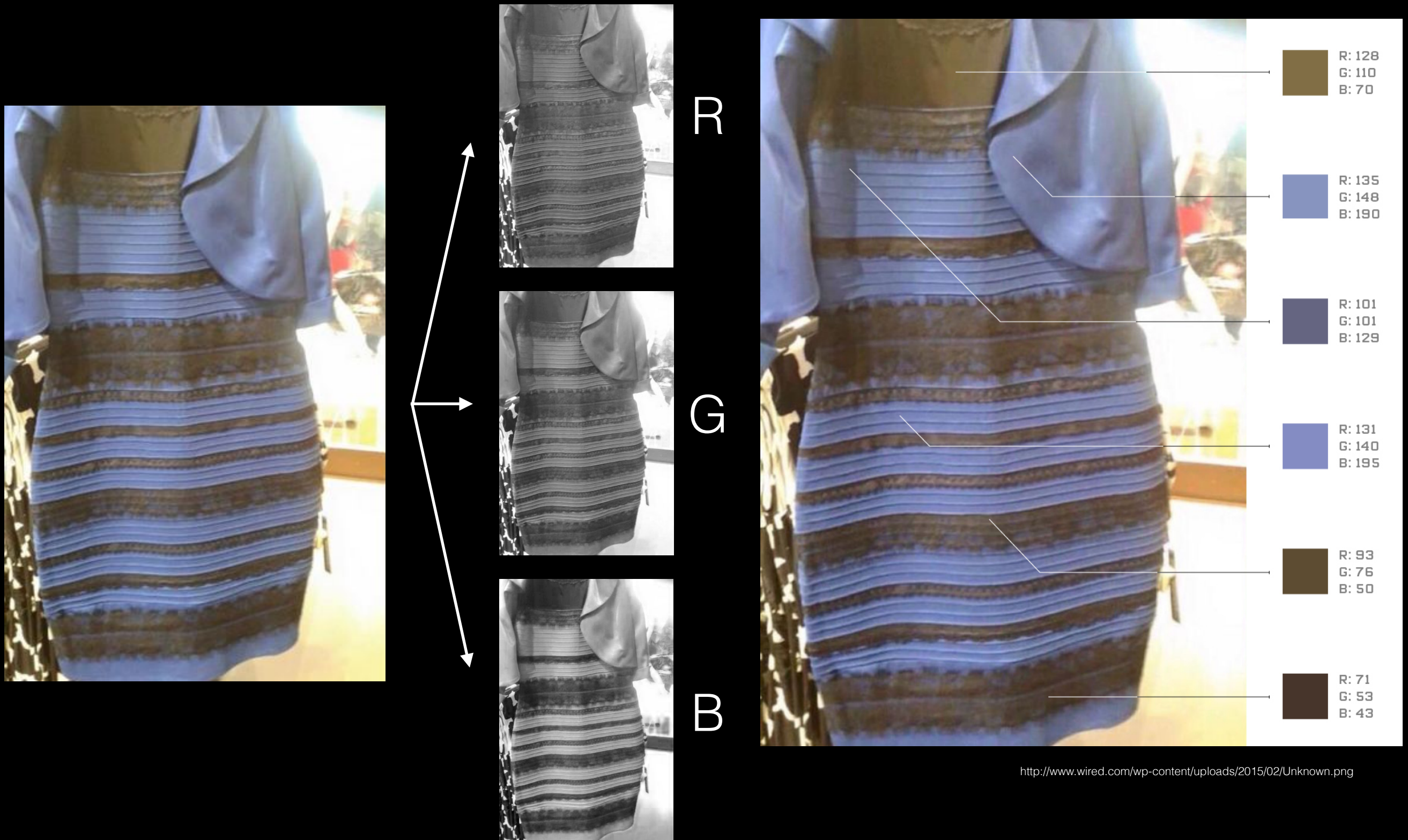
0 - 1	1-bit	0 - 1
0 - 11	2-bit	0 - 3
:	:	:
:	:	:
0 - 11111111	8-bit	0 - 255
:	:	:
0 - 11111111111111	12-bit	0 - 4095
:	:	:
0 - 111111111111111111	16-bit	0 - 65535

Dynamic Range

Image bit-depth



Colored Images



Colored Images



A



B



C



ABC



BAC



CBA



ACB

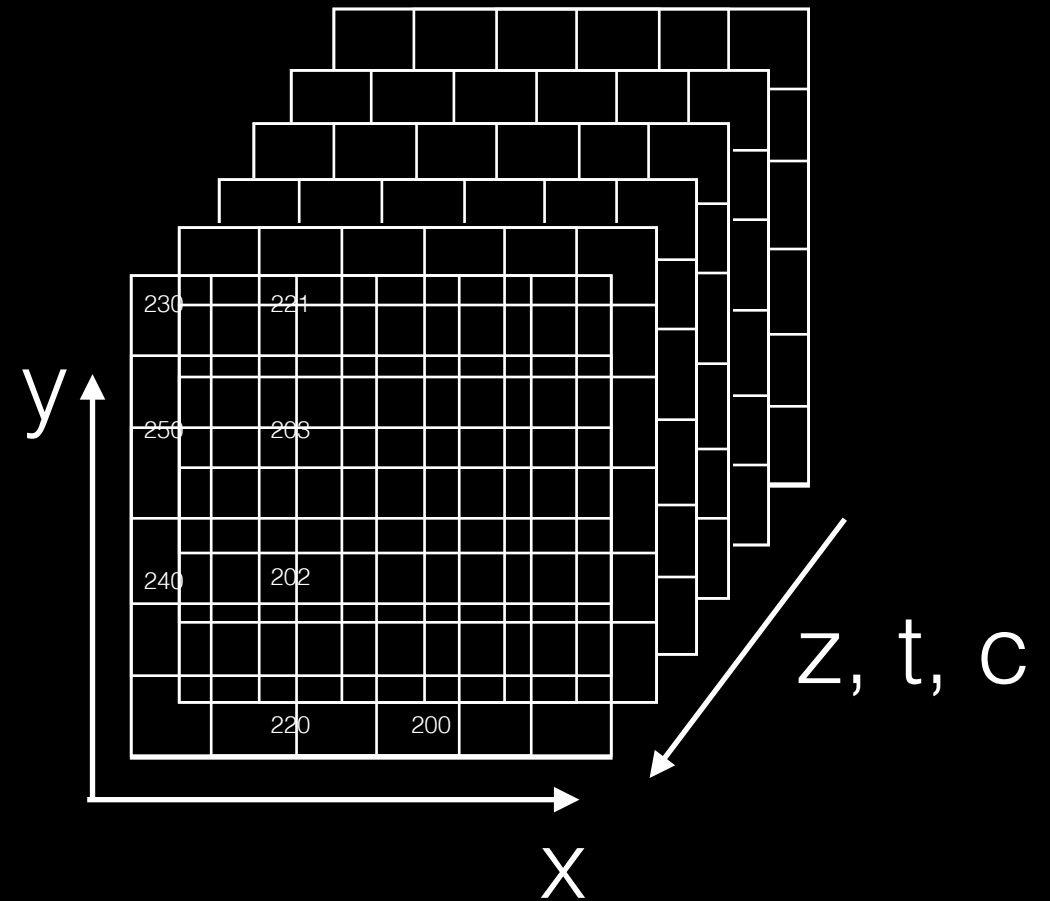
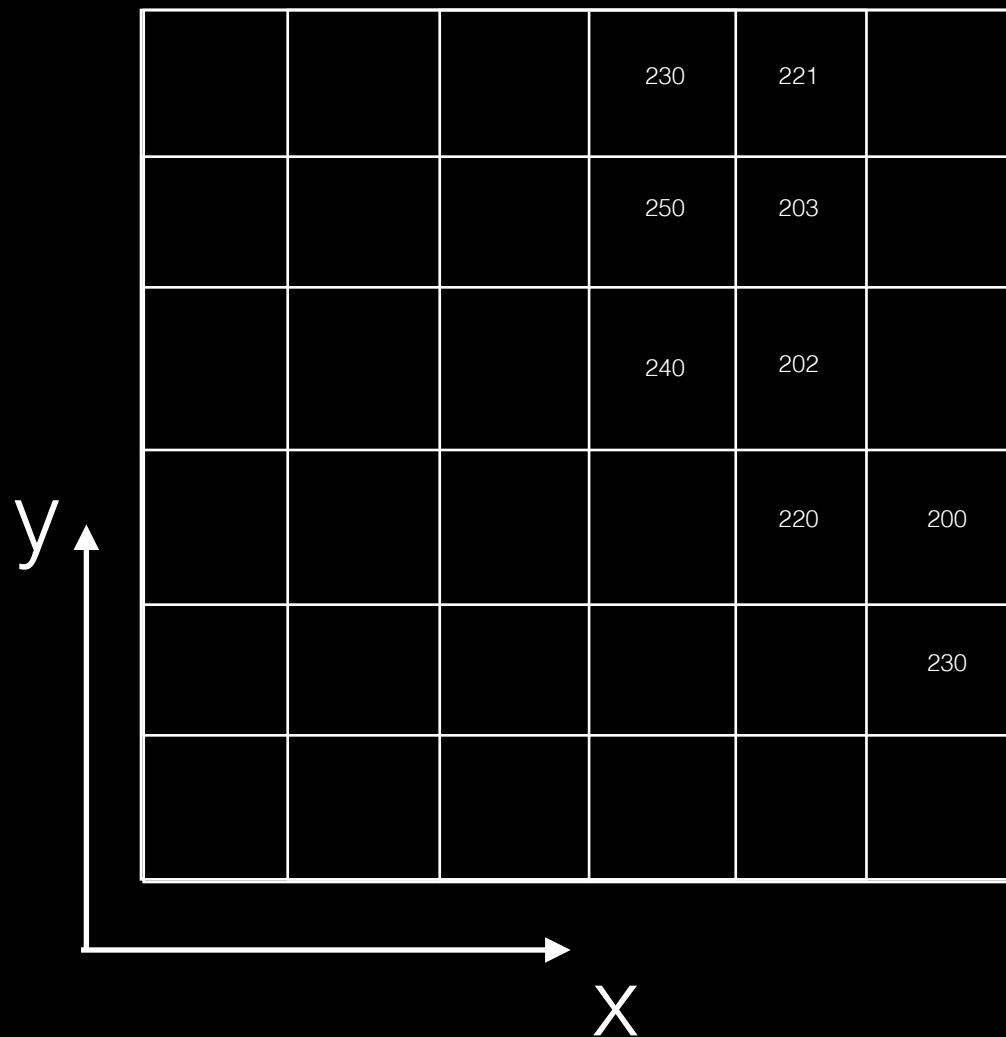


BCA



CAB

Stacks and Hyper-stacks

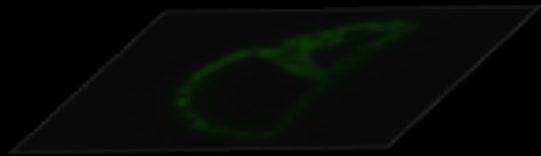


Meta-data

metadata	value
FileName	/Labs/images/EX36-CG-3.lsm
FileModDate	23-Aug-2011 13:20:11
FileSize	877951876
Format	TIF
Width	2048
Height	1024
BitDepth	16
ColorType	Indexed
BitsPerSample	16
Compression	Uncompressed
PhotometricInterpretation	RGB Palette
MaxSampleValue	65535
MinSampleValue	0
Binning	1
PixelResolutionUnitX	microns
PixelResolutionUnitY	microns
PixelResolutionUnitZ	microns
PixelResolutionX	0.155370
PixelResolutionY	0.155370
PixelResolutionZ	4.754000

- information about image content
- acquisition parameters
- manufacturer-specific data

Outline



Digital image formation

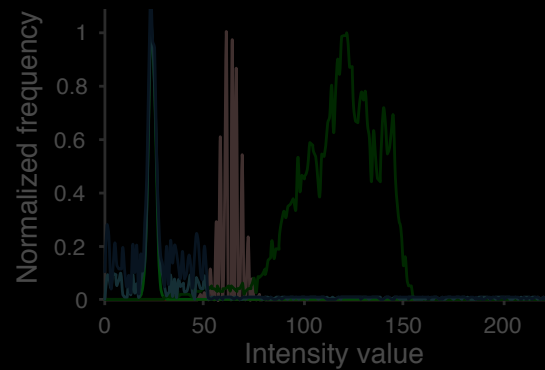


Image properties

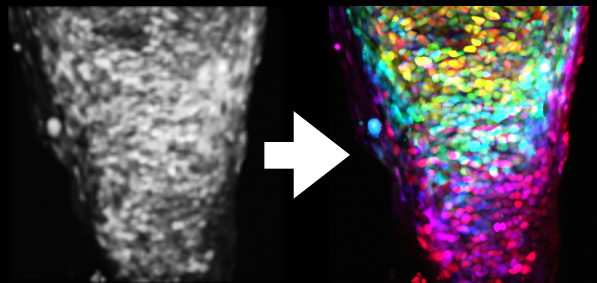


Image processing



Available softwares

Resources

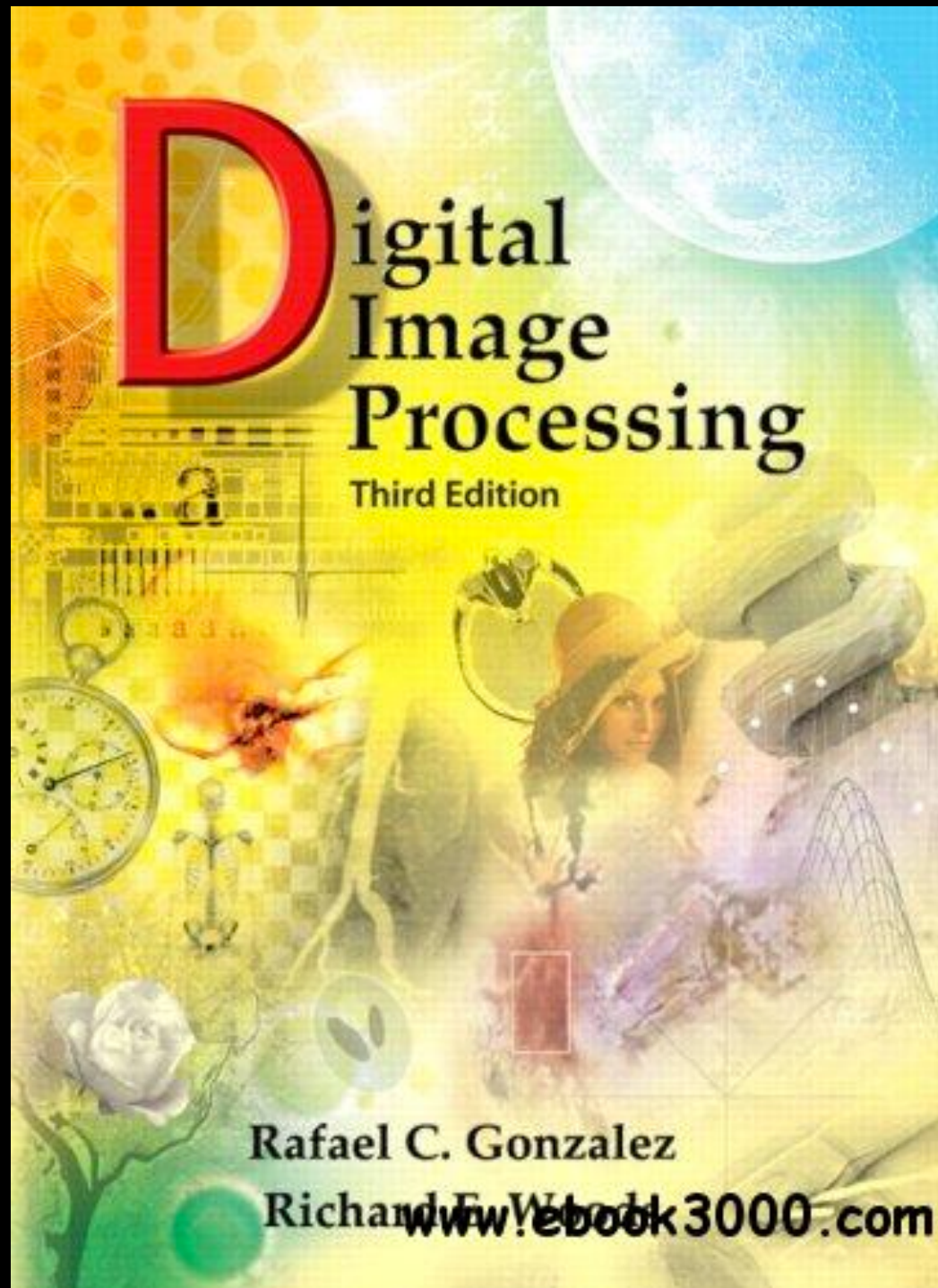


Image Processing In Life Science

www.weizmann.ac.il/vet/IC/content/image-processing-life-science

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Image Processing Course

Image Processing In Life Science

Image Processing In Life Science- Enrichment Course

By: Dr. Aviad Baram, Yoseph (Sefi) Addadi, Dr. Guy Malkinson, Ofra Golani

This is an introductory course on digital image processing directed towards scientists in life science disciplines. The course will cover theory of various image processing concepts. The students will gain hands on practical experience in applying various image processing tools and methods to data collected in life science laboratories. Image processing concepts and techniques will be demonstrated on the open source, freely available, software package: ImageJ.

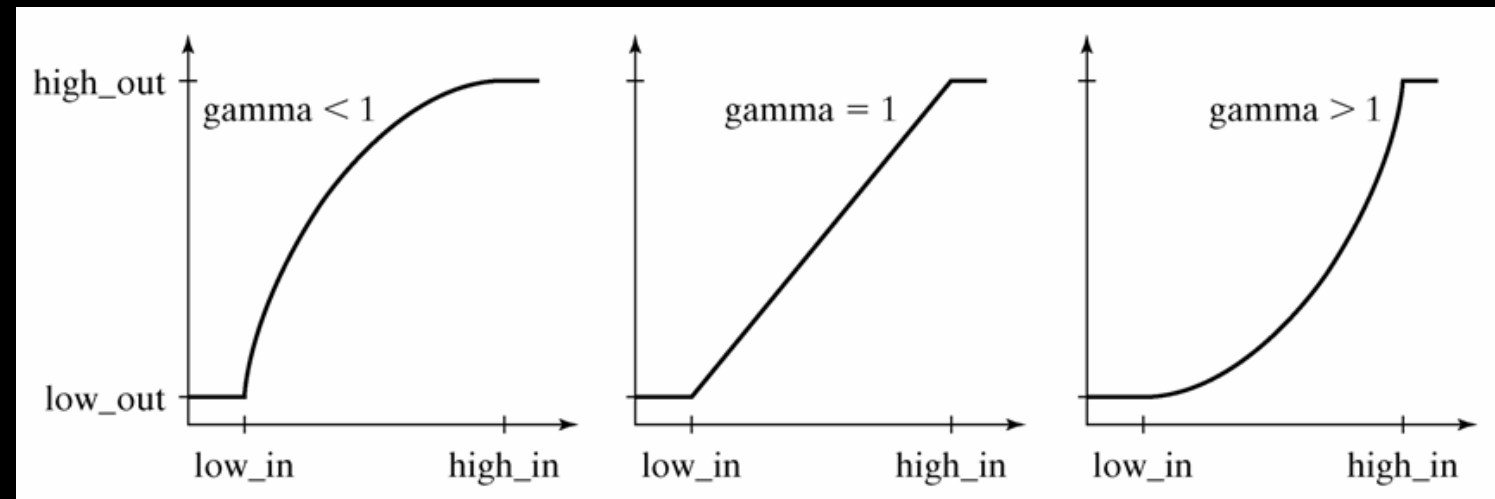
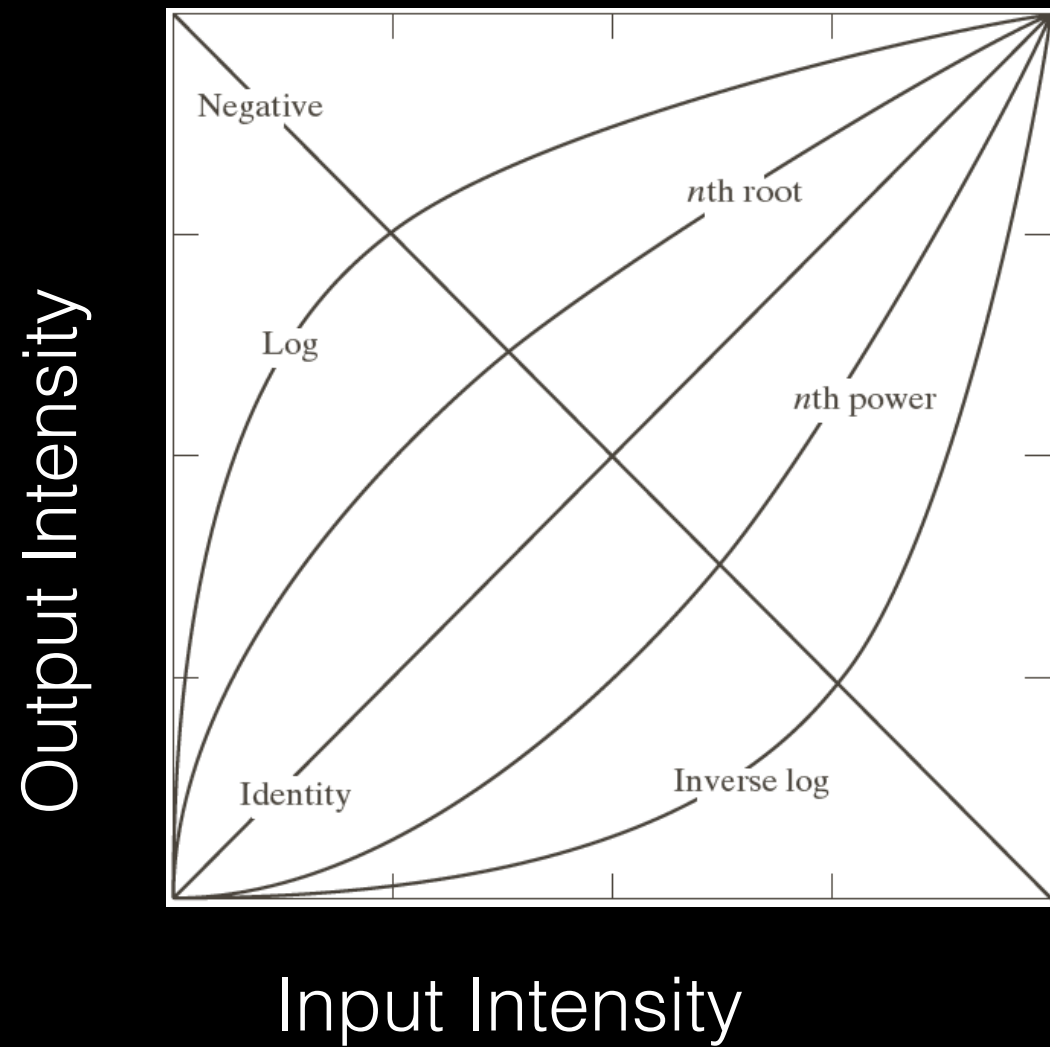
Course outline:

1. Digital image fundamentals (Guy Malkinson, 14.3, [PPT](#))
2. Image enhancement in the spatial domain (Sefi Addadi, 21.3, [PDF](#))
3. Segmentation and Morphological Operations (Aviad Baram, 28.3, [PPT](#))
4. Image enhancement in the frequency domain (Aviad Baram, 4.4)
5. Multi dimensional image processing (Guy Malkinson, 18.4, [PPT](#))
See also: [3D Processing and Analysis with ImageJ](#)
6. ImageJ Macro Language (Aviad Baram, 2.5)
7. Image Analysis Applications (Maya Dadiani, Zvi Kam, 9.5)

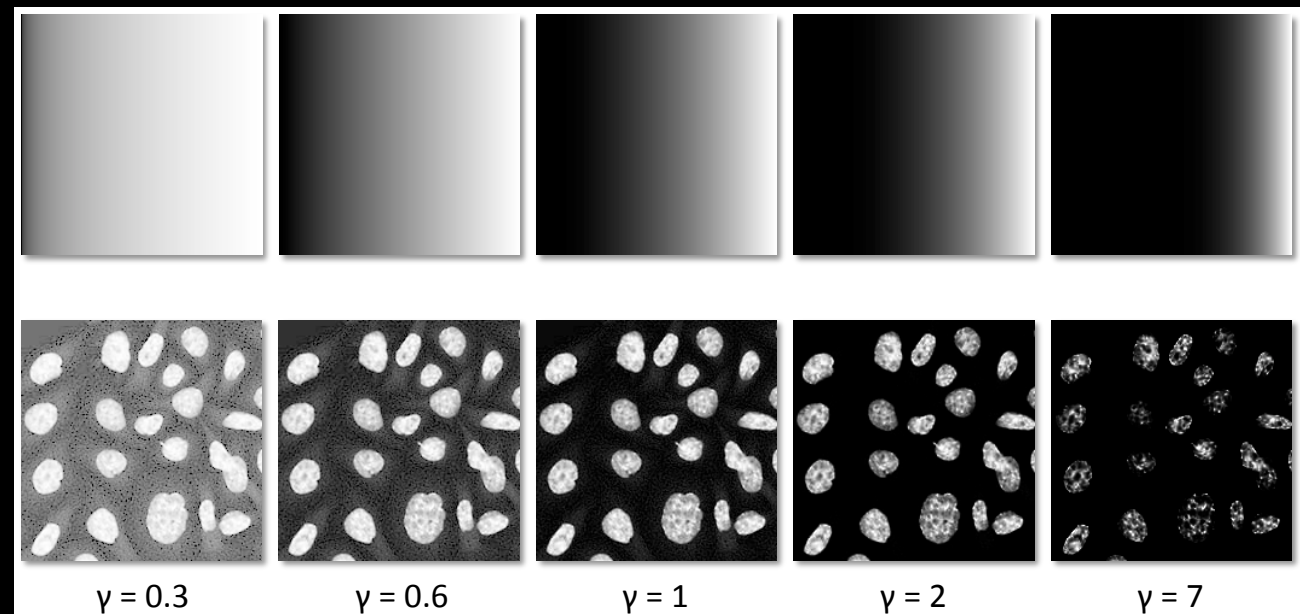
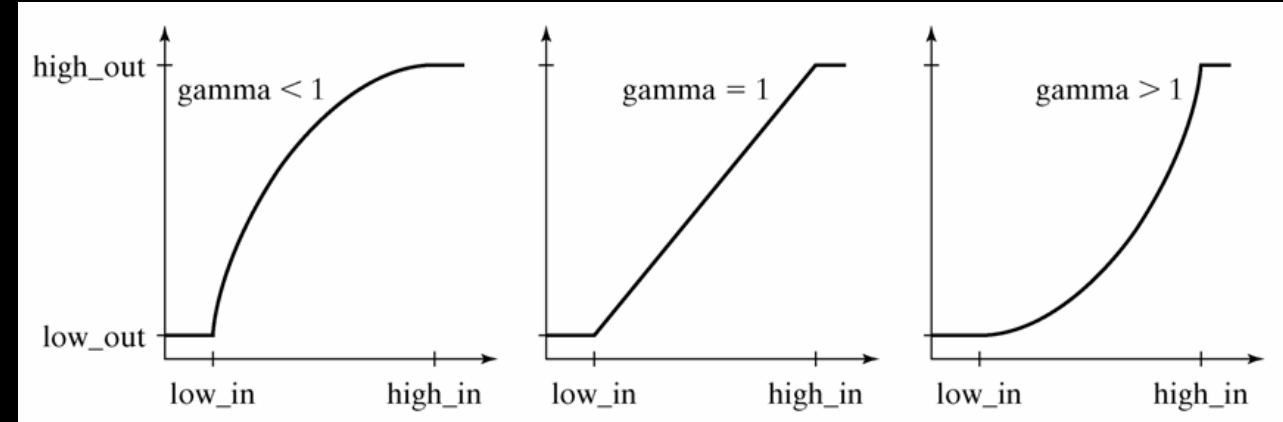
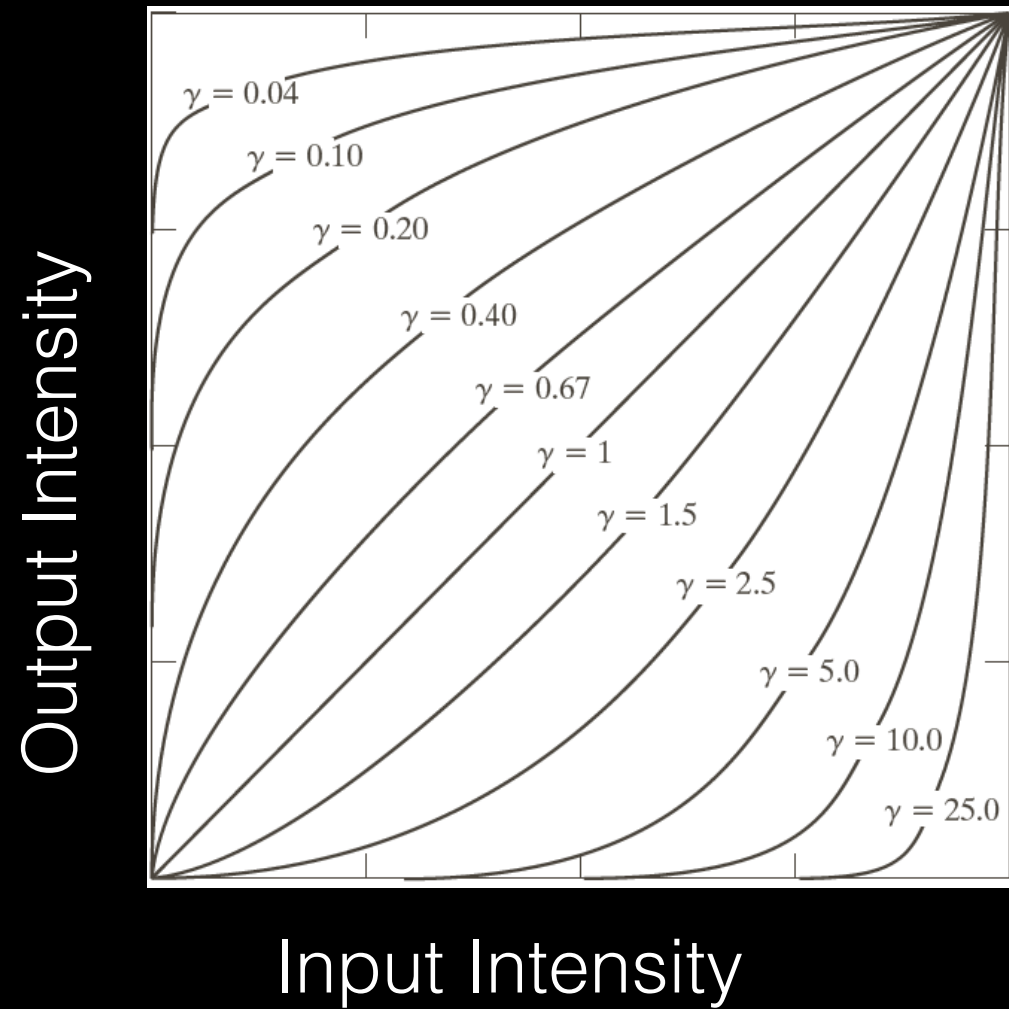
Links to Image Analysis Courses and Lectures

[Zvi Kam: Quantitative Image Analysis](#)
[Basics of Quantitative Image Analysis](#)
[Image Processing and Analysis Courses @ EMBL \(ImageJ\)](#)
[Stefan Terjung: Basics in Image Processing, EMBL](#)
[Yury Belyaev: Rendering and analysis of confocal datasets, EMBL](#)
[Biological Image Analysis Primer](#)

Contrast adjustment

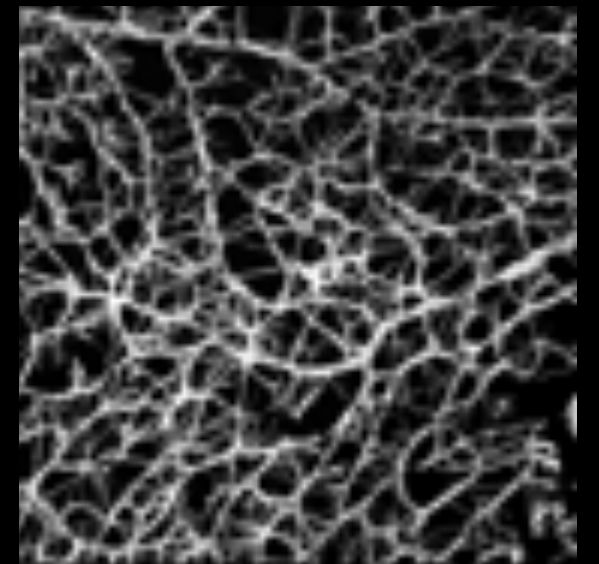
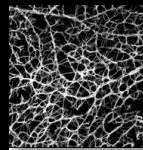
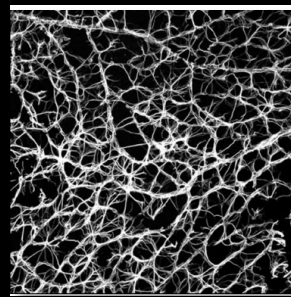
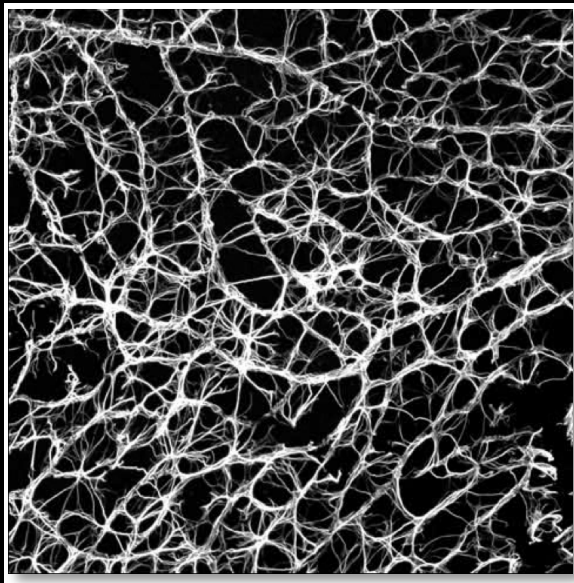


Contrast adjustment

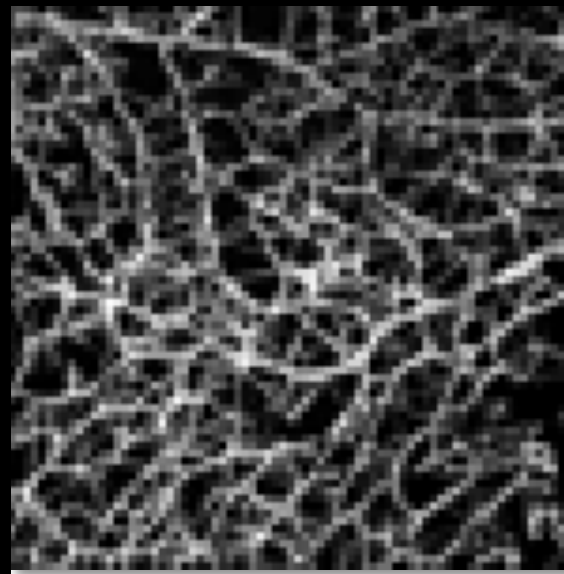
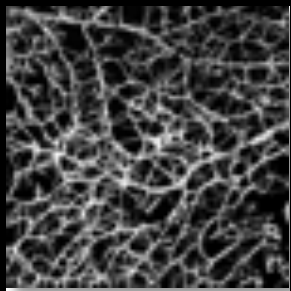


$$\text{Output} = c. (\text{Input Image})^{\gamma}$$

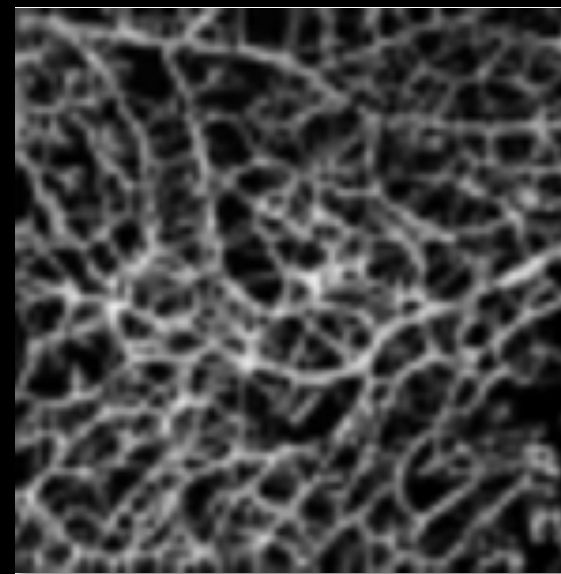
Binning



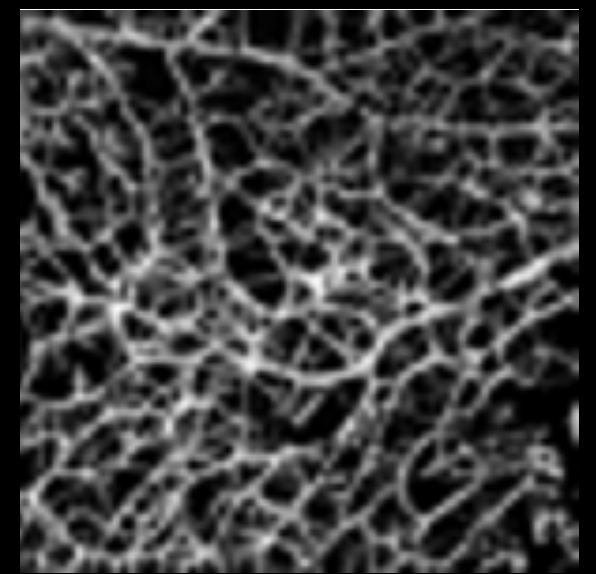
Interpolation



nearest neighbor
impose a finer grid,
pick nearest pixel in
original



bilinear
weighted average
of 2 x 2 neighbors

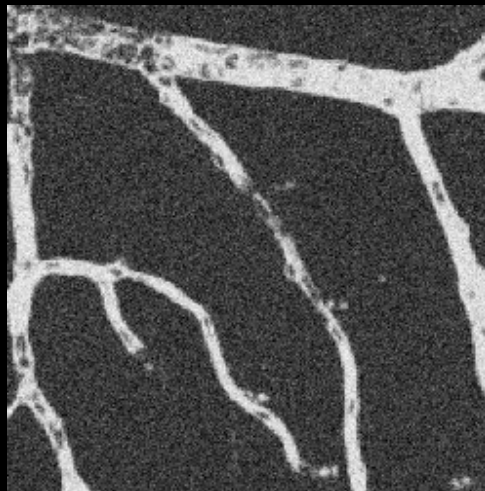


bicubic
weighted average
of 4 x 4 neighbors

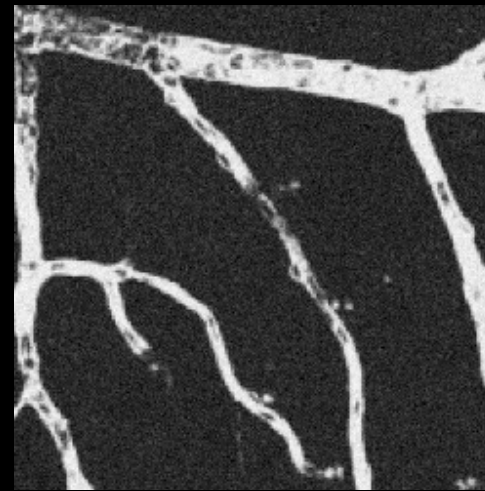
Average



single frame

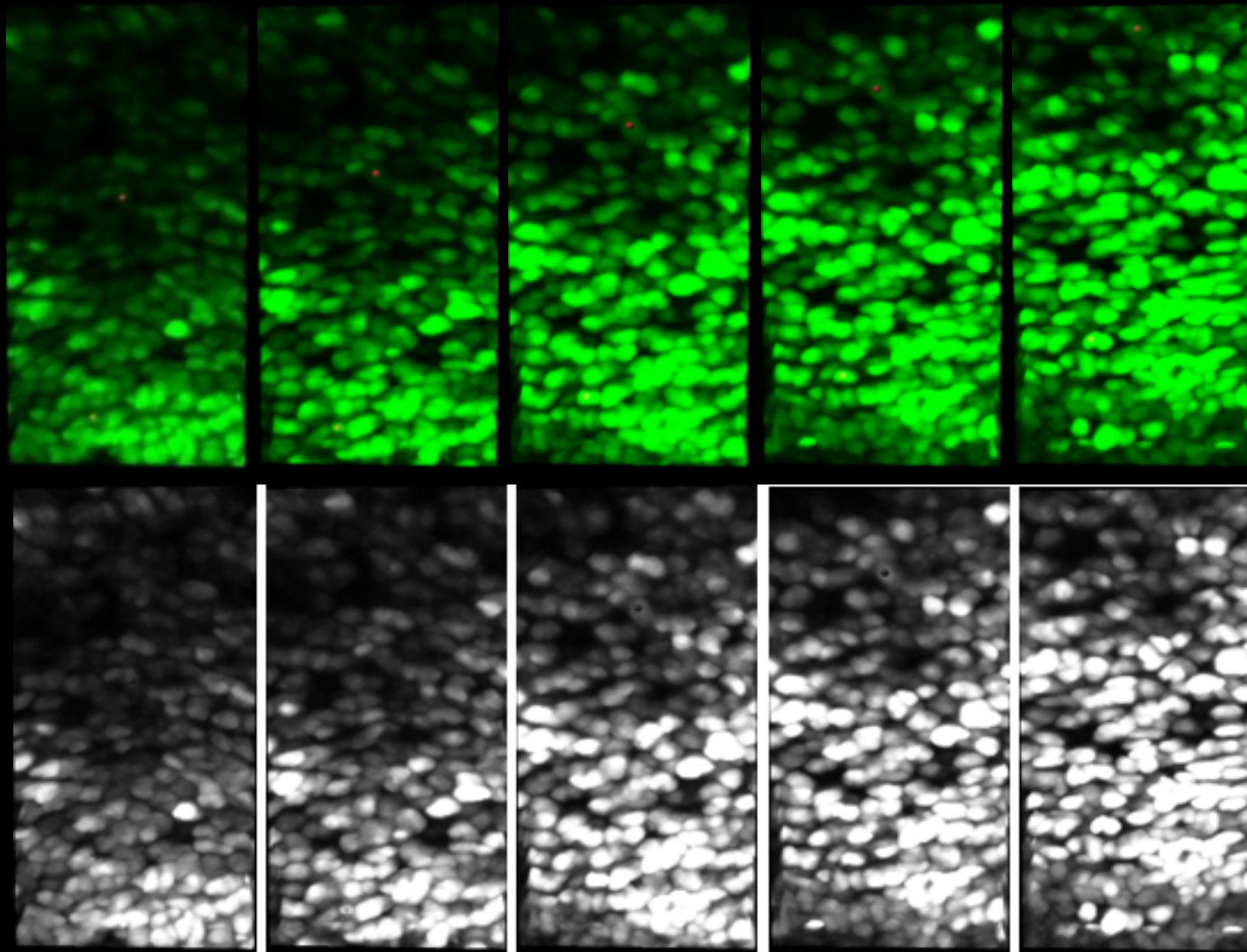


average of 20

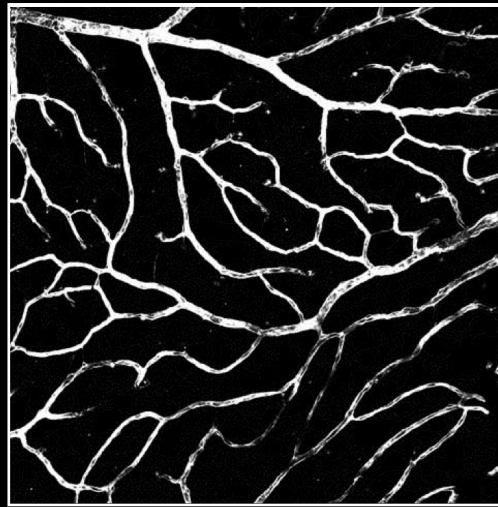


average of 100

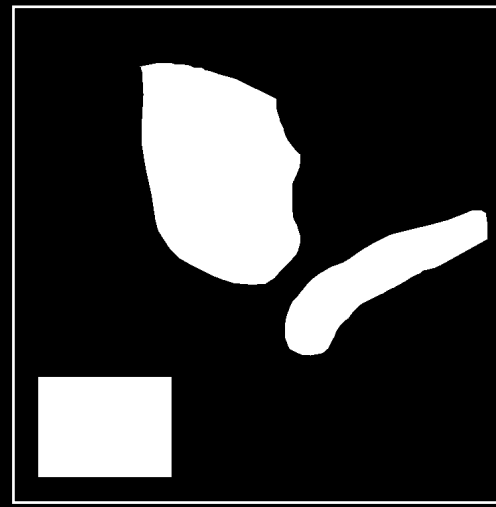
Binary images (thresholding)



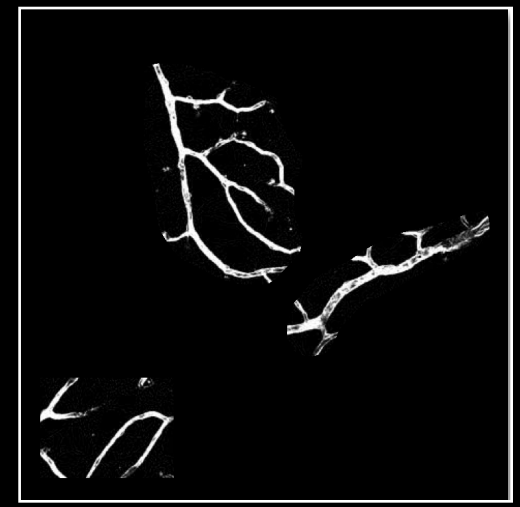
Mask



*



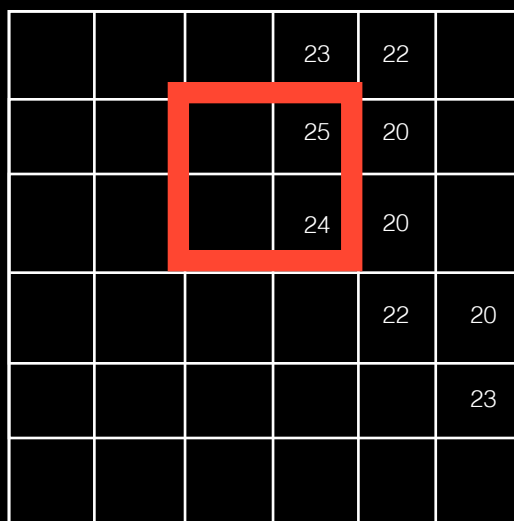
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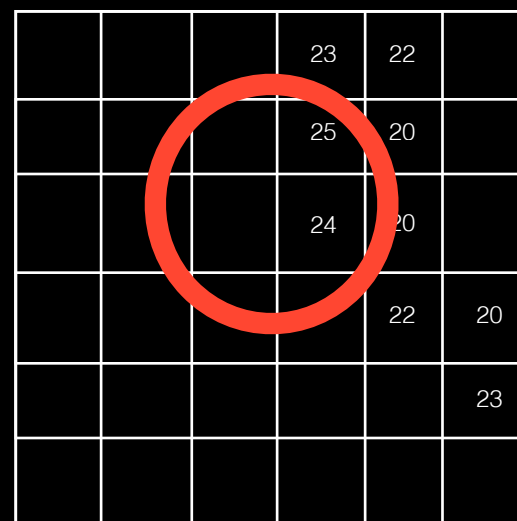
Spatial filtering

Smoothing

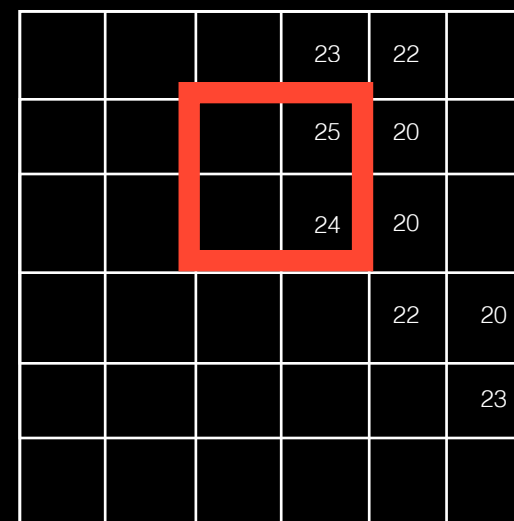
Average (Mean)



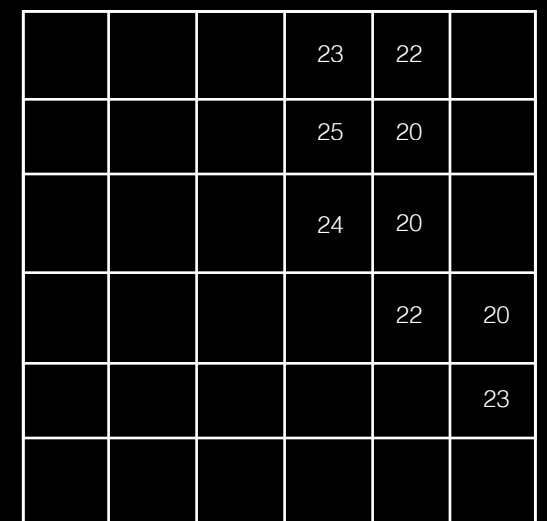
Disk



Median



Gaussian



Spatial filtering

Derivatives

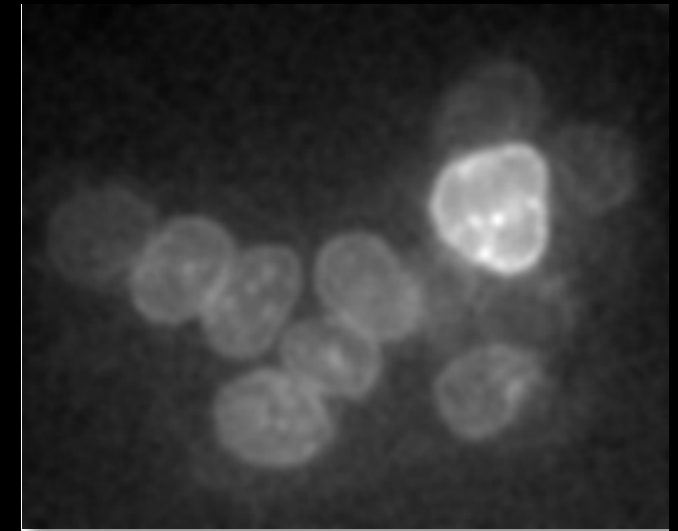
$$\frac{\partial f}{\partial x} = f(x+1) - f(x)$$

$$\frac{\partial^2 f}{\partial x^2} = f(x+1) + f(x-1) - 2f(x)$$

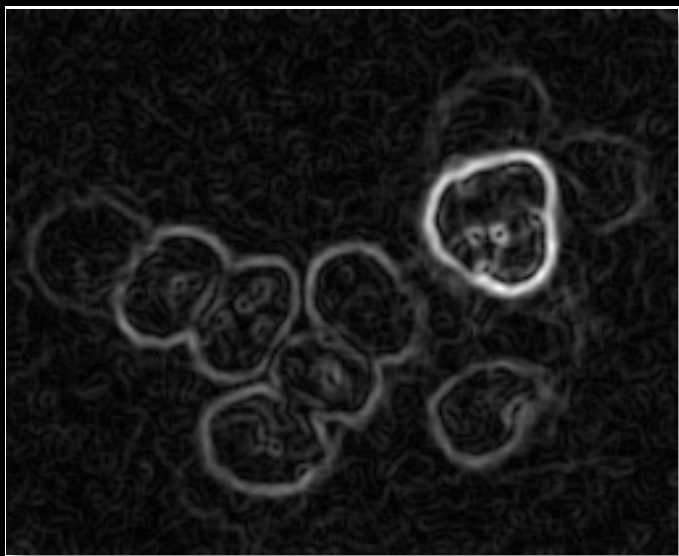
$$\nabla^2 f = \frac{\partial^2 f}{\partial x^2} + \frac{\partial^2 f}{\partial y^2}$$

$$\frac{\partial^2 f}{\partial x^2} = f(x+1, y) + f(x-1, y) - 2f(x, y)$$

$$\frac{\partial^2 f}{\partial y^2} = f(x, y+1) + f(x, y-1) - 2f(x, y)$$

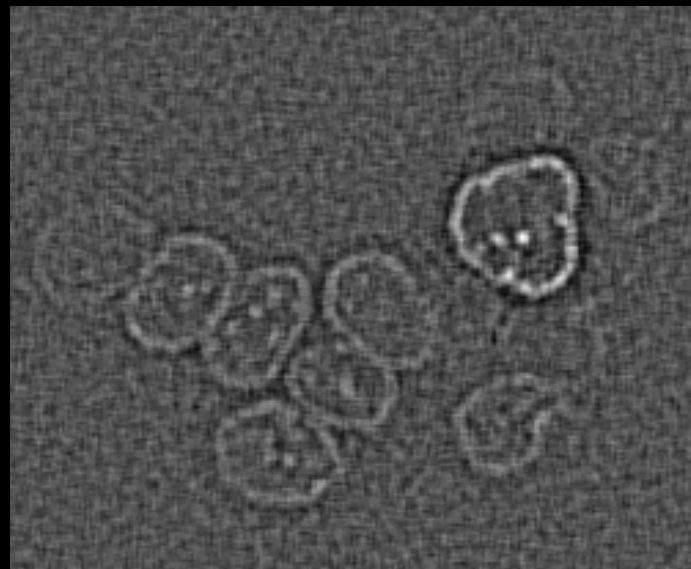


Gradient



$$\left| \frac{\partial f}{\partial x} \right| + \left| \frac{\partial f}{\partial y} \right|$$

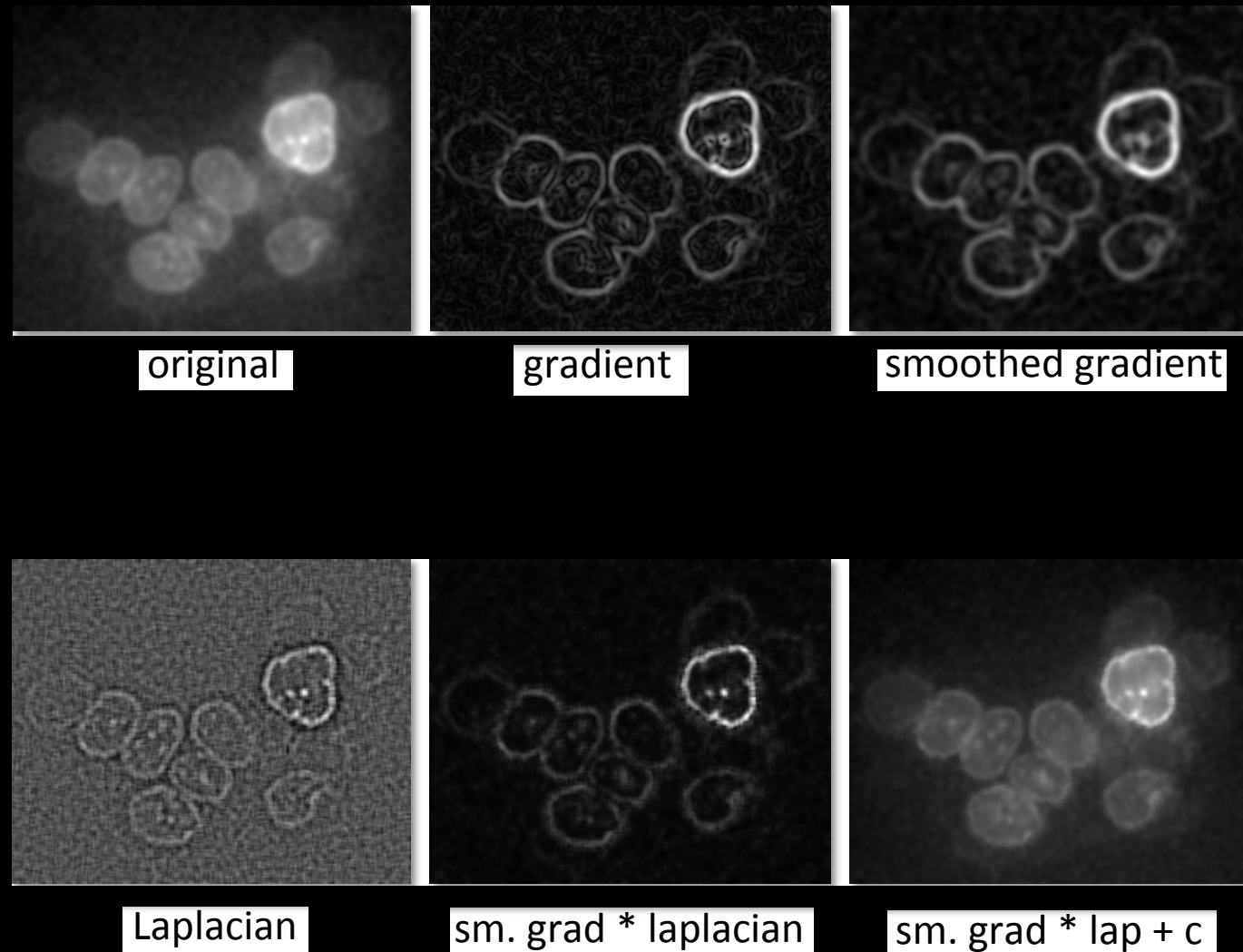
Laplacian



$$\nabla^2 f(x, y)$$

LoG

Combined filtering



Edge detection and segmentation

Image similarity

Image differences

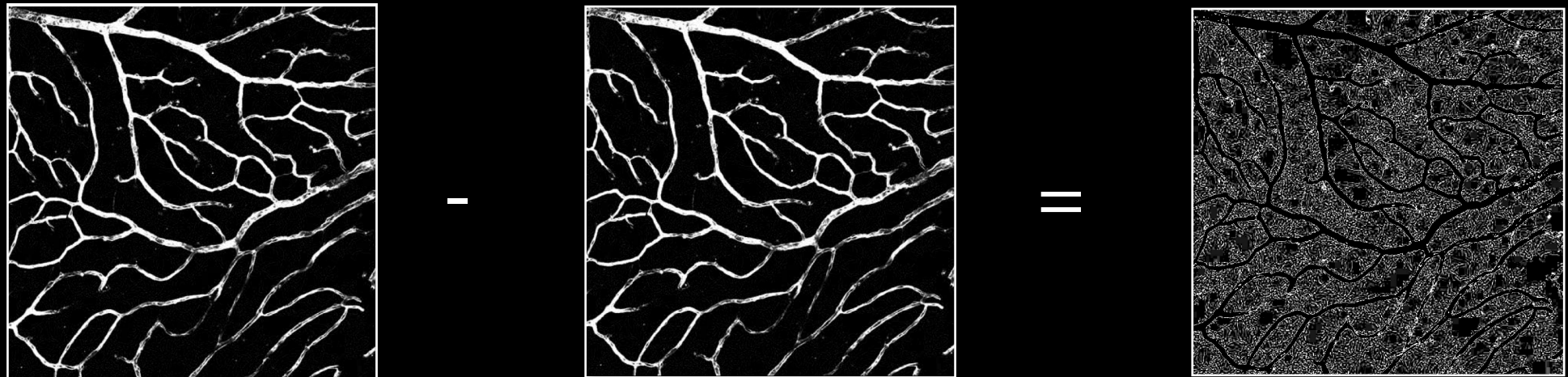


Image similarity

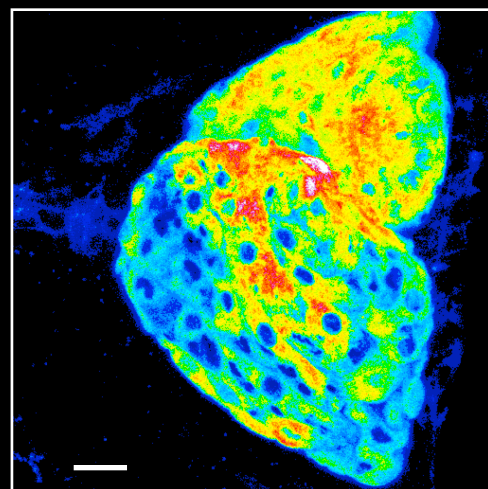
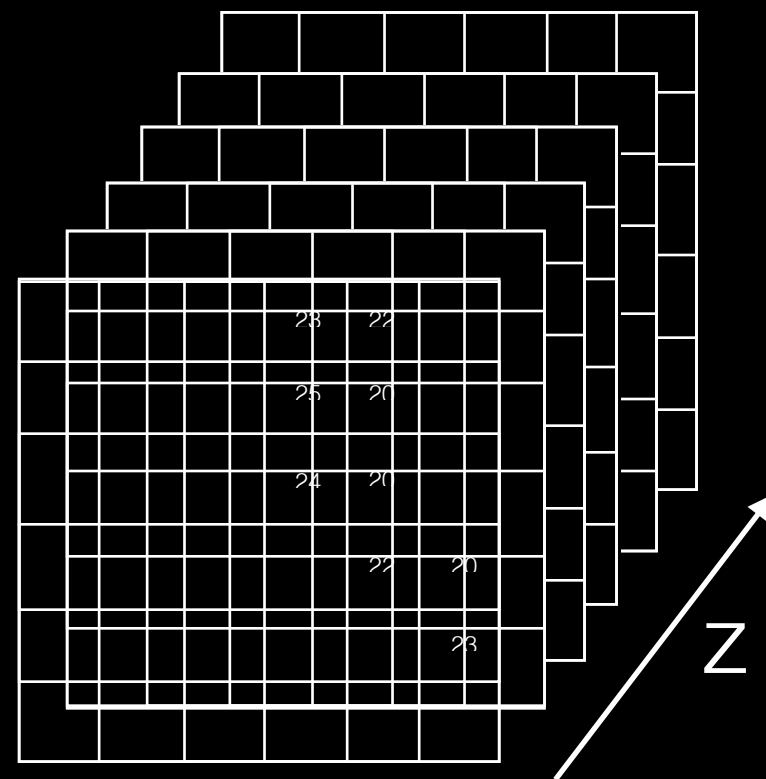
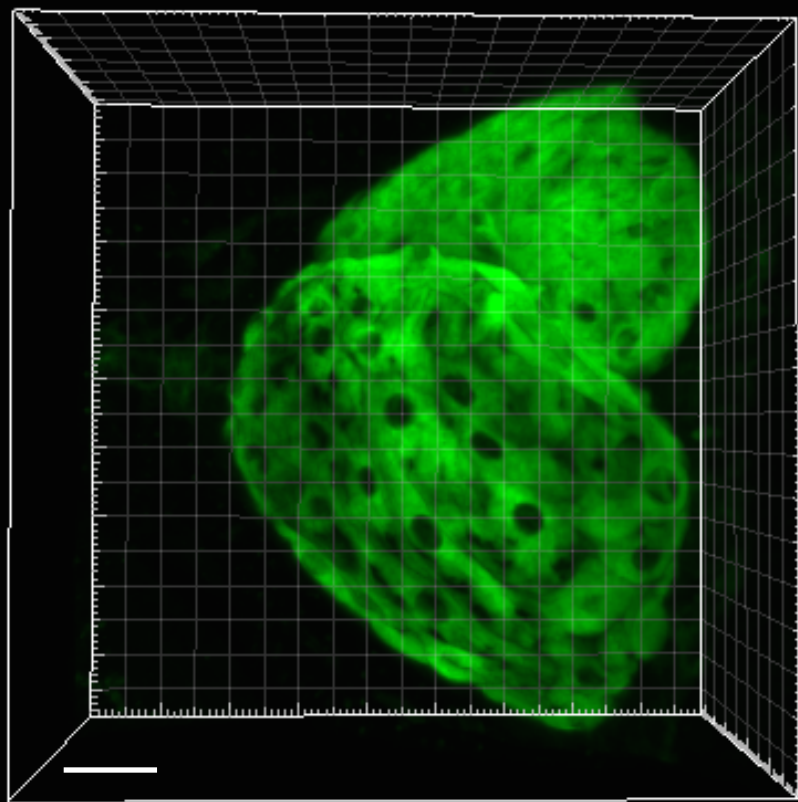
Image correlation

			23	22	
			25	20	
			24	20	
				22	20
					23

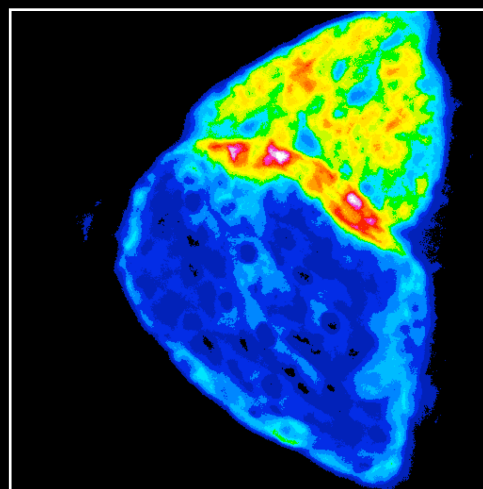
			23	22	
			25	20	
			24	20	
				22	20
					23

Image convolution

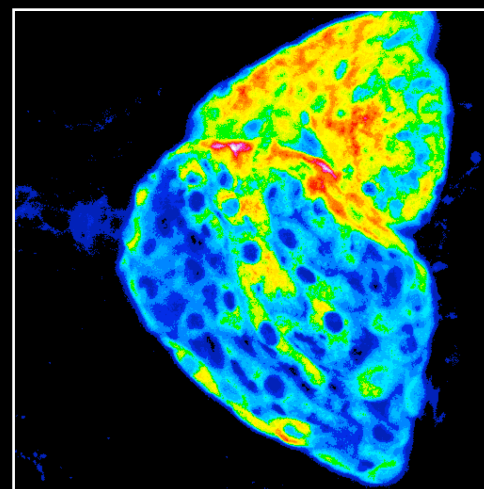
Intensity Projections



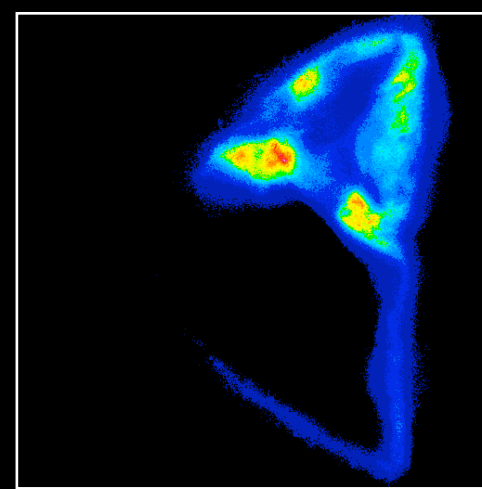
Maximum



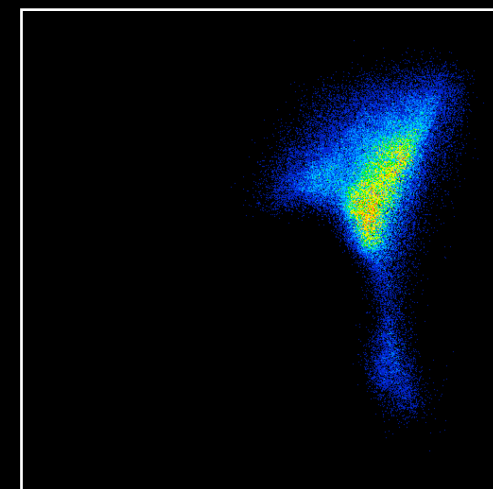
Average



Standard Dev.



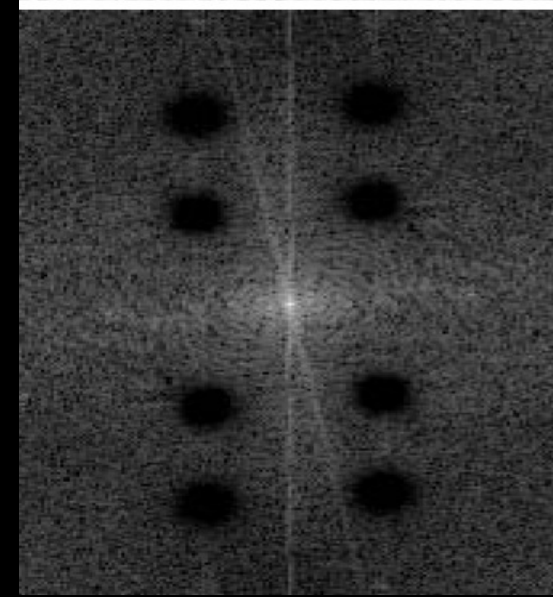
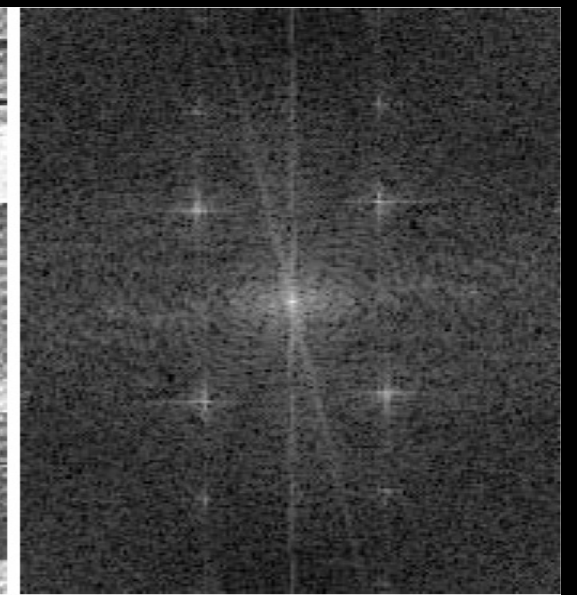
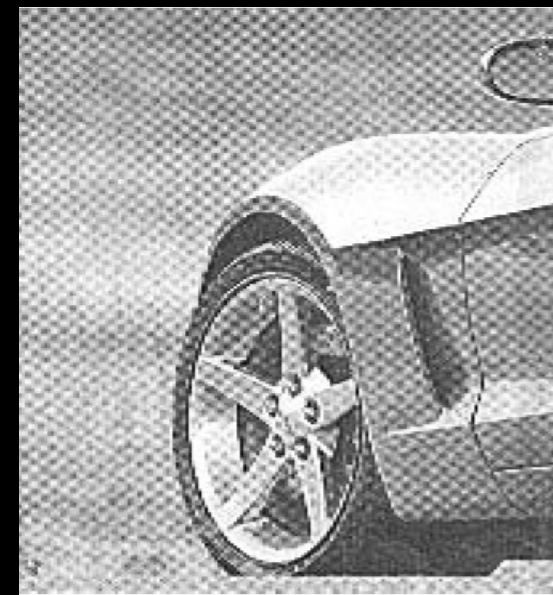
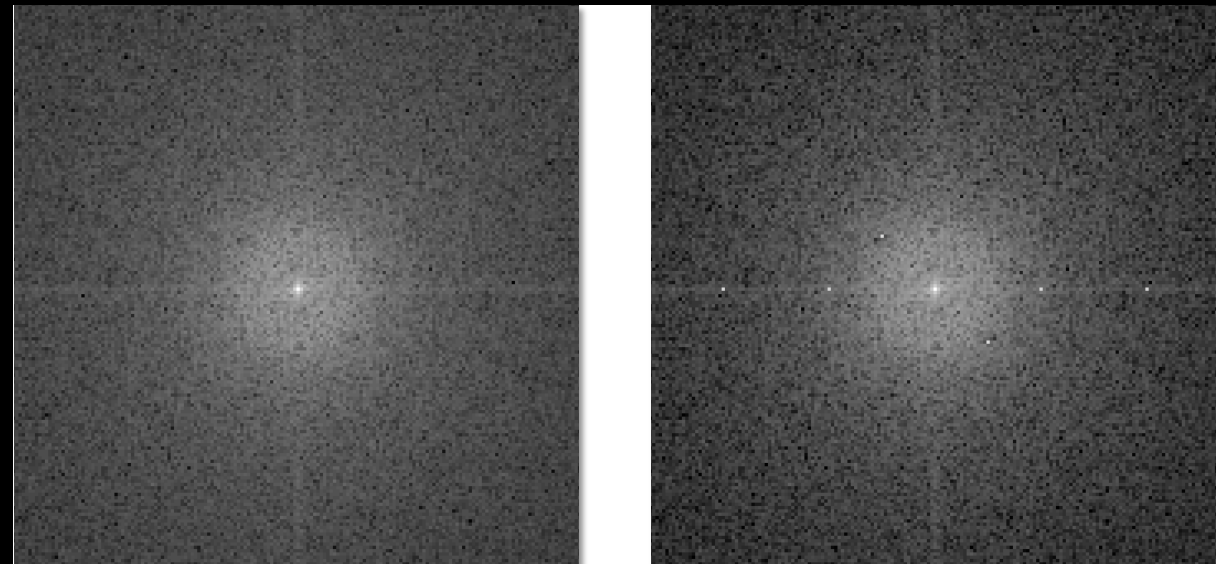
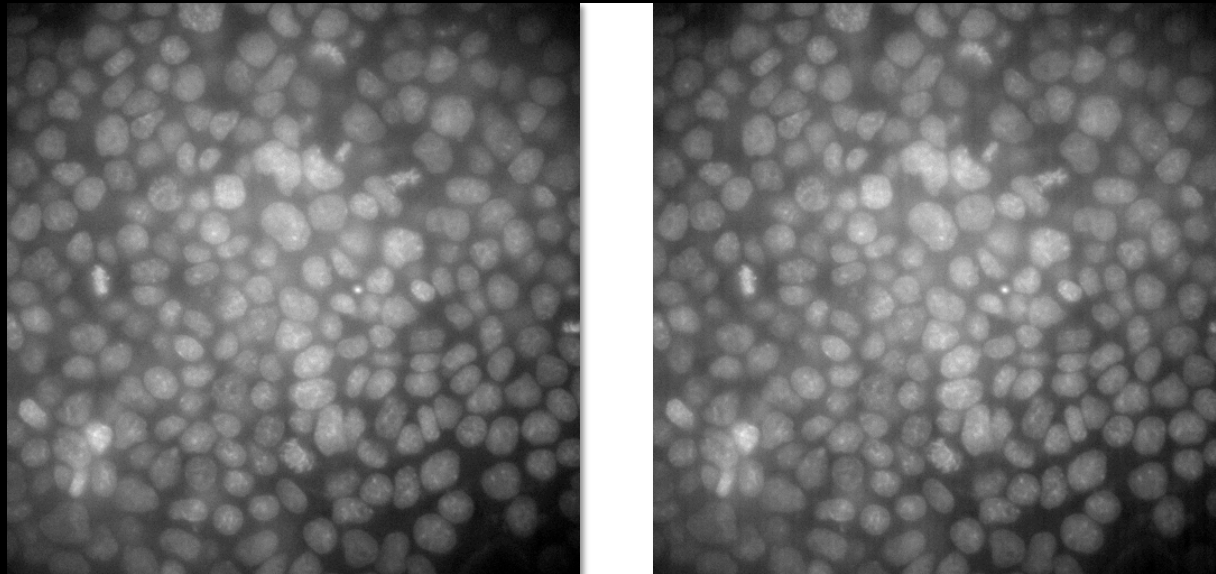
Median



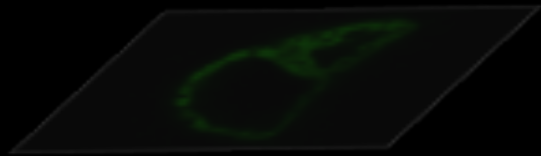
Minimum

Image transforms

Fourier transform



Outline



Digital image formation

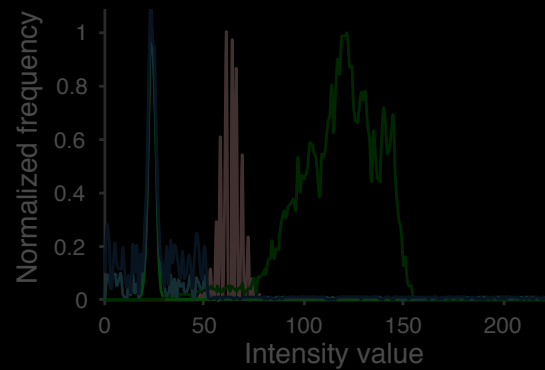


Image properties

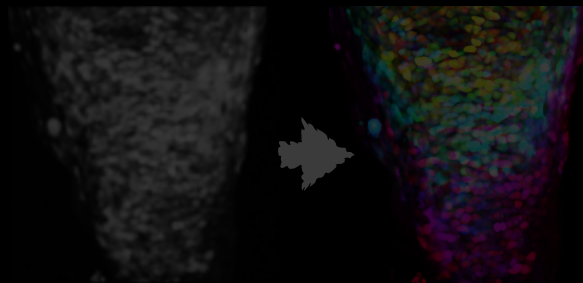


Image processing

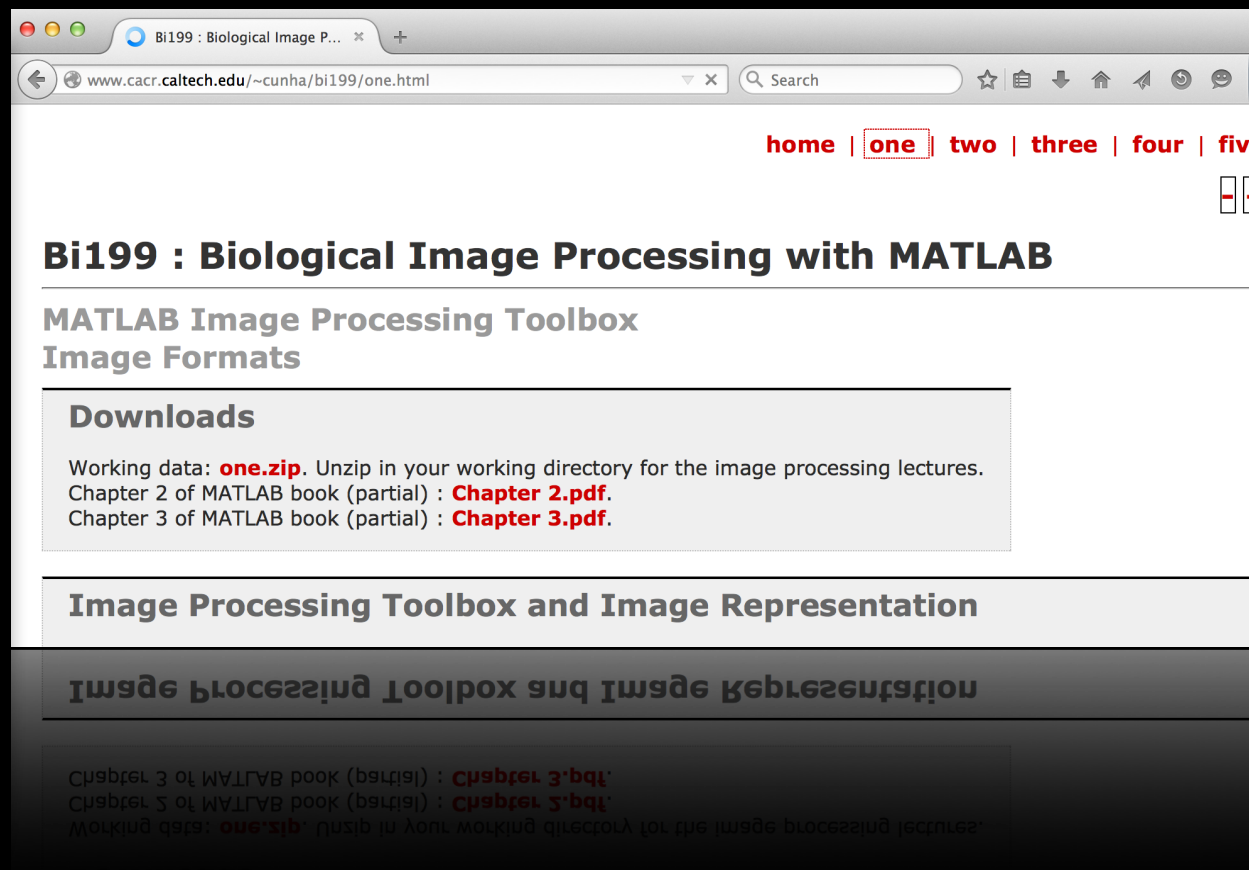


Available softwares

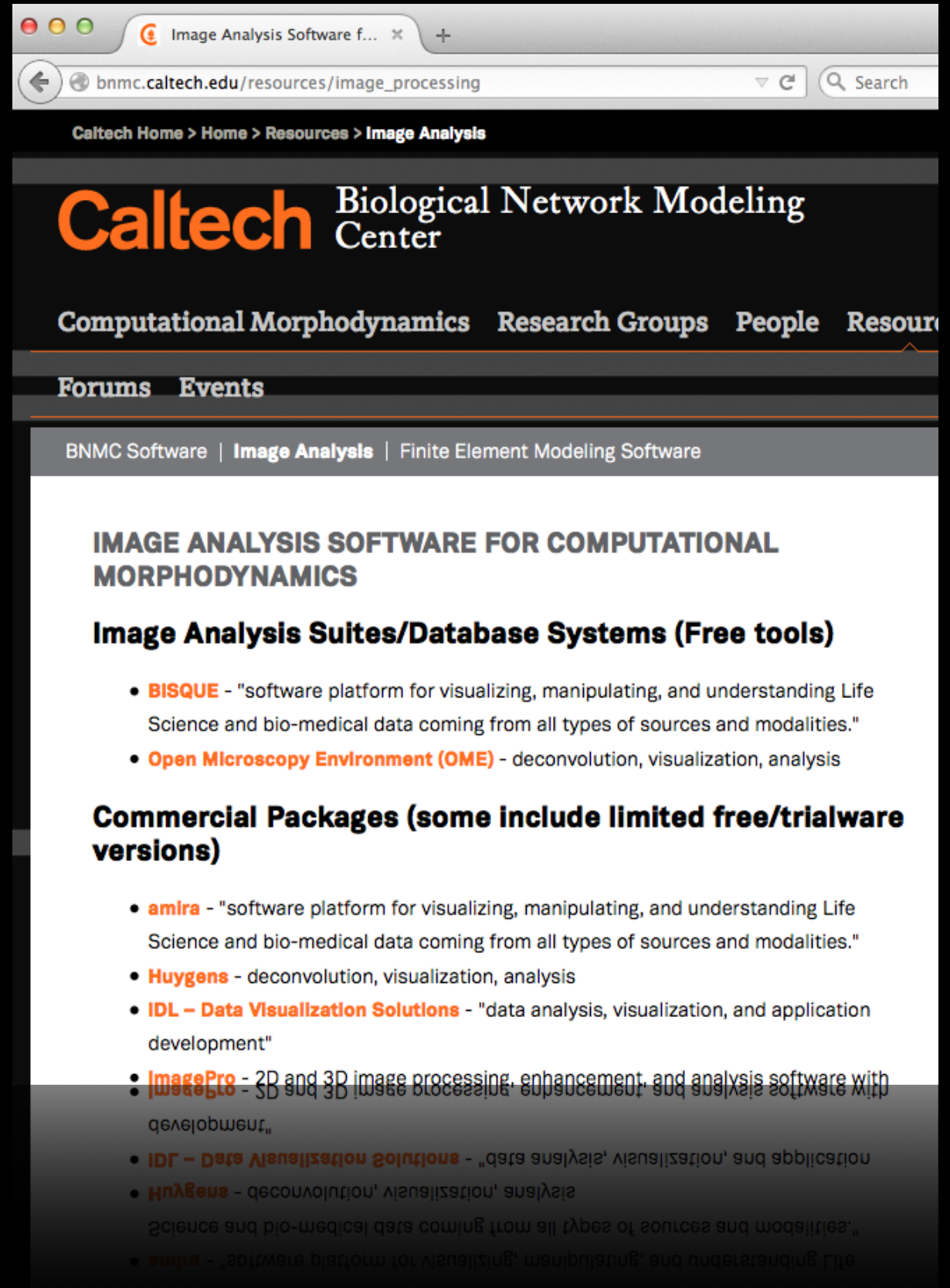
Resources @ Caltech

Bi 270. Special Topics in Biology. Units to be arranged; first, second, third terms. Students may register with permission of the responsible faculty member.

Bi 270/BioE 240 course offered by Dr. Alphan Altinok.



A screenshot of a web browser showing the Bi199 course website. The browser's address bar displays 'www.cacr.caltech.edu/~cunha/bi199/one.html'. The website has a navigation bar with links: 'home', 'one' (highlighted), 'two', 'three', 'four', and 'five'. The main heading is 'Bi199 : Biological Image Processing with MATLAB'. Below this is a section titled 'MATLAB Image Processing Toolbox Image Formats'. A 'Downloads' box contains the text: 'Working data: **one.zip**. Unzip in your working directory for the image processing lectures. Chapter 2 of MATLAB book (partial) : **Chapter 2.pdf**. Chapter 3 of MATLAB book (partial) : **Chapter 3.pdf**.' Below the downloads box is a section titled 'Image Processing Toolbox and Image Representation'.



A screenshot of a web browser showing the Caltech Biological Network Modeling Center website. The browser's address bar displays 'bnmc.caltech.edu/resources/image_processing'. The website has a navigation bar with links: 'Caltech Home > Home > Resources > Image Analysis'. The main heading is 'Caltech Biological Network Modeling Center'. Below this is a navigation bar with links: 'Computational Morphodynamics', 'Research Groups', 'People', 'Resources', 'Forums', and 'Events'. The main content area is titled 'BNMC Software | Image Analysis | Finite Element Modeling Software'. It contains two sections: 'IMAGE ANALYSIS SOFTWARE FOR COMPUTATIONAL MORPHODYNAMICS' and 'Image Analysis Suites/Database Systems (Free tools)'. The 'Free tools' section lists: 'BISQUE' - 'software platform for visualizing, manipulating, and understanding Life Science and bio-medical data coming from all types of sources and modalities.' and 'Open Microscopy Environment (OME)' - 'deconvolution, visualization, analysis'. Below this is a section titled 'Commercial Packages (some include limited free/trialware versions)' which lists: 'amlra' - 'software platform for visualizing, manipulating, and understanding Life Science and bio-medical data coming from all types of sources and modalities.', 'Huygens' - 'deconvolution, visualization, analysis', 'IDL - Data Visualization Solutions' - 'data analysis, visualization, and application development', and 'ImagePro' - '2D and 3D image processing, enhancement, and analysis software with development'.

Image formats

raster : storing as a grid

vector : storing shapes

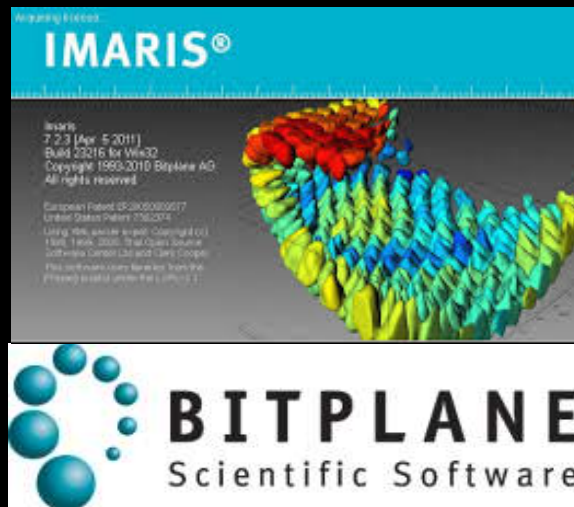
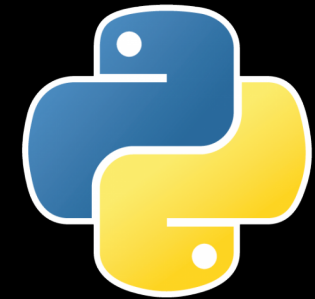
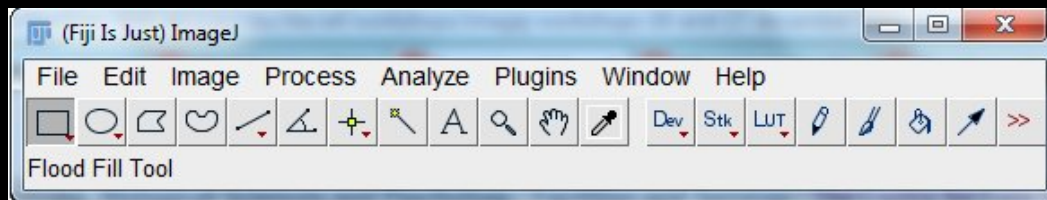
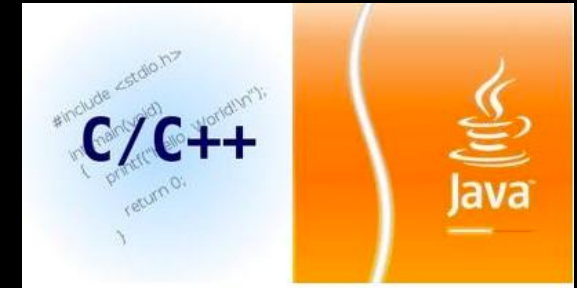
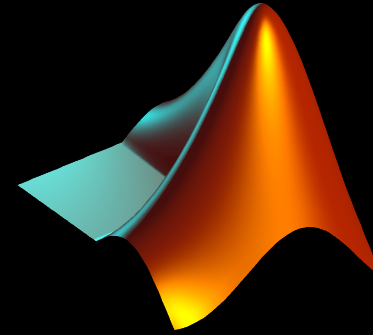
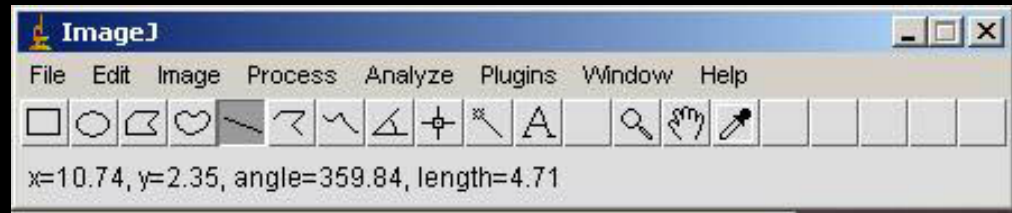
compression/no compression

reducing irrelevant/redundant information

TIFF	compression metadata annotation multiple frames
PNG	alpha channel compression
JPEG	compression RGB storage

LSM, LEI, OIB encapsulated TIFF images
many others : BMP, GIF, RAW, XWD, ...

Available softwares



Amira	Bio-Formats	Bitplane Imaris	CellProfiler
Comstat2	Endrov	Farsight	Fiji
FocalPoint	i3dcore	IDL	ImageJ
ImageMagick	Imago	ImagePro	Imglib
lqm	ITK-VTK	Macnification	Metamorph
MIPAV	PIL	Qu-Matlab	V3D
VisAD	VisBio	XuvTools	Bisque
Omero	Morphographix	OpenCV	

List compiled by Dr. Alphan Altinok

Questions